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Halo acids and DMSO: Electrophilic aromatic substitution at room temperature conditions

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HALO ACIDS AND DMSO: ELECTROPHILIC AROMATIC SUBSTITUTION AT ROOM TEMPERATURE CONDITIONS

PANKAJ KULSHRESTHA

AUGUST, 2000

THESIS

SUBMITTED IN PARTIAL FULFILLMENT OF THE REQUIREMENTS
FOR THE DEGREE OF MASTER OF SCIENCE

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AT ROOM TEMPERATURE CONDITIONS

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Lastly I would like to take this opportunity to thank almighty God, my parents, all my professors and my friends for their moral support and being there to help me whenever I needed their help.

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ABSTRACT

HALO ACIDS AND DMSO: ELECTROPHILIC AROMATIC SUBSTITUTION AT ROOM TEMPERATURE CONDITIONS

By Pankaj Kulshrestha under the guidance of Dr. James J. Worman

A simple, safe and cost effective experiment to demonstrate electrophilic aromatic substitution in the undergraduate laboratory can be accomplished by placing a small amount of N,N Dimethyl Aniline or a few crystals of Phenol in an NMR tube containing an appropriate level of deuterio DMSO. After observation of the expanded proton NMR spectrum of the aromatic region, a few drops of concentrated HBr and HCl aqueous acids are added and the NMR spectrum recorded after one hour.

Observation of an A,A' B,B' typical four line pattern in the aromatic region of NMR spectrum is direct evidence for the halogenation on the aromatic ring at para position. Addition of gaseous hydrogen chloride provides a faster and cleaner reaction. A mechanism for the reaction, GC/MS data to support the NMR results, along with other examples are presented.

Application of the reaction for halogenation of complex biomolecules in an attempt to enhance their activity is plausible because room temperature experimental conditions should prevent structural degradation of sensitive molecules and produce fewer non-toxic byproducts.

INTRODUCTION

The objective of my research was to synthesize halogenated biomolecules in an attempt to enhance the activity against antibiotic resistant strains of bacteria and to reduce toxic byproducts of halogenation reactions in an undergraduate laboratory, thereby minimizing environmental pollution.

Current methods of aromatic halogenation involve the use of reagents and catalysts under experimental conditions which present a safety hazard when performed in an undergraduate teaching laboratory. In addition, the disposal of wastes are costly and definitely not benign to the environment.

Existing methods of halogenation involve the use of liquid bromine, chlorine gas and other hazardous halogenating reagents in the presence of catalysts (1,2). The methods employ drastic conditions which would destroy the structure of the biomolecules and perhaps decrease the pharmacological activity of the biomolecules, even causing them to be acutely toxic. The reaction conditions presently employed may also produce byproducts, many of which are hazardous to the environment (3,4).

Experimental conditions which on one hand involve halogenation of biomolecules safely with the formation of fewer toxic byproducts and on the other hand stereospecifically and

stereoselectively allow for the synthesis of biomolecules targeted to resistant microbial strains, would minimize environmental pollution and add knowledge to the field of medicinal chemistry.

Biomolecular target specificity can reduce the toxic ill effects and side effects on human or animal species and also prevent environmental pollution, which in turn may provide a safe living environment for other organisms in air, soil and water.

Safe disposal of the toxic byproducts of chemical reactions, in particular reactions involving halogenation, have been a matter of deep concern because byproducts have the potential to act as environmental pollutants. A part of this research focuses on the issue of trying to design safe and environmentally benign synthetic reactions for the research labs and for industry.

The design of such reactions promises the generation of stereoselective and stereospecific products and the safe disposal of unwanted byproducts, primarily by the use of haloacids in dimethyl sulfoxide under room temperature conditions. This same experimental design can be used in undergraduate laboratories to demonstrate halogenation reactions which can be accomplished with minimum waste and improved safety, thereby decreasing the risk of severe laboratory accidents.

EARLY INVESTIGATIONS

Before and during the 1960's and the 1970's electrophilic aromatic halogenation using liquid bromine/ferric bromide and chlorine gas/ferric chloride or other hazardous halogenating reagents and catalysts was not only studied in theory (1,2,5,6) but also included in laboratory coursework (7).

This important chemical reaction and its mechanism is taught in undergraduate coursework (8,9,10,11,12) and graduate coursework (13,14), but due to the use of expensive and hazardous reagents and unsafe experimental conditions, these experiments were removed from the undergraduate laboratory experimental curriculum (15,16).

As early as 1970's (17,18) during the ongoing research (scheme 1 below) by Dr. James J.Worman, it was found that the dihydrobromide salt of 3,7 dimethoxy-1,5 diphenyloctahydro-1,5 diazocine, underwent halogenation in the presence of dimethyl sulfoxide in the NMR tube when kept for a few hours.

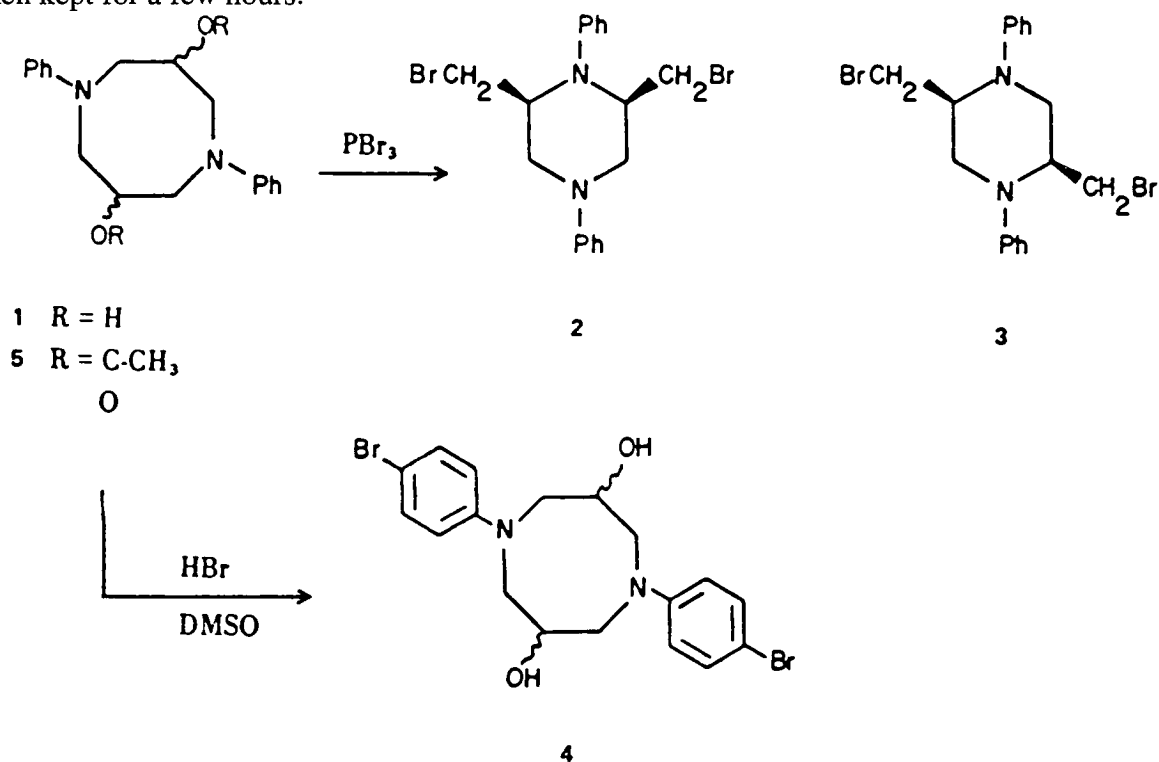


Figure 1. below shows a typical four line pattern of the aromatic region of a para substituted aromatic system (1,5-Bis-(p-bromo phenyl)octahydro-3,7- dimethoxy-1,5-diazocine) – the compound which was formed during the ongoing researchwork done by Dr.Worman's research group, when dimethyl sulfoxide was added to the dihydrobromide salt of octahydro 3,7-dimethoxy- 1,5- diphenyl- 1,5- diazocine in an NMR tube.

Figure 1.

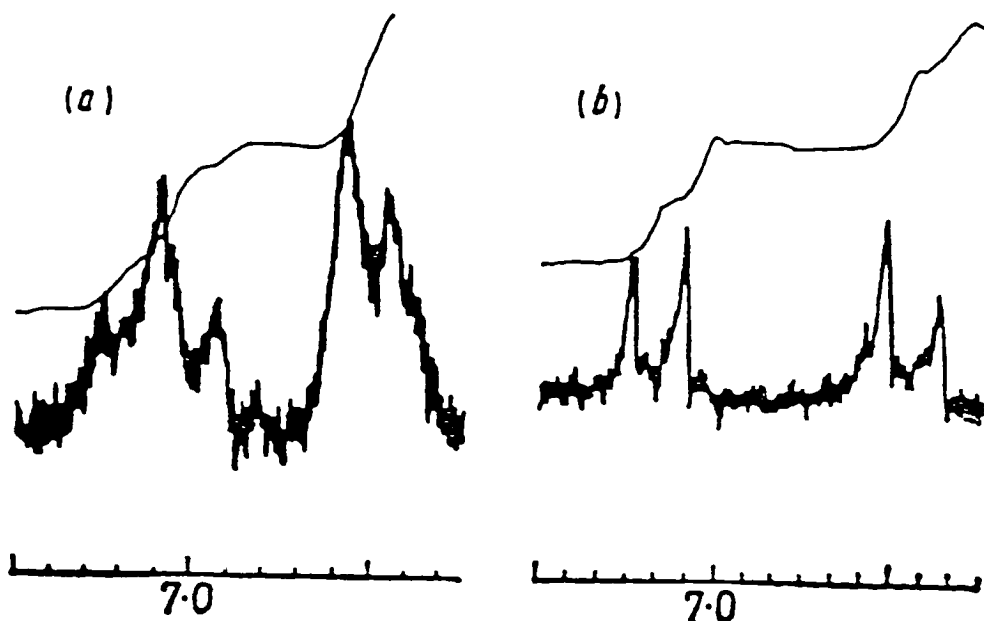
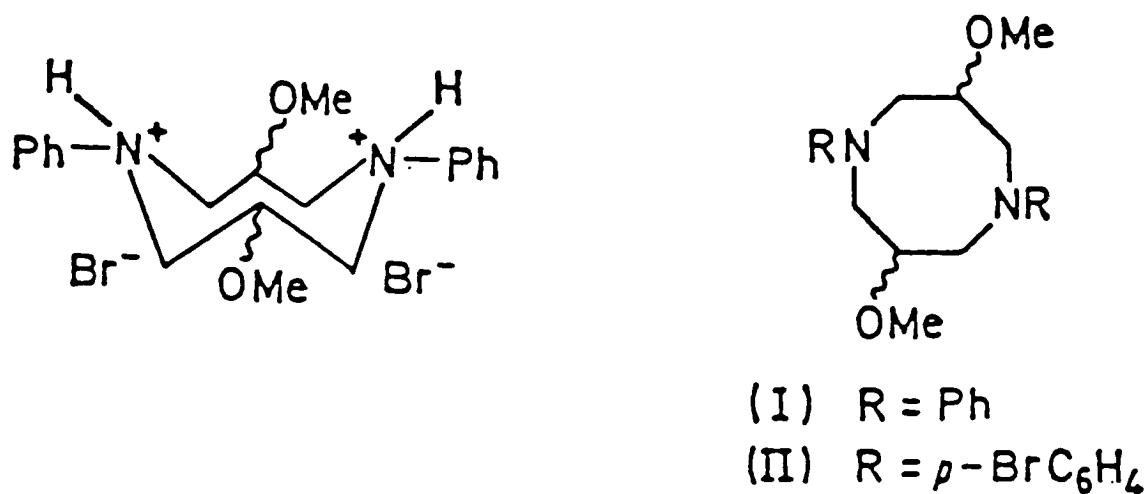
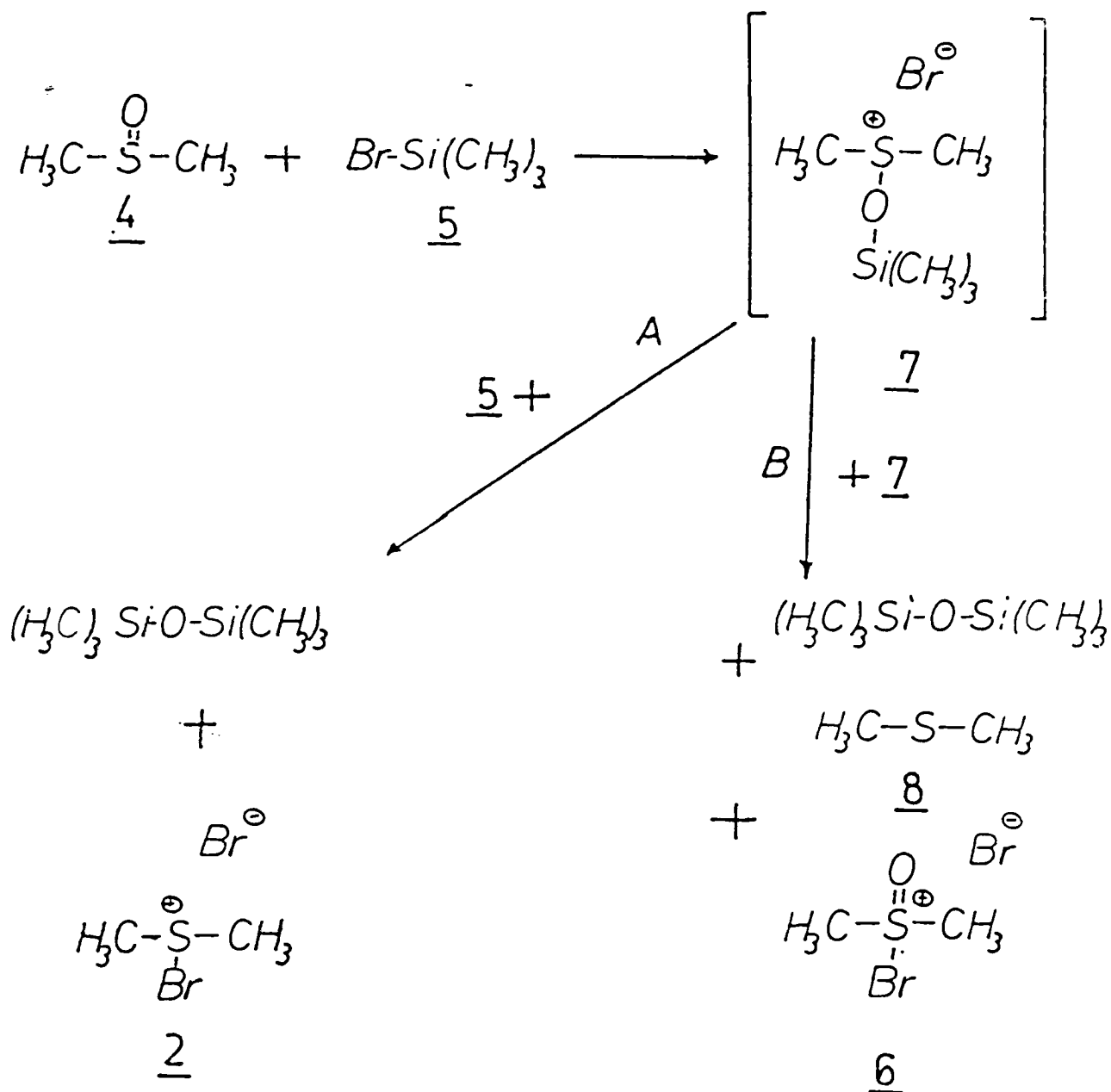
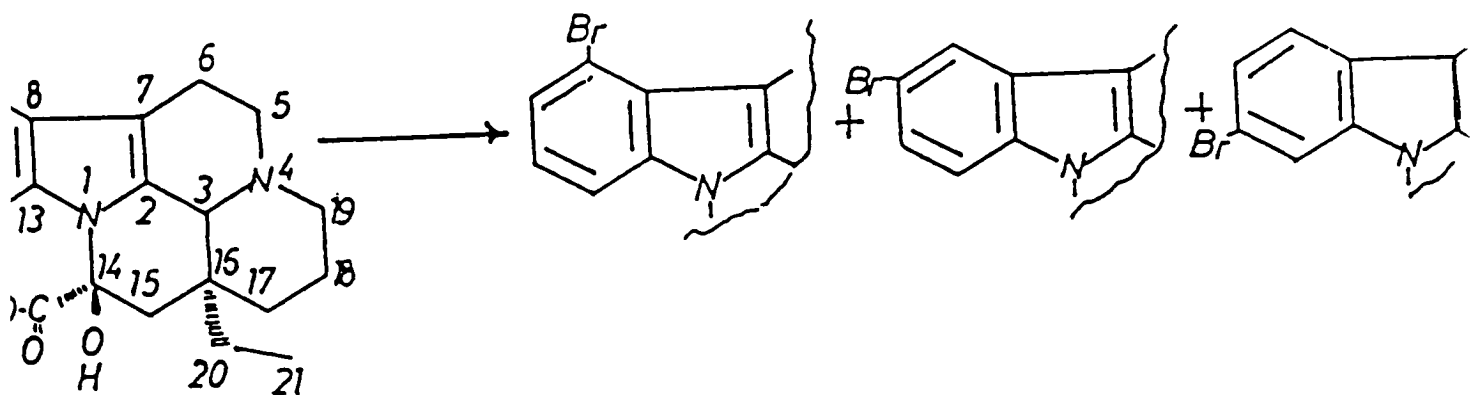
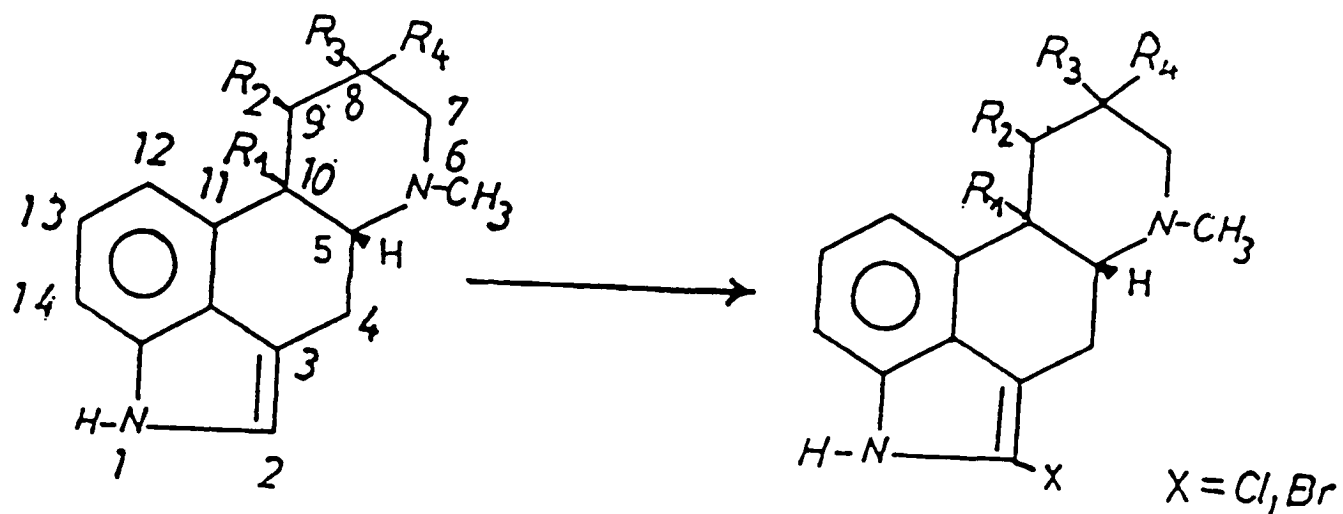


FIGURE 1 N.m.r. spectra of the dihydrobromide of octahydro-3,7-dimethoxy-1,5-diphenyl-1,5-diazocine (I) in dimethyl sulphoxide at room temperature; (a) aromatic-proton absorption immediately after dissolution of dihydrobromide in solvent; (b) same absorption after 8 h

Very little work was done in this area until 1989 when Dr. Megyeri Gabor and Dr. Tibor Keve of Hungary halogenated indole alkaloids using halodimethylsulfonium halogenids (compound no.2 in scheme 2 below) and halodimethylsulfoxonium halogenids (compound no.6 in scheme 2 below)(19).



These are only a few examples out of many reactions carried out by Dr.Gabor's research group which show that halogenation can take place not only at the vinyl position but also on the aromatic ring using previously stated conditions (scheme 3 below).



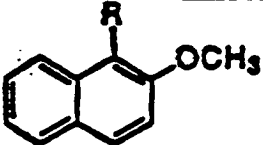
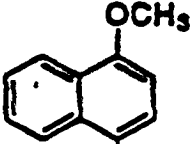
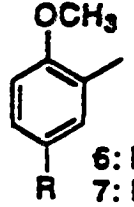
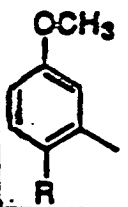
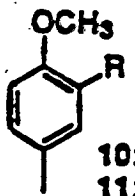
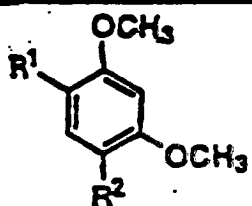
In 1997, pioneer work was done by Dr. George Majetich's research group when they presented a myriad of reaction conditions (table 1, 2 and 3 below) and procedures to specifically brominate various aromatic and heterocyclic compounds (20).

It is a distinct possibility that these conditions can be used to brominate aromatic macromolecules to give useful chemical compounds and produce few non-toxic byproducts, the reaction conditions are easy to follow and have the potential for reaction specificity, which could form targeted biomolecules. The utility of such halogenation conditions and reaction methodologies, which were not attempted earlier, were investigated.

Table 1

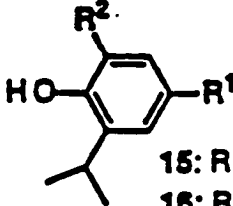
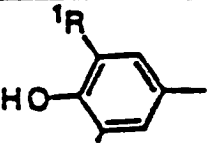
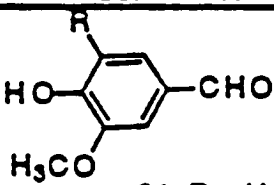
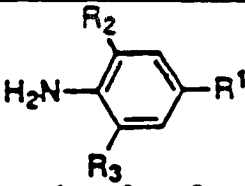
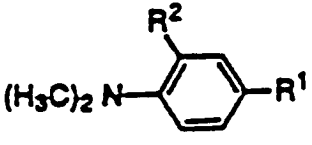
solvent	reaction condns	yield of 3 (%)	reaction completion
DMSO	2 h at rt	96	complete
acetonitrile	32 h at rt	84	trace
THF	1 h at reflux	trace	unreacted 2 only
THF	24 h at reflux	99	unreacted 2 complete
THF/AcOH (3:1)	2 h at rt	50	~50% complete
AcOH/DMSO (2:1)	<5 min	96	complete

Table 2

Substrate / Product	Percent Yield and Reaction Time Using Standard Conditions			
	HBr, DMSO "A"	HBr, AcOH, DMSO "B"	Br ₂ , CCl ₄ "C"	Br ₂ , AcOH "D"
 2: R = H 3: R = Br	96% of 3 after 2-h	96% of 3 after 5-min	90% of 3 after 5-min	89% of 3 after 5-min
 4: R = H 5: R = Br	77% of 5 after 72-h	97% of 5 after 24-h	90% of 5 after 72-h	85% ^a of 5 after 72-h
 6: R = H 7: R = Br	82% of 7 after 12-h	86% of 7 after 12-h	75% of 7 after 5-min	75% of 7 after 2-h
 8: R = H 9: R = Br	91% of 9 after 12-h	90% of 9 after 12-h	68% ^a of 9 after 20-min	89% ^a of 9 after 20-min
 10: R = H 11: R = Br	77% of 11 after 72-h	75% of 11 after 72-h	74% of 11 after 20-min	91% of 11 after 5-min
 12: R ¹ = R ² = H 13: R ¹ = H; R ² = Br 14: R ¹ = R ² = Br	34% of 13 after 5-min <hr/> 89% of 14 after 20-min	73% of 14 after 5-min <hr/> 87% of 14 after 20-min	80% ^a of 14 after 5-min	77% of 14 after 5-min

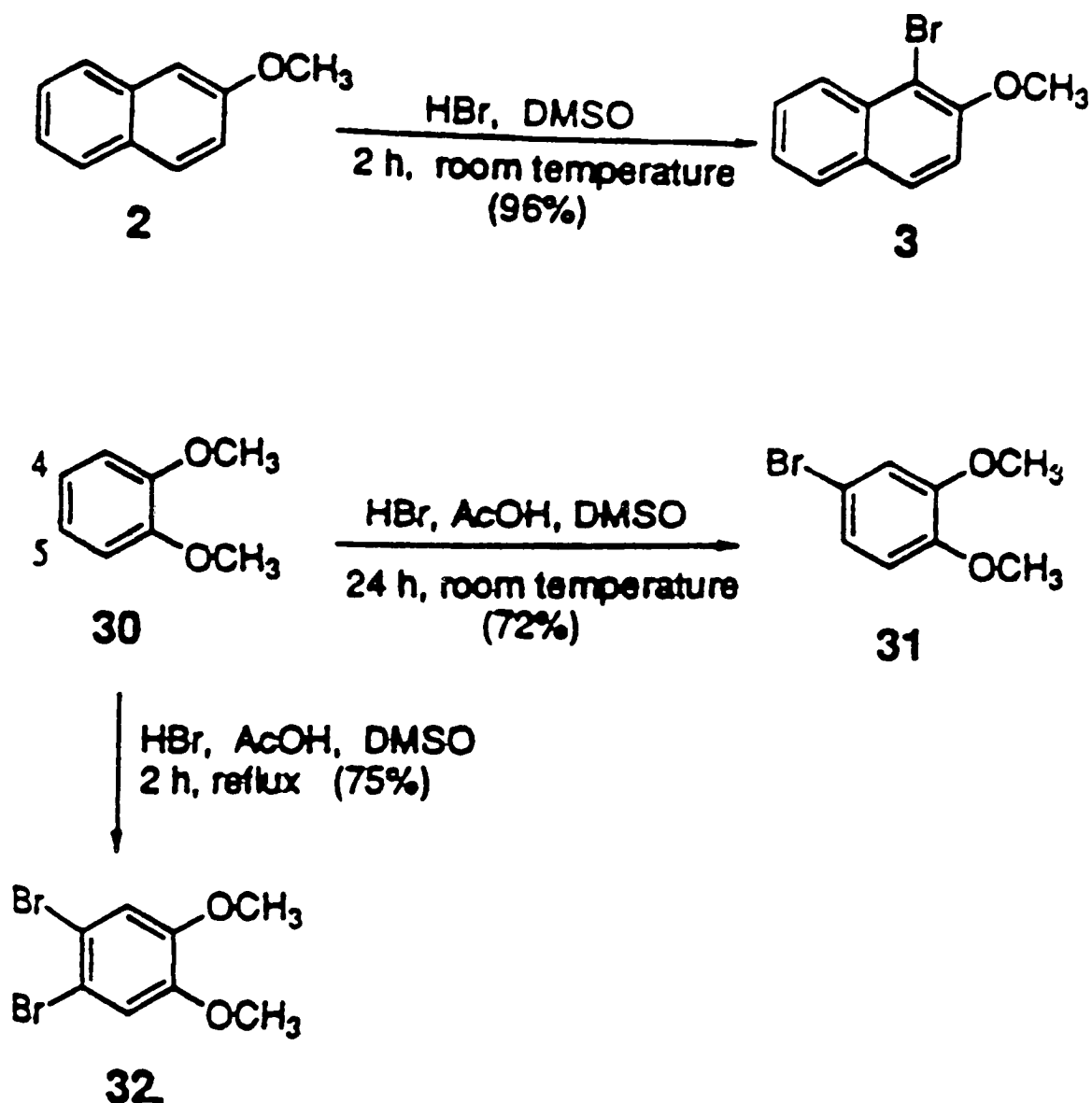
Product contained inseparable impurities. The yield shown is based on analysis of the ¹H NMR spectrum.

Table 3

Substrate / Product	Percent Yield and Reaction Time Using Standard Conditions			
	HBr, DMSO "A"	HBr, AcOH, DMSO "B"	Br ₂ , CCl ₄ "C"	Br ₂ , AcOH "D"
 <p>15: R¹ = R² = H 16: R¹ = Br; R² = H 17: R¹ = R² = Br</p>	90% of 16 after 24-h	91% of 16 after 5-min	23% of 16 and 64% of 17 after 5-min	63% of 16 and 31% of 17 after 5-min
 <p>18: R¹ = R² = H 19: R¹ = Br; R² = H 20: R¹ = R² = Br</p>	88% of 19 after 48-h	87% of 19 after 2-h	87% of 19 after 30-min	83% of 19 after 30-min; trace 20
 <p>21: R = H 22: R = Br</p>	72% of 22 after 12-h	57% of 22 after 12-h	48% of 22 after 24-h	80% of 22 after 20-min
 <p>23: R¹ = R² = R³ = H 24: R¹ = Br; R² = R³ = H 25: R¹ = R² = Br; R³ = H 26: R¹ = R² = R³ = Br</p>	76% of 24 after 6-h	74% of 24 after 24-h	19% of 24; 33% of 25 and 30% of 26 after 5-min	21% of 24; 41% of 25 and 31% of 26 after 5-min
 <p>27: R¹ = R² = H 28: R¹ = Br; R² = H 29: R¹ = R² = Br</p>	43% of 28 after 24-h	42% of 28 after 24-h	60% of 28 after 5-min	68% of 28 and 29% of 29 after 5-min

^a Inseparable byproducts were observed. The yield shown is based on analysis of the ¹H NMR data.

Scheme 4 and 5 show a few examples of many reactions carried out by Dr. Majetich's research group. Scheme 5 shows that under room temperature halogenation conditions one position on the aromatic ring of Veratrole (compound 30 in **scheme 5**) is halogenated and under severe reflux halogenation conditions more than one position on the aromatic ring of the same compound is halogenated.



EXPERIMENTAL WORK

The following experimental work was done in the Department of Chemistry at the Rochester Institute of Technology, Rochester, NY under the supervision of Dr. James J. Worman.

All nuclear magnetic resonance data were obtained on a 7 Tesla Bruker DRX – 300 300 MHz spectrometer. Deuterated dimethyl sulfoxide was used as the solvent for all samples.

All GC/MS data were obtained on a Hewlett Packard model HP190915S-936. The column used for the entire analysis was a capillary column (60.0mts * 250.0m * 0.25m in dimensions) tightly packed with HP-1MS-crosslinked Methyl Siloxane. Conditions employed were kept uniform during the entire analysis of samples. Uniform conditions used were constant flow of gas at the rate of 1ml/min and total flow being 33.4 ml/min under a total pressure of 35.23 psi, the temperature of the heater/injector was set at 250°C. The column was operated at a temperature range of 50°C-280°C. The temperature of the column was held at 50°C for one minute, then ramped at 20°C/min till 180°C then reached a final temperature of 280°C to which it was set at the rate 30°C/min and held for 2 minutes. Dimethyl Sulfoxide, Tetrahydrofuran and Methylene Chloride were used as diluting solvents during the GC/MS run of various samples.

Typical NMR and GC/MS spectra are shown in Appendix A and B respectively.

THREE PHASES OF MY EXPERIMENTAL WORK INCLUDE

PHASE I :- involves the use of NMR to determine the formation of para substituted halogenated compounds of simple aromatic systems.

PHASE II :- involves the use of GC/MS analysis to verify the NMR results and to determine the isomeric distribution of halogenated species formed.

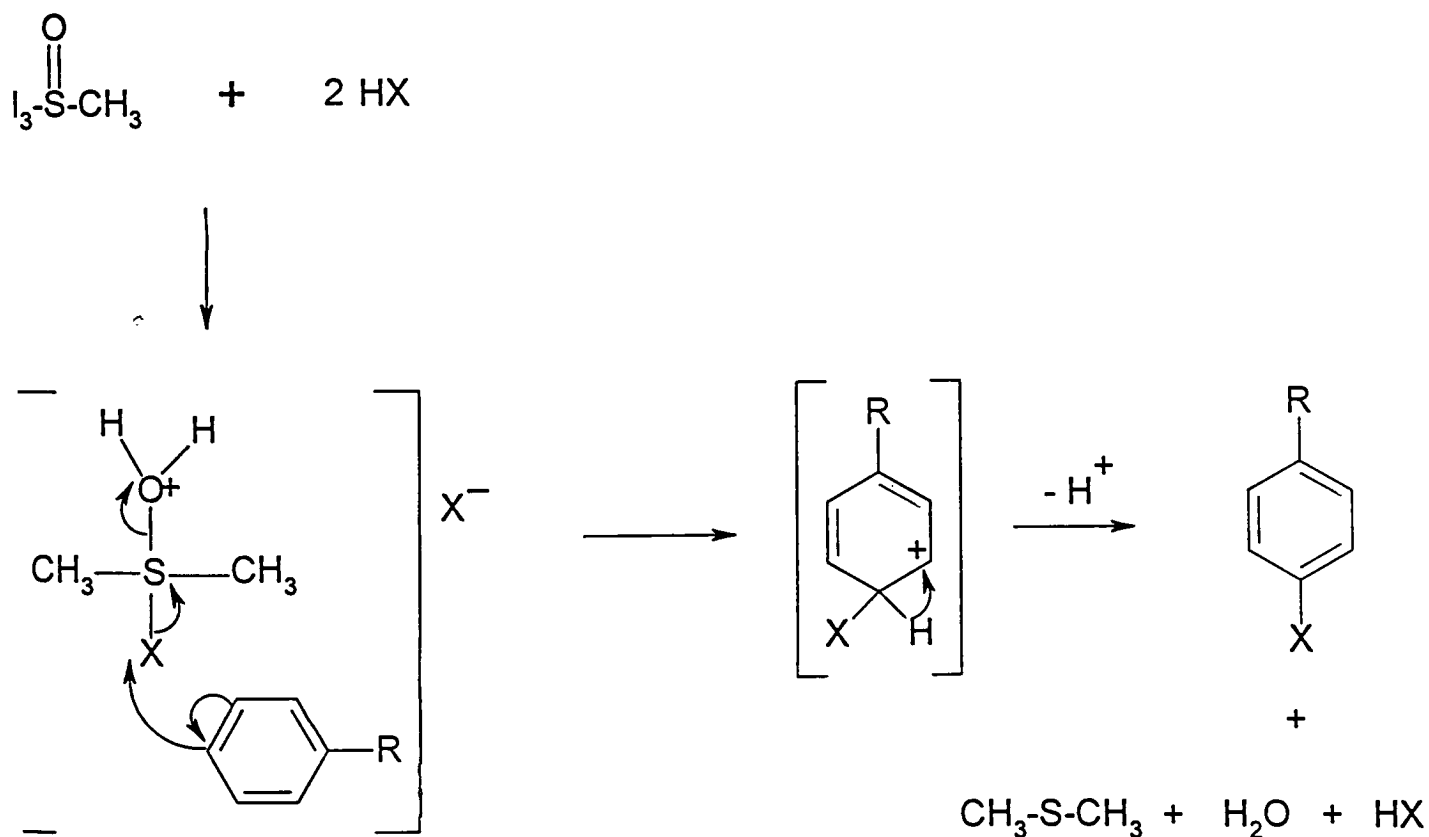
PHASE III :- involves the use of NMR and GC/MS analysis to determine the position of halogen substitution on the macro/biomolecules.

TWO POINT APPROACH UTILIZED IN THE RESEARCH

- 1) Use of room temperature conditions for halogenation.
- 2) Ensure safe halogenation of aromatic systems by utilizing novel but simple laboratory techniques.

REACTION MECHANISM

Two moles of a halo acid react with one mole of dimethyl sulfoxide. Halogen anion (X^-) of one mole of the halo acid attacks the sulfur of dimethyl sulfoxide and hydrogen cation H^+ of the same halo acid attacks the oxygen of the dimethyl sulfoxide giving rise to a tetrahedral intermediate state. The hydrogen cation H^+ of a second mole of the haloacid also attacks the oxygen of the hydroxy group to give a positive oxonium ion balanced by the halogen anion(X^-). The halogen attached to the sulfur of the tetrahedral intermediate attacks the para position of the aromatic ring of an aromatic system with the simultaneous in-situ formation of the para substituted halogenated aromatic system, water and dimethyl sulfide.

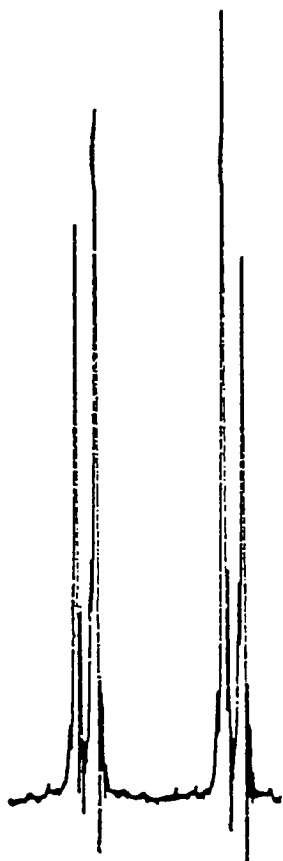


EXPERIMENTAL PROCEDURE

Aqueous Experimental Conditions:- To two drops of N,N-Dimethylaniline in an NMR tube was added 1 ml of deuterated dimethyl sulfoxide followed by the addition of 2-4 drops of concentrated haloacid and the solution was left at room temperature for one to two hours. Thereafter, the NMR and GC/MS analysis was performed to determine the in-situ formation of para substituted halogenated aromatic species and other isomers (Typical NMR results are shown in **figure 2.** on next page.)

Gaseous Experimental Conditions:- To two drops Anisole in an NMR tube was added 1 ml of deuterio DMSO followed by passing gaseous HCl at a controlled rate of one gas bubble/second for 30 seconds and then the solution was heated in an oil bath at the temperature of 100°C for one hour. The solution was left to stand at room temperature for one hour, afterwhich NMR and GC/MS analysis was performed to observe the in-situ formation of para substituted halogenated aromatic species (Typical NMR results are shown in **figure 2.** on next page.)

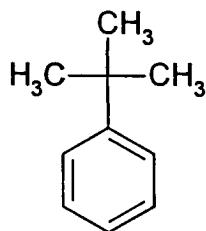
Figure 2. TYPICAL FOUR LINE PATTERN OF A PARA SUBSTITUTED AROMATIC SYSTEM WITH TWO DIFFERENT SUBSTITUENTS



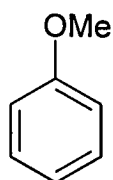
**TYPICAL FOUR LINE PATTERN OF A PARA
SUBSTITUTED AROMATIC SYSTEM WITH
TWO DIFFERENT SUBSTITUENTS**

EXPERIMENTAL PHASE I

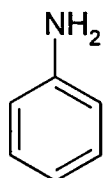
The following aromatic systems were halogenated in an effort to establish environmentally benign room temperature conditions for safe and simple halogenation experiments in an undergraduate laboratory. At the same time a search was made to develop the best halogenating reagents under room temperature conditions which could be used for halogenation of biomolecules.



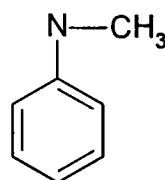
t-BUTYL BENZENE



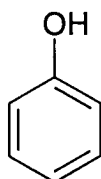
ANISOLE



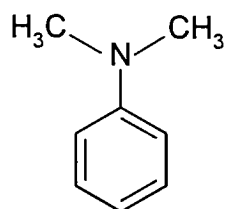
ANILINE



N METHYL ANILINE



PHENOL



N,N DIMETHYL ANILINE

Table 4 : LIST OF HALOGENATION REACTIONS AND NMR ANALYSIS RESULTS

LIST OF REACTIONS	NMR RESULTS
TERT.BUTYLBENZENE +CONC.HF +DMSO	- ve
TERT.BUTYLBENZENE +CONC.HCl +DMSO	- ve
TERT.BUTYLBENZENE +CONC.HBr +DMSO	- ve
TERT.BUTYLBENZENE +CONC.HI +DMSO	- ve
TERT.BUTYLBENZENE +GASEOUS HCl + DMSO	- ve
ANISOLE + CONC.HF + DMSO	- ve
ANISOLE + CONC.HCl + DMSO	- ve
ANISOLE + CONC.HBr + DMSO	- ve
ANISOLE + CONC.HI + DMSO	- ve
ANISOLE + GASEOUS HCl + DMSO (HEATED IN OIL BATH FOR 1 Hr.)	+ ve
ANILINE + CONC.HF + DMSO	- ve
ANILINE + CONC.HCl + DMSO	+ ve
ANILINE + CONC.HBr + DMSO	+ ve
ANILINE + CONC.HI + DMSO	- ve
ANILINE + GASEOUS HCl + DMSO	+ ve
N-METHYLANILINE + CONC. HF + DMSO	ve
N-METHYLANILINE + CONC.HCl + DMSO	+ ve
N-METHYLANILINE + CONC.HBr. + DMSO	+ ve
N-METHYLANILINE + CONC.HI + DMSO	- ve
N-METHYLANILINE + GASEOUS HCl + DMSO	+ ve
N,N-DIMETHYLANILINE + CONC. HF + DMSO	- ve
N,N-DIMETHYLANILINE + CONC. HCl + DMSO	+ ve
N,N-DIMETHYLANILINE + CONC. HBr + DMSO	+ ve

N,N-DIMETHYLANILINE + CONC. HI + DMSO	- ve
N,N-DIMETHYLANILINE + GASEOUS HCl + DMSO	+ ve
PHENOL + CONC.HF + DMSO	ve
PHENOL + CONC.HCl + DMSO	+ ve
PHENOL + CONC.HBr + DMSO	+ ve
PHENOL + CONC.HI + DMSO	- ve
PHENOL + GASEOUS HCl + DMSO	+ ve

+ ve means that typical four line pattern of a para substituted aromatic system was seen during NMR analysis

- ve means that typical four line pattern of a para substituted aromatic system was not seen during NMR analysis

EXPERIMENTAL PHASE II

Table 5 : NMR RESULTS(PARA, META, ORTHO SUBSTITUED HALOGENATED AROMATIC SYSTEMS)SUPPORTED BY GC/MS ANALYSIS

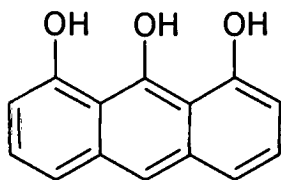
S.N.	LIST OF REACTIONS	PARA PRODUCT	META PRODUCT	ORTHO PRODUCT	% FORMED OUT OF ALL THE COMPOUNDS ELUTED
1.	TERT.BUTYL - BENZENE + CONC. HF + DMSO	- ve	-ve	-ve	-ve
2.	TERT.BUTYL - BENZENE + CONC. HCl + DMSO	- ve	- ve	- ve	- ve
3.	TERT.BUTYL - BENZENE + CONC. HBr + DMSO	- ve	- ve	- ve	- ve
4.	TERT.BUTYL - BENZENE + CONC. HI + DMSO	- ve	- ve	- ve	- ve
5.	TERT.BUTYL - BENZENE + GAS. HCl + DMSO	- ve	- ve	- ve	- ve
6.	ANISOLE + CONC. HF + DMSO	- ve	- ve	- ve	- ve
7.	ANISOLE + CONC. HCl + DMSO	- ve	- ve	- ve	- ve
8.	ANISOLE + CONC. HBr + DMSO	- ve	- ve	- ve	- ve
9.	ANISOLE + CONC. HI + DMSO	- ve	- ve	- ve	- ve
10.	ANISOLE + GAS. HCl + DMSO (HEATED IN OIL BATH FOR 2 Hrs.)	+ ve	- ve	+ ve	29.15% p-Chloroanisole 8.77% o-Chloroanisole
11.	ANILINE + CONC. HF + DMSO	- ve	- ve	- ve	- ve
12.	ANILINE + CONC. HCl + DMSO	+ ve	- ve	+ ve	26.36% p-Chloroaniline 4.85% o-Chloroaniline 2.13% 2,4-Dichloroaniline
13.	ANILINE + CONC. HBr + DMSO	+ ve	+ ve	+ ve	20.46% p-Bromoaniline 4.10% m-Bromoaniline 2.01% 2,4-Dibromoaniline

14.	ANILINE + CONC. HI + DMSO	- ve	- ve	- ve	- ve
15.	ANILINE + GAS. HCl + DMSO	+ ve	- ve	- ve	22.56% p-Chloroaniline 5.06% o-Chloroaniline 3.21% 2,4-Dichloroaniline
16.	N-METHYL - ANILINE + CONC. HF + DMSO	- ve	- ve	-ve	-ve
17.	N-METHYL - ANILINE + CONC. HCl + DMSO	+ ve	+ ve	+ ve	38.75% p-Chloro-N-methyl Aniline 9.25% m-Chloro-N-methyl Aniline 0.46% o-Chloro-N-methyl Aniline 1.19% 2,4-Dichloro-N-methyl Aniline
18.	N-METHYL - ANILINE + CONC. HBr + DMSO	+ ve	- ve	- ve	58.57% p-Bromo-N-methyl Aniline 5.01% 2,4-Dibromo-N-methyl Aniline
19.	N-METHYL - ANILINE + CONC. HI + DMSO	- ve	- ve	- ve	- ve
20.	N-METHYL - ANILINE + GAS. HCl + DMSO	+ ve	+ ve	+ ve	35.05% p-Chloro-N-methyl Aniline 6.23% m-Chloro-N-methyl Aniline 1.08% o-Chloro-N-methyl Aniline 0.77% 2,4-Dichloro-N-methyl Aniline
21.	N,N-DIMETHYL - ANILINE + CONC. HF + DMSO	- ve	- ve	- ve	- ve
22.	N,N-DIMETHYL - ANILINE + CONC. HCl +DMSO	+ ve	- ve	- ve	15.03% p-Chloro-N,N-dimethyl Aniline
23.	N,N-DIMETHYL - ANILINE + CONC. HBr +DMSO	+ ve	ve	- ve	60.32% p-Bromo-N,N-dimethyl Aniline
24.	N,N-DIMETHYL - ANILINE + CONC. HI + DMSO	- ve	- ve	- ve	- ve

25.	N,N DIMETHYL - ANILINE + GAS. HCl + DMSO	+ ve	- ve	- ve	17.78% p-Chloro-N,N-dimethyl Aniline
26.	PHENOL + CONC. HF + DMSO	- ve	- ve	- ve	- ve
27.	PHENOL + CONC. HCl + DMSO	+ ve	- ve	- ve	4.57% p-Chloro phenol
28.	PHENOL + CONC. HBr + DMSO (2 DIFFERENT GC/MS RESULTS)	A) + ve B) + ve	+ ve + ve	- ve + ve	12.86% p- & m-Bromo phenol in equal quantities 15.39% p-Bromo phenol 11.37% m-Bromo phenol 7.01% o-Bromo phenol
29.	PHENOL + CONC. HI + DMSO	- ve	- ve	- ve	- ve
30.	PHENOL + GAS. HCl. + DMSO	+ ve	- ve	- ve	5.03% p-Chloro phenol

EXPERIMENTAL PHASE III

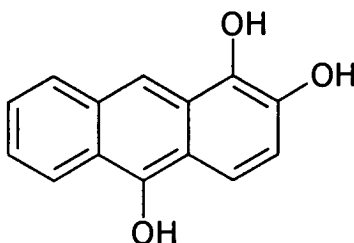
The following macro/biomolecules were selected for halogenation after thorough study of the Merck Index (21) and other organic medicinal chemistry books (22, 23, 24). They were halogenated with a possibility of enhancing their activity and forming a minimal quantity of toxic byproducts.



DITHRANOL

1,8,9 ANTHRACENETRIOL

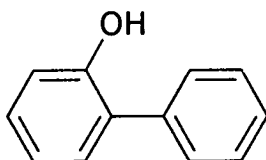
USED IN DERMATOLOGIC PREPARATIONS AND ANTIFUNGAL PREPARATIONS



ANTHRAROBIN

1,2,10 ANTHRACENETRIOL

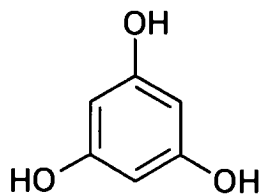
USED AS PARASITICIDE



O- PHENYL PHENOL (DOWICIDE I)

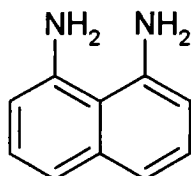
[1,1'-BIPHENYL] 2 - OL

USED IN GERMICIDE AND FUNGICIDE PREPARATIONS
AND IN COMMERCIAL RUBBER INDUSTRY



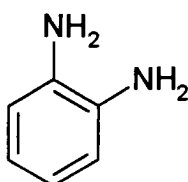
PHLOROGLUCINOL (1,3,5 BENZENETRIOL)

USED AS AN ANTISPASMODIC



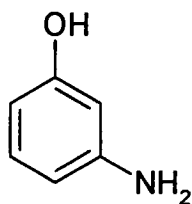
1,8 DIAMINO NAPHTHALENE

USED AS AN ANTIOXIDANT FOR LUBRICATING OILS AND FOR THE DETECTION OF
SELENIUM AND NITRITES



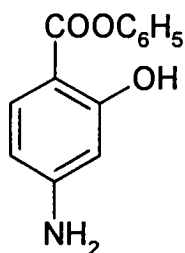
O- PHENYLENEDIAMINE(1,2- BENZENEDIAMINE)

USED IN THE MANUFACTURE OF DYES



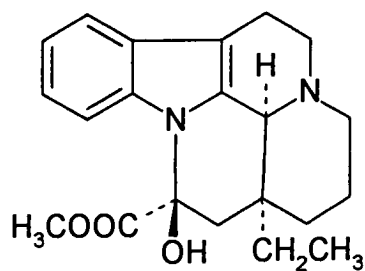
3 AMINO PHENOL

USED IN THE MANUFACTURE OF ANTITUBERCLAR AGENT P- AMINO SALICYLIC ACID(PAS)



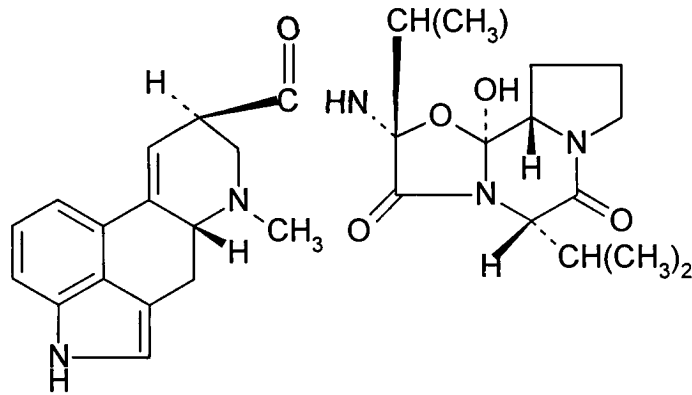
PHENYL AMINO SALICYLATE (PHENYL PAS)

USED AS AN ANTIBACTERIAL (TUBERCULOSTATIC)



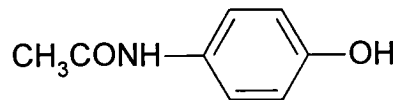
VINCAMINE

USED AS VASODILATOR



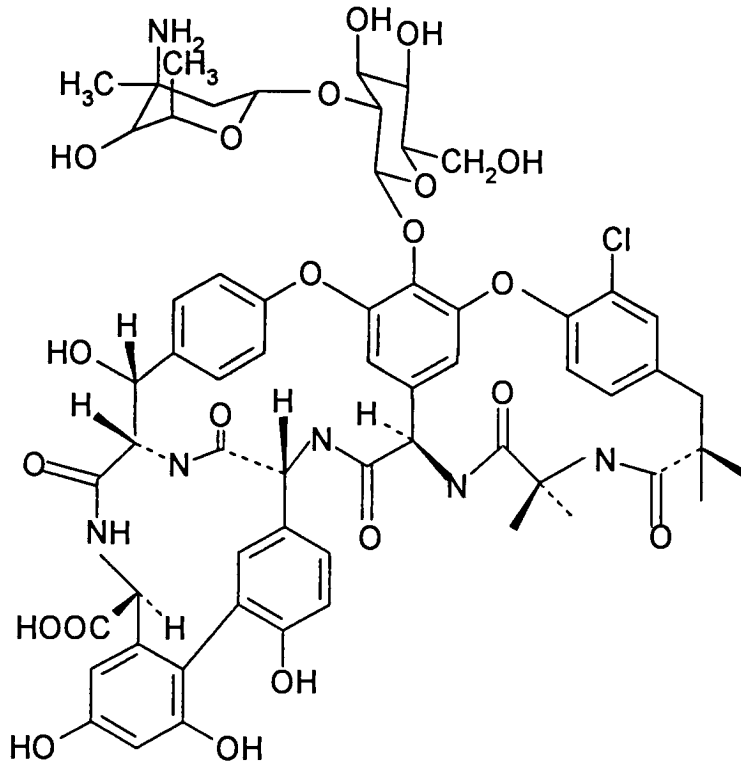
ERGOCORNINE

USED AS VASOCONSTRICTOR (SPECIFIC IN MIGRAINE).
IT HAS BEEN USED AS AN OXYTOCIC IN VETERINARY MEDICINE



4 ACETAMIDOPHENOL

USED AS AN ANTIPYRETIC AND ANALGESIC
IT HAS BEEN USED COMMERCIALY FOR THE MANUFACTURE OF AZO DYES
AND PHOTOGRAPHIC CHEMICALS



VANCOMYCIN (VANCOCIN)

ANTIBIOTIC SUBSTANCE PRODUCED BY STREPTOMYCES ORIENTALIS FROM INDONESIAN AND INDIAN SOIL.

USED AS AN ANTIBACTERIAL

**TABLE 6 : LIST OF HALOGENATION REACTIONS
WITH MACRO/BIOMOLECULES AND NMR ANALYSIS
RESULTS**

LIST OF REACTIONS	NMR RESULTS
DITHRANOL + CONC. HCl + DMSO	UNINTERPRETABLE
DITHRANOL + CONC. HBr + DMSO	UNINTERPRETABLE
ANTHRAROBIN + CONC. HCl + DMSO	UNINTERPRETABLE
ANTHRAROBIN + CONC. HBr + DMSO	UNINTERPRETABLE
O- PHENYLPHENOL + CONC. HCl + DMSO	UNINTERPRETABLE
O- PHENYLPHENOL + CONC. HBr + DMSO	UNINTERPRETABLE
PHLOROGLUCINOL + CONC. HCl + DMSO	UNINTERPRETABLE
PHLOROGLUCINOL + CONC. HBr + DMSO	UNINTERPRETABLE
1,8 DIAMINONAPHTHALENE + CONC. HCl + DMSO	UNINTERPRETABLE
1,8 DIAMINONAPHTHALENE + CONC. HBr + DMSO	UNINTERPRETABLE
O- PHENYLENEDIAMINE + CONC. HCl + DMSO	UNINTERPRETABLE
O- PHENYLENEDIAMINE + CONC. HBr + DMSO	UNINTERPRETABLE
3 AMINOPHENOL + CONC. HCl + DMSO	UNINTERPRETABLE
3 AMINOPHENOL + CONC. HBr + DMSO	UNINTERPRETABLE
PHENYLAMINO SALICYLATE + CONC. HCl + DMSO	UNINTERPRETABLE
PHENYLAMINO SALICYLATE + CONC. HBr + DMSO	UNINTERPRETABLE
4 ACETAMIDOPHENOL + CONC. HCl + DMSO	UNINTERPRETABLE
4 ACETAMIDOPHENOL + CONC. HBr + DMSO	UNINTERPRETABLE
VINCAMINE + CONC.HCl + DMSO	UNINTERPRETABLE
VINCAMINE + CONC. HBr + DMSO	UNINTERPRETABLE

ERGOCORNINE + CONC. HCl + DMSO	UNINTERPRETABLE
ERGOCORNINE + CONC. HBr + DMSO	UNINTERPRETABLE
VANCOMYCIN + CONC. HCl +DMSO	UNINTERPRETABLE
VANCOMYCIN + CONC. HBr + DMSO	UNINTERPRETABLE

TABLE 7 : GC/MS ANALYSIS OF HALOGENATED PRODUCTS FORMED AS A RESULT OF HALOGENATION REACTIONS OF MACRO/BIOMOLECULES USING THE SAME CONDITIONS

S.N	LIST OF REACTIONS	HALOGENATED PRODUCT FORMED	PERCENTAGE PRODUCT FORMED
1.	DITHRANOL + CONC. HCl + DMSO	- ve	- ve
2.	DITHRANOL + CONC. HBr + DMSO	- ve	- ve
3.	ANTHRAROBIN + CONC. HCl + DMSO	- ve	- ve
4.	ANTHRAROBIN + CONC. HBr + DMSO	- ve	- ve
5.	O- PHENYLPHENOL + CONC. HCl + DMSO	A) 5-Chloro-o-Phenyl phenol B) 3-Chloro-o-Phenyl phenol C) 2,2',3,4,5-pentaChloro-o-Phenyl phenol	3.57% 2.08% 0.4%
6.	O- PHENYLPHENOL + CONC. HBr + DMSO	A) 3,5-Dibromo-o-Phenyl phenol B) Bromophenoxy-Benzene	31.38% 3.78%
7.	PHLOROGLUCINOL + CONC. HCl + DMSO	- ve	- ve
8.	PHLOROGLUCINOL + CONC. HBr + DMSO	- ve	- ve
9.	1,8 DIAMINO - NAPHTHALENE + CONC. HCl + DMSO	- ve	- ve
10.	1,8 DIAMINO - NAPHTHALENE + CONC. HBr + DMSO	- ve	- ve
11.	O-PHENYLENEDIAMINE + CONC.HCl + DMSO	- ve	- ve
12.	O-PHENYLENEDIAMINE + CONC.HBr + DMSO	- ve	- ve
13.	3 AMINOPHENOL + CONC. HCl + DMSO	2,4-Dichloro 3-amino phenol	4.00%
14.	3 AMINOPHENOL + CONC. HBr + DMSO	2,4-Dibromo 3-amino phenol	18.35%
15.	PHENYLAMINO - SALICYLATE + CONC. HCl + DMSO	- ve	- ve

16.	PHENYLAMINO - SALICYLATE + CONC. HBr + DMSO	- ve	- ve
17.	4 ACETAMIDOPHENOL + CONC. HBr + DMSO	- ve	- ve
18.	4 ACETAMIDOPHENOL + CONC. HCl + DMSO	- ve	- ve
19.	VINCAMINE + CONC. HCl + DMSO	ve	- ve
20.	VINCAMINE + CONC. HBr + DMSO	- ve	- ve
21.	ERGOCORNINE + CONC. HCl + DMSO	- ve	- ve
22.	ERGOCORNINE + CONC. HBr + DMSO	- ve	- ve
23.	VANCOMYCIN + CONC. HCl + DMSO	- ve	- ve
24.	VANCOMYCIN + CONC. HBr + DMSO	- ve	- ve

RESULTS AND DISCUSSION

SIGNIFICANCE OF THE RESULTS ACHIEVED:-

- 1) Environmentally benign conditions were established for safe and simple halogenation experiments in the undergraduate laboratory
- 2) NMR and GC/MS analysis were used to verify the formation of specific isomers

POTENTIAL USE:-

- 1) Synthesis of halogenated biomolecules sensitive to pathogenic microbes in an attempt to enhance their bioactivity and reduce toxicity.
- 2) Minimizing environmental pollution, lessening hazards and ensuring safe disposal of waste products and toxic byproducts specifically to demonstrate electrophilic aromatic substitution in the undergraduate laboratory.

CONCLUSION

Aqueous HBr and HCl in the presence of DMSO prove to be good halogenating reagents of aromatic systems in the reactions previously described. Aqueous HI shows very little or no reactivity at all even though it is a stronger acid and should protonate the DMSO more effectively. Aqueous HF was also used, but there was no evidence that a reaction took place. This would be expected because fluorine does not like to be even partially positive, which is a necessity for electrophilic aromatic substitution. Gaseous HCl has almost the same reactivity as aqueous HCl. The only advantage in using gaseous HCl over aqueous HCl is that water interferes during NMR or GC/MS analysis of the reaction samples. The disadvantage in using gaseous HCl is that it is hazardous to use gas cylinders in undergraduate laboratories which besides being expensive, can pose a great safety hazard, cause environmental pollution and lead to unsafe handling problems.

The NMR data were uninterpretable for the reactions of macro/biomolecules in the presence aqueous acids and DMSO, because it seemed that water present in haloacids and deuterated DMSO(which absorbs water readily) was interfering with the final results, and the expected NMR pattern was not showing up clearly in the final results. Gaseous HCl was used for some of the reactions *instead* of aqueous HCl acid, but again because of interference of water present in DMSO (which appeared as a dominant peak in the NMR spectrum) and also

because of the complex aromatic region of most of these macro/biomolecules, the NMR spectrum was difficult to interpret.

Since all macro/biomolecules selected had $-OH$ and $-NH_2$ as activating groups, and conditions for halogenation were also set according to those used for halogenating simple aromatic systems, it was assumed that some kind of halogenation reaction is taking place in the NMR tube even though NMR patterns were uninterpretable. Most of the reactions with macro/biomolecules did not show any positive GC/MS results. In some cases it was concluded that since most of these macromolecules have complex chemical structures with polar and non-polar groups and high boiling points, they tend to stick to the methyl siloxane GC/MS column and do not elute. This can be supported by the fact that even the parent molecule of these macro/biomolecules did not elute from the column. In some cases, as with Vancomycin and Phenyl Amino Salicylate, fragments of the molecules which were halogenated eluted from the column. It was inferred that halogenation might be taking place on the complex aromatic structure of these molecules, but since they did not elute as whole molecules during GC/MS analysis the above statement cannot be supported by any kind of evidence. Molecules like O-Phenyl phenol and 3-Amino phenol showed GC/MS evidence of halogenation when their halogenated products were successfully eluted from the GC/MS column by using a different eluting solvent i.e. CH_2Cl_2 rather than regular DMSO or THF. From this, a conclusion can be drawn that infact some macro/biomolecules can be halogenated using the conditions established for halogenation. Slight modification of the

halogenation conditions used and the use of suitable diluting and eluting solvents for analysis could be useful.

During GC/MS analysis, the percentage of halogenated products formed from all the products eluted from the GC/MS column and reported in tables 5 and 7 are not in very high yields. Many of the products could have been obtained from previous analysis and eluted with the solvents used in our GC/MS method of analysis. Isomer percentages were based on the area of individual isomers divided by the total area observed for all the products eluting from the GC/MS column.

Whatever the limitations, it can be successfully concluded from extensive halogenation reactions with simple aromatic systems that the reactions of haloacids with DMSO can be used to demonstrate electrophilic aromatic substitution in an undergraduate laboratory.

Similar conditions with slight modifications can be used to halogenate some bio/macromolecules.

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APPENDIX A

Page

Spectrum 1. Expanded ^1H NMR Spectrum of Standard Aniline in DMSO.....	A1
Spectrum 2. Expanded ^1H NMR Spectrum of Aniline and aqueous HCl in DMSO.....	A1
Spectrum 3. Expanded ^1H NMR Spectrum of Aniline and aqueous HBr in DMSO.....	A2
Spectrum 4. Expanded ^1H NMR Spectrum of Standard N-Methylaniline in DMSO.....	A3
Spectrum 5. Expanded ^1H NMR Spectrum of N-Methylaniline + aqueous HCl in DMSO.....	A3
Spectrum 6. Expanded ^1H NMR Spectrum of N-Methylaniline + aqueous HBr in DMSO.....	A4
Spectrum 7. Expanded ^1H NMR Spectrum of Standard N,N-Dimethylaniline in DMSO.....	A5
Spectrum 8. Expanded ^1H NMR Spectrum of N,N-Dimethylaniline + aqueous HCl in DMSO.....	A5
Spectrum 9. Expanded ^1H NMR Spectrum of N,N-Dimethylaniline + aqueous HBr in DMSO.....	A6
Spectrum 10. Expanded ^1H NMR Spectrum of N,N-Dimethylaniline + aqueous HI in DMSO.....	A6
Spectrum 11. Expanded ^1H NMR Spectrum of Standard Phenol in DMSO.....	A7
Spectrum 12. Expanded ^1H NMR Spectrum of Phenol + aqueous HCl in DMSO.....	A7
Spectrum 13. Expanded ^1H NMR Spectrum of Phenol + aqueous HBr in DMSO.....	A8

Spectrum 14. Expanded ^1H NMR Spectrum of Phenol + aqueous HI in DMSO.....	A8
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Spectrum 16. Expanded ^1H NMR Spectrum of Anisole + gaseous HCl in DMSO.....	A9
Spectrum 17. ^1H NMR Spectrum of Standard 3-Aminophenol in DMSO.....	A10
Spectrum 18. ^1H NMR Spectrum of 3-Aminophenol + aqueous HCl in DMSO.....	A10
Spectrum 19. ^1H NMR Spectrum of 3-Aminophenol + aqueous HBr in DMSO.....	A11
Spectrum 20. ^1H NMR Spectrum of Standard o-Phenylphenol in DMSO.....	A12
Spectrum 21. ^1H NMR Spectrum of o-Phenylphenol + aqueous HCl in DMSO.....	A12
Spectrum 22. ^1H NMR Spectrum of o-Phenylphenol + aqueous HBr in DMSO.....	A13

APPENDIX B

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Spectrum 1. GC/MS Spectrum of Aniline (Note:-This GC/MS spectrum is shown as as a representative example of many spectrums of parent compound which were taken each time prior to analysis of various reaction sample mixtures).....	B1
Spectrum 2. GC/MS Spectrum of p-Chloroaniline.....	B2
Spectrum 3. GC/MS Spectrum of 4-Bromoaniline.....	B3
Spectrum 4. GC/MS Spectrum of 4-Chloro-N-methylaniline.....	B4
Spectrum 5. GC/MS Spectrum of 4-Bromo-N-methylaniline.....	B5
Spectrum 6. GC/MS Spectrum of 4-Bromo-N,N-dimethylaniline.....	B6
Spectrum 7. GC/MS Spectrum of p-Chlorophenol.....	B7
Spectrum 8. GC/MS Spectrum of 4-Bromophenol & 3-Bromophenol (Note:-This GC/MS spectrum is shown as a representative example of many spectrums which showed 2 or more isomers during analysis of various sample mixtures but here GC/MS spectrums of only one important isomer of each reaction sample type is shown).....	B8
Spectrum 9. GC/MS Spectrum of 1-Chloro-4-methoxybenzene (4-Chloroanisole).....	B9
Spectrum 10. GC/MS Spectrum of 2,4-dichloro-3-aminophenol.....	B10
Spectrum 11. GC/MS Spectrum of 2,4-dibromo-3-aminophenol.....	B11
Spectrum 12. GC/MS Spectrum of 3-Chloro-[1,1'-Biphenyl]-2-ol (3-Chloro-o-Phenylphenol).....	B12
Spectrum 13. GC/MS Spectrum of 3,5-dibromo-[1,1'-Biphenyl]-2-ol (3,5-dibromo-o-Phenylphenol).....	B13

APPENDIX C

Page

Table of NMR Parameters.....	C1
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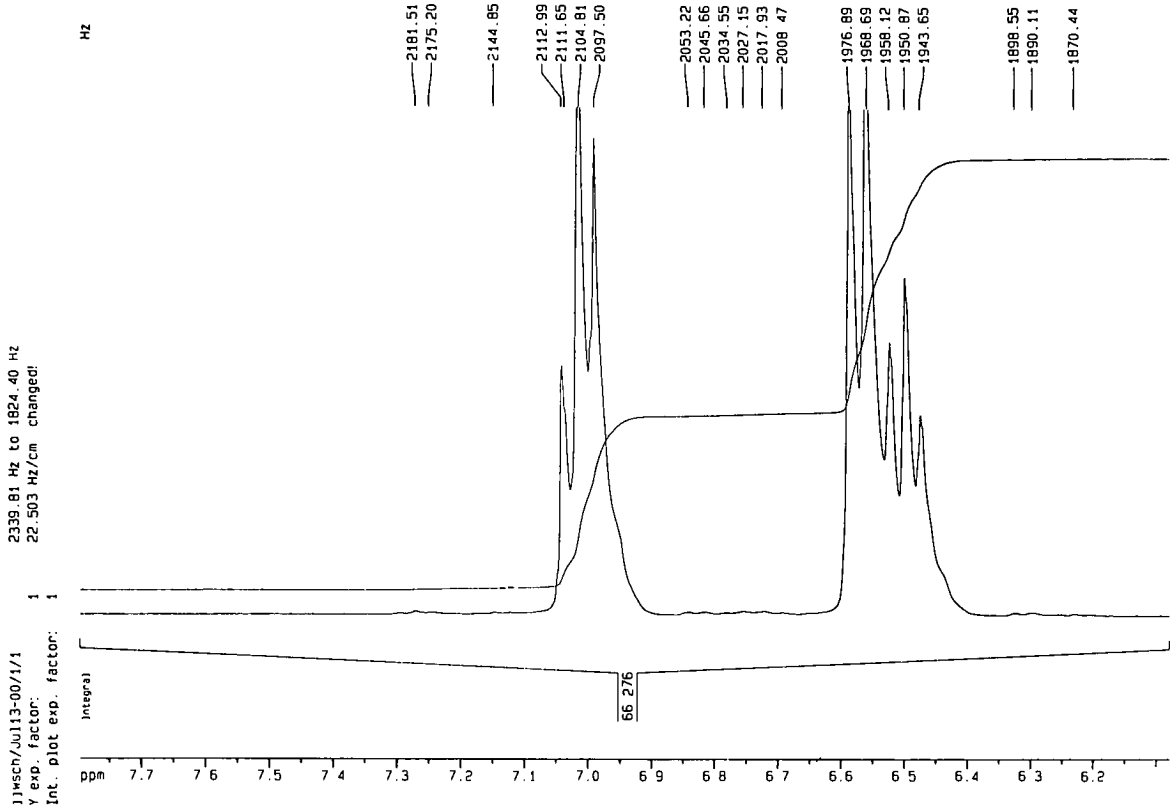
APPENDIX D

Page

Data Analysis (used during GC/MS analysis) Parameters.....D1

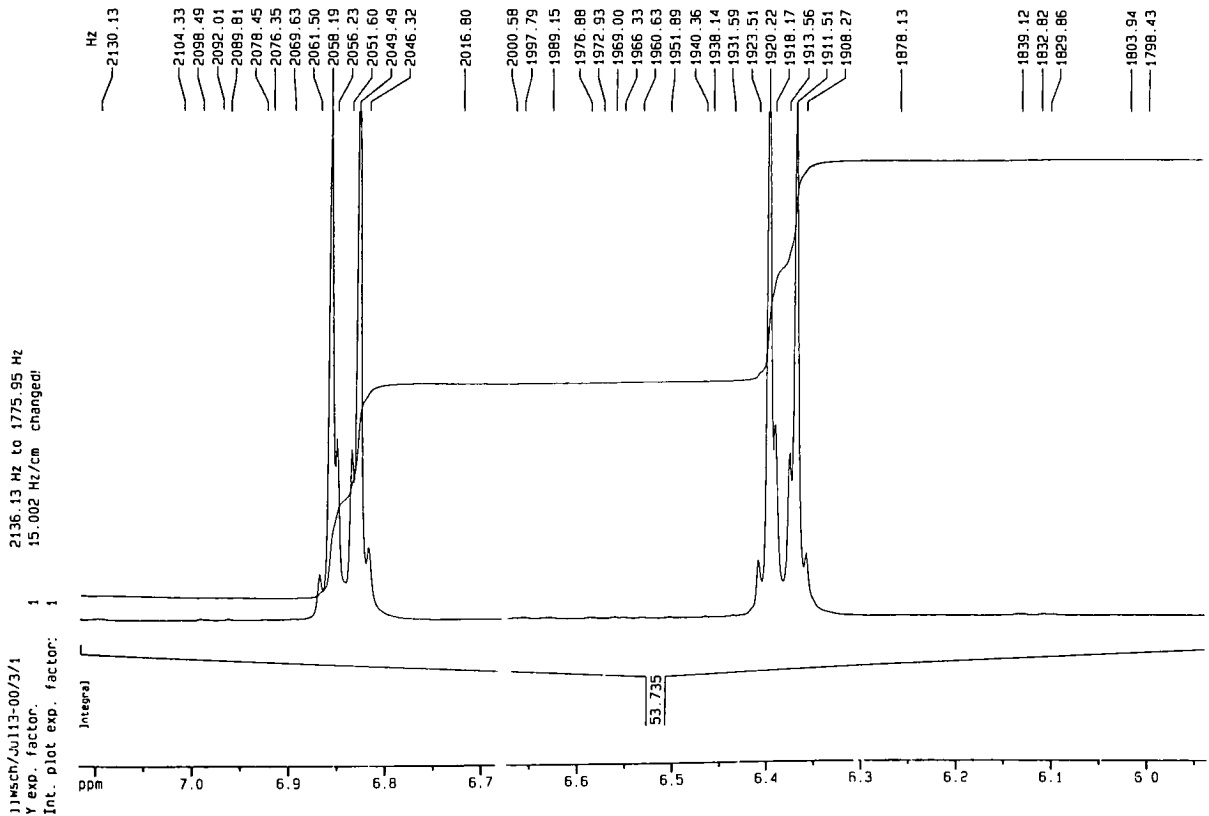
APPENDIX A

Spectrum 1.

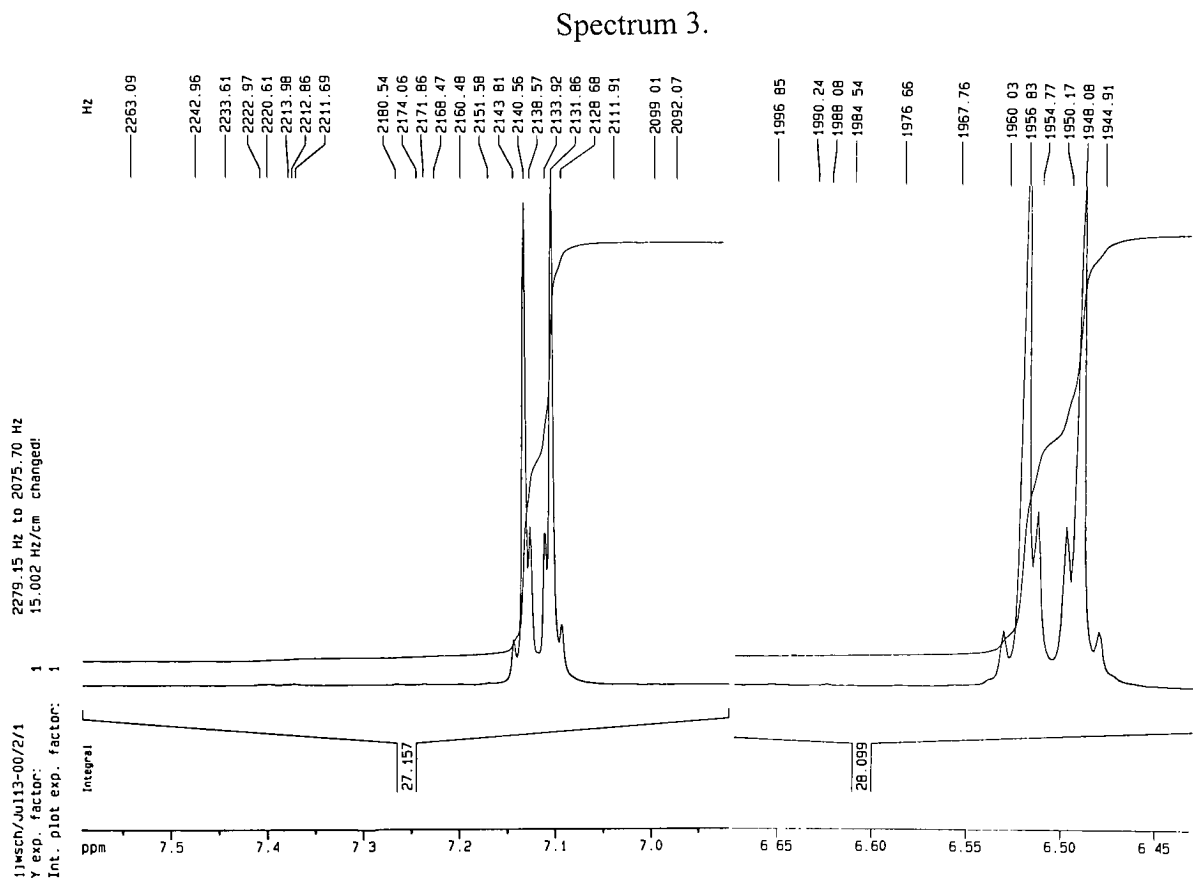


ANILINE (STANDARD) + DMSO

Spectrum 2.

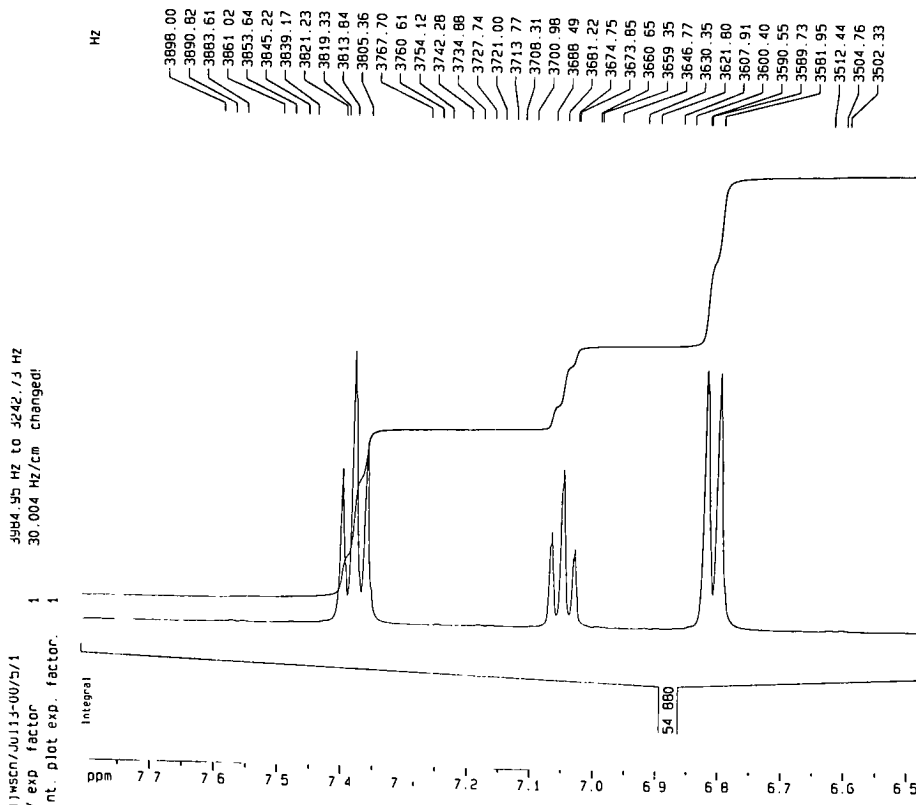


ANILINE + AQUEOUS HCl + DMSO



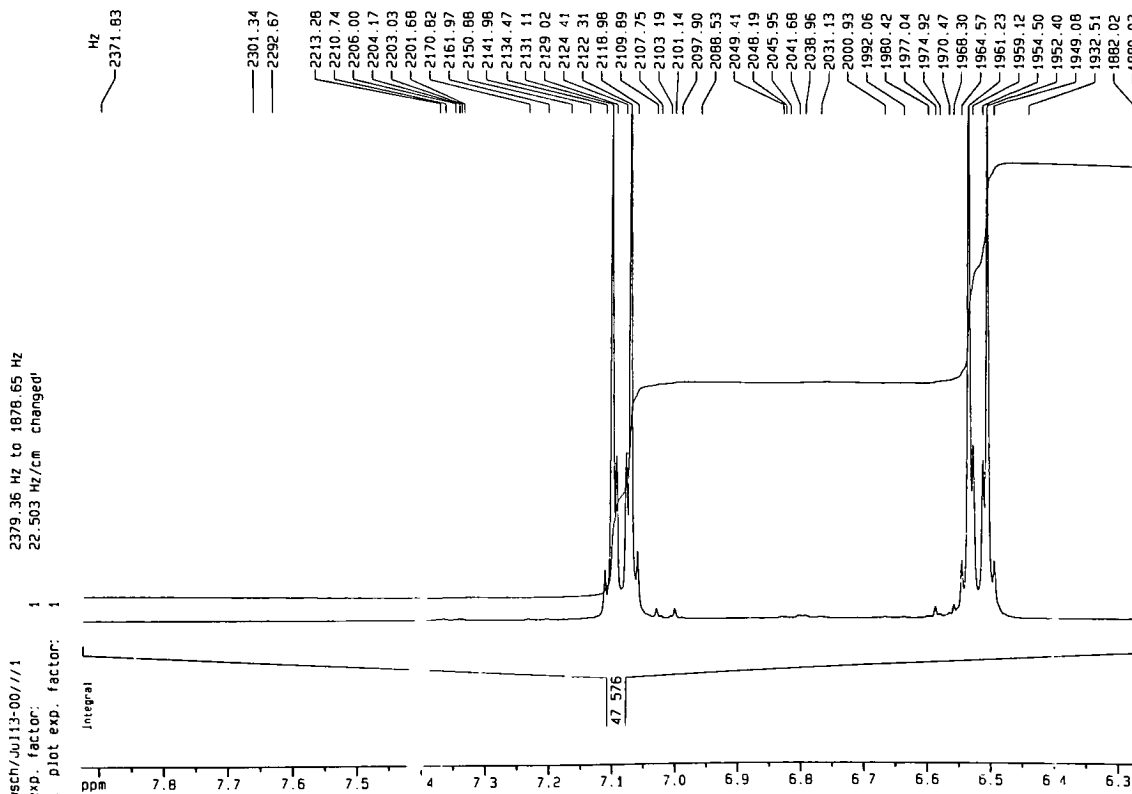
ANILINE + AQUEOUS HBr + DMSO

Spectrum 4.



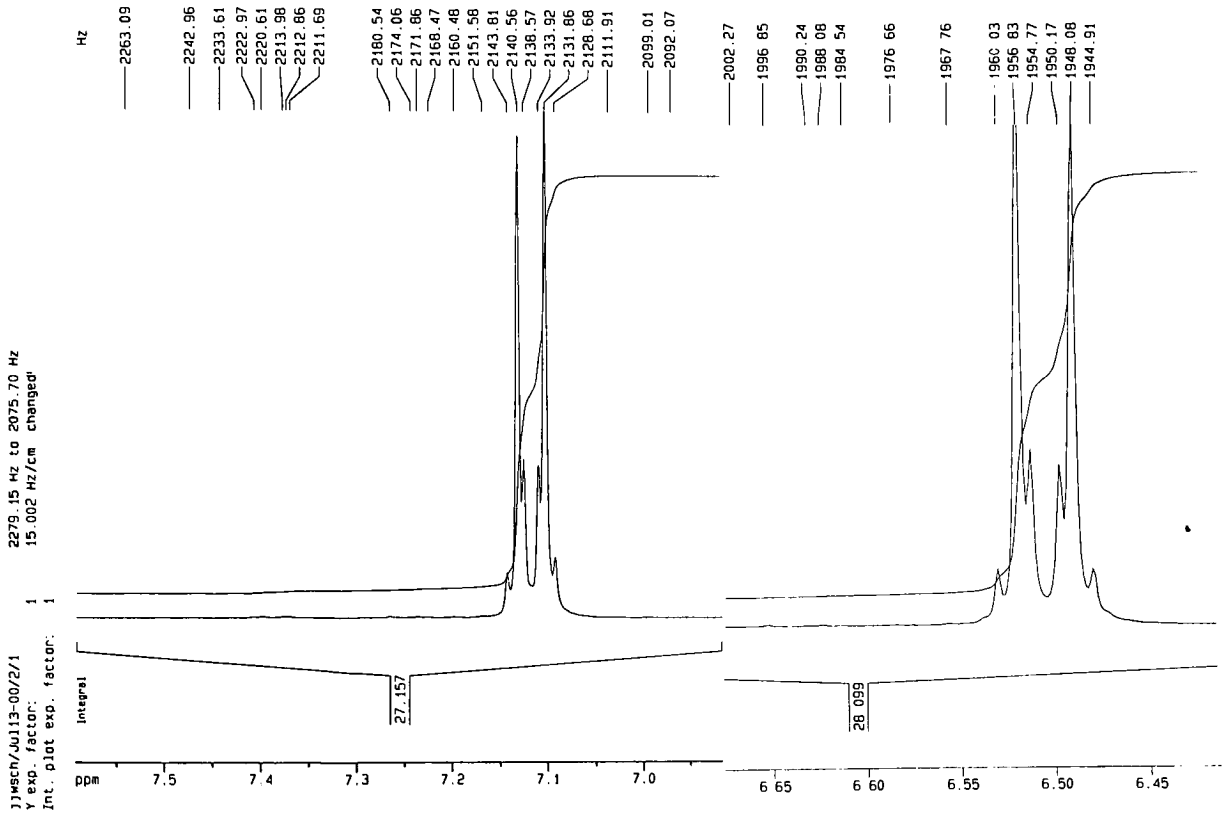
N METHYL ANILINE (STANDARD) + DMSO

Spectrum 5.



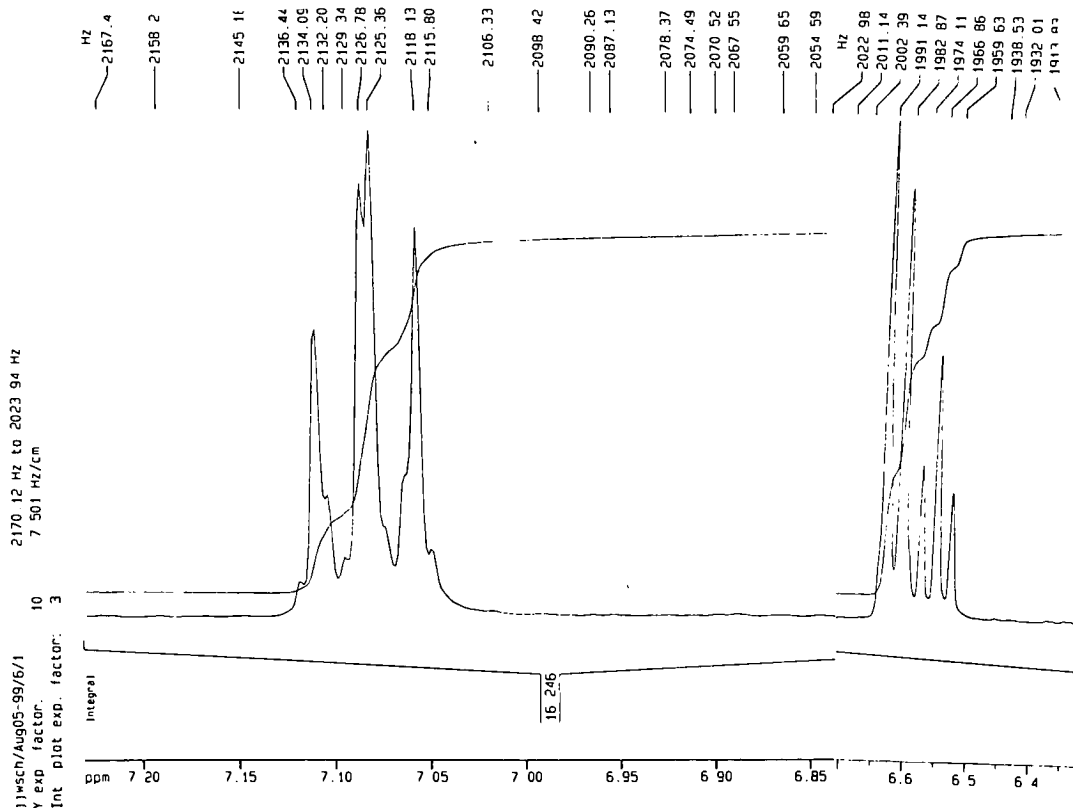
N METHYL ANILINE + AQUEOUS HCl + DMSO

Spectrum 6.



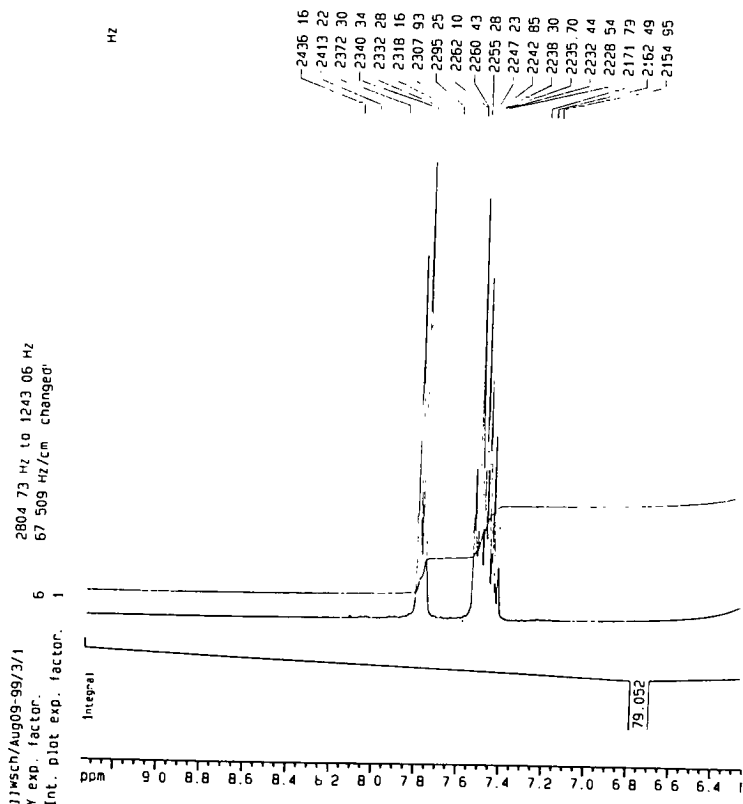
N METHYL ANILINE + AQUEOUS HBr + DMSO

Spectrum 7.



N,N DIMETHYL ANILINE (STANDARD) + DMSO

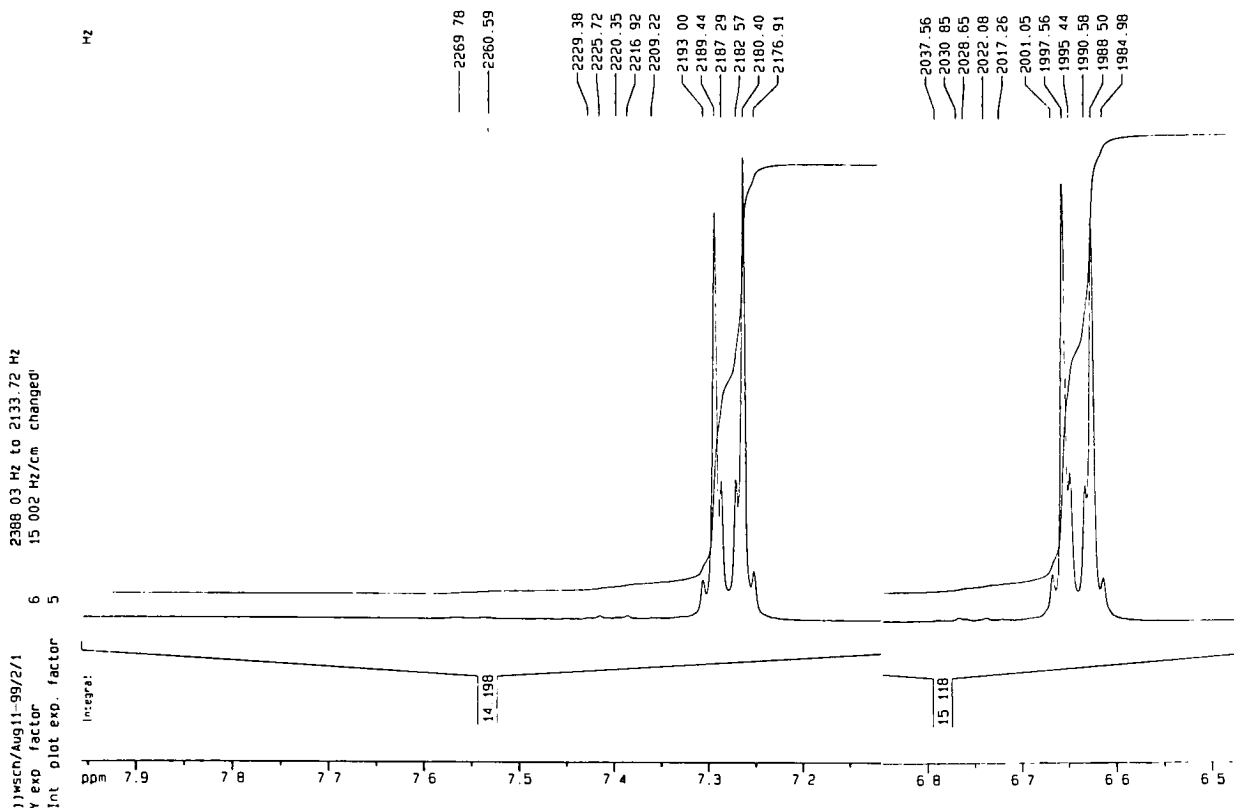
Spectrum 8.



N,N DIMETHYL ANILINE + AQUEOUS HCl + DMSO

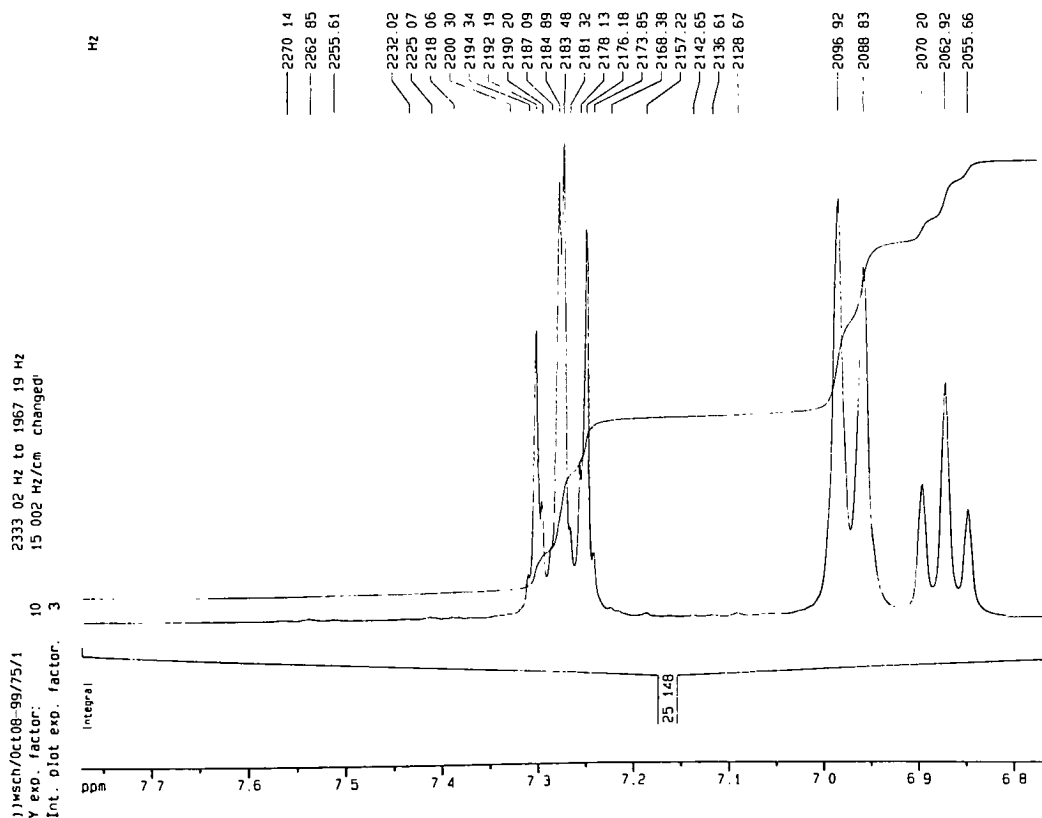
Spectrum 9.

A6



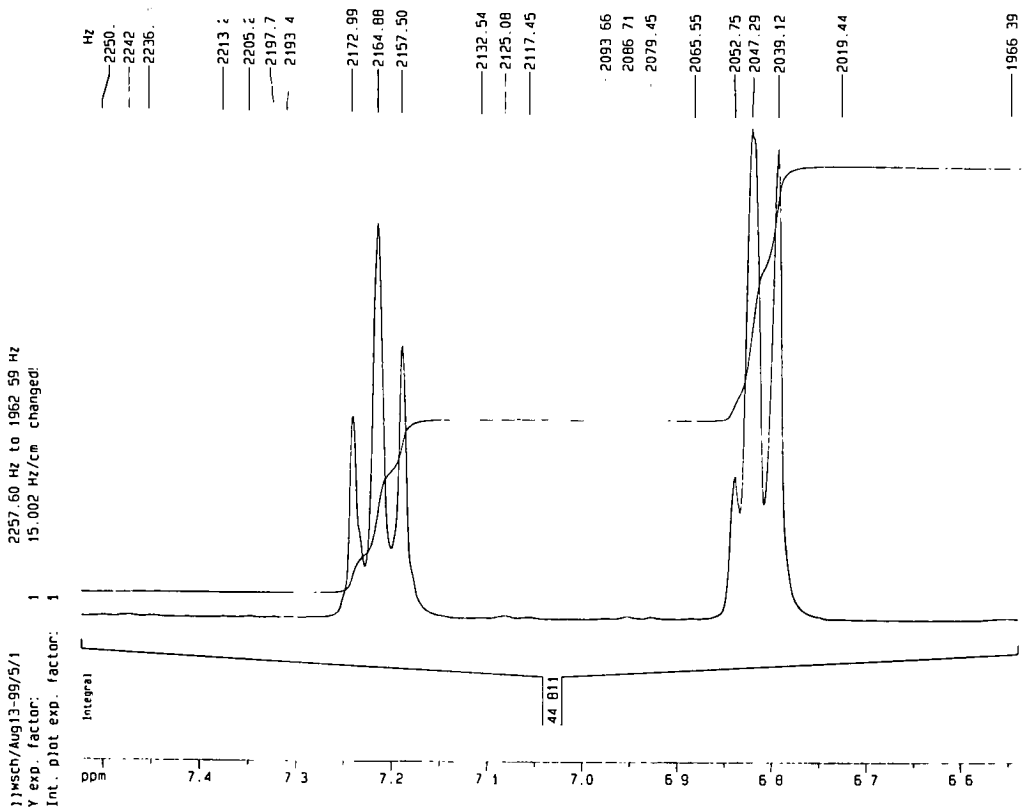
N,N DIMETHYL ANILINE + AQUEOUS HBr + DMSO

Spectrum 10.



N,N DIMETHYL ANILINE + AQUEOUS HI + DMSO

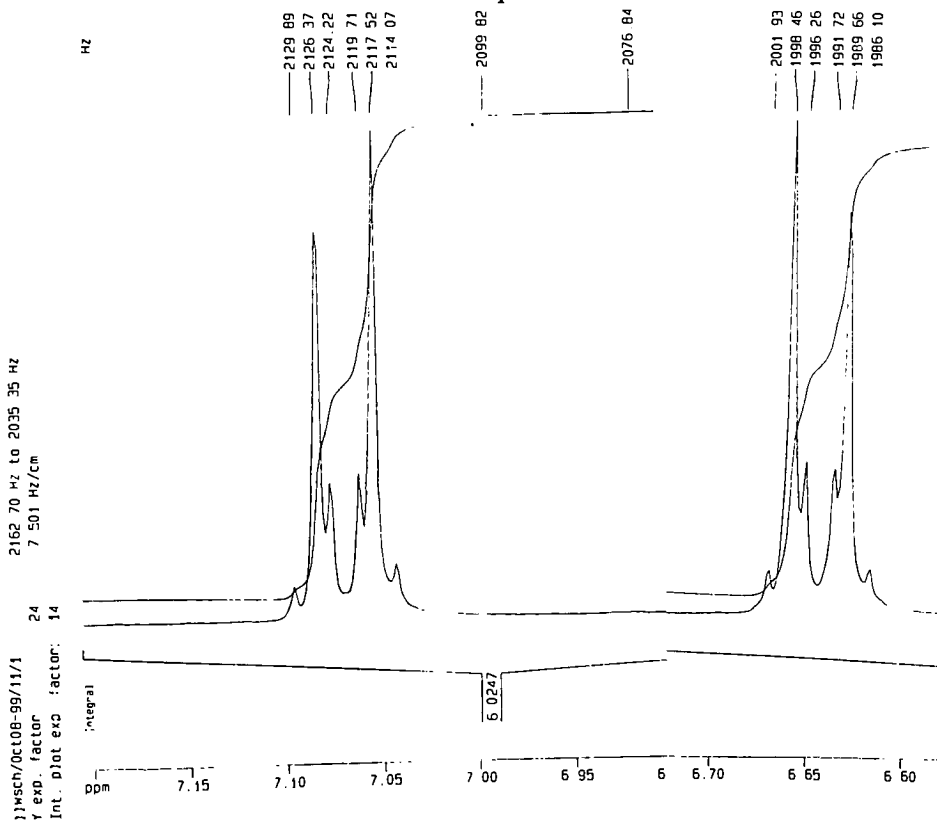
Spectrum 11.



A7

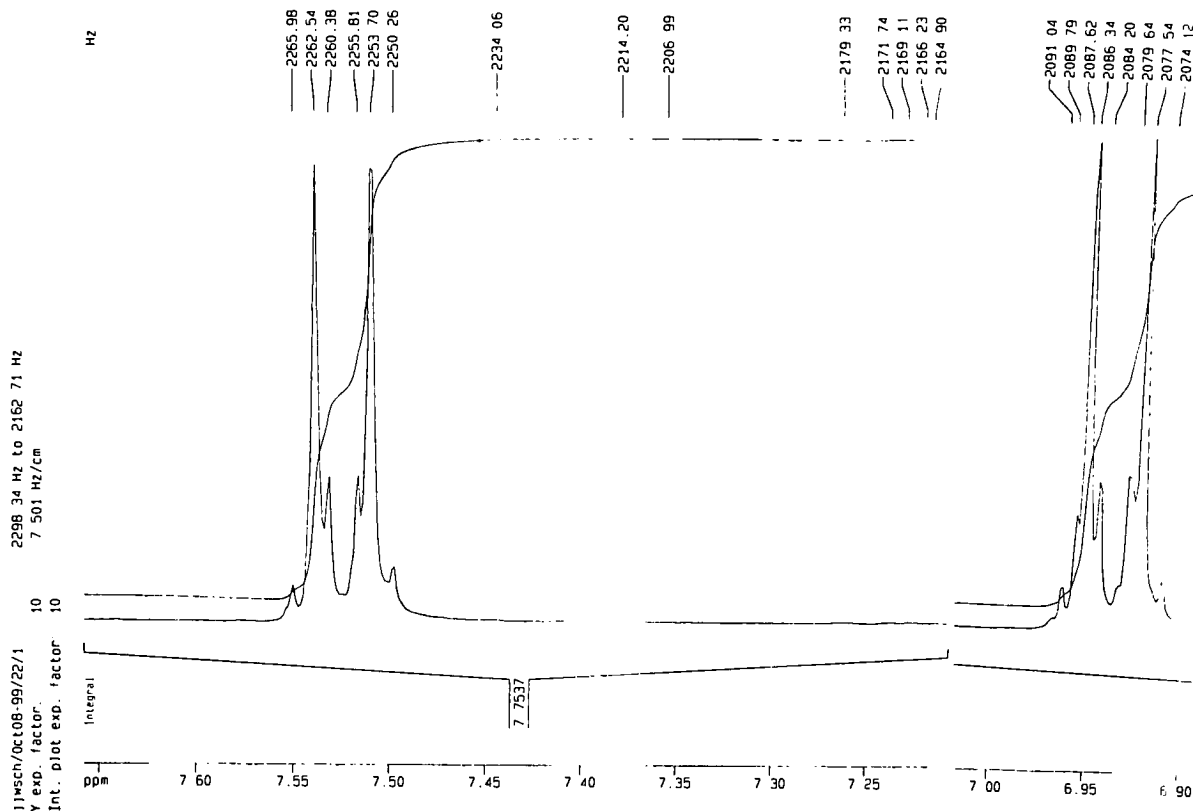
PHENOL (STANDARD) + DMSO

Spectrum 12.



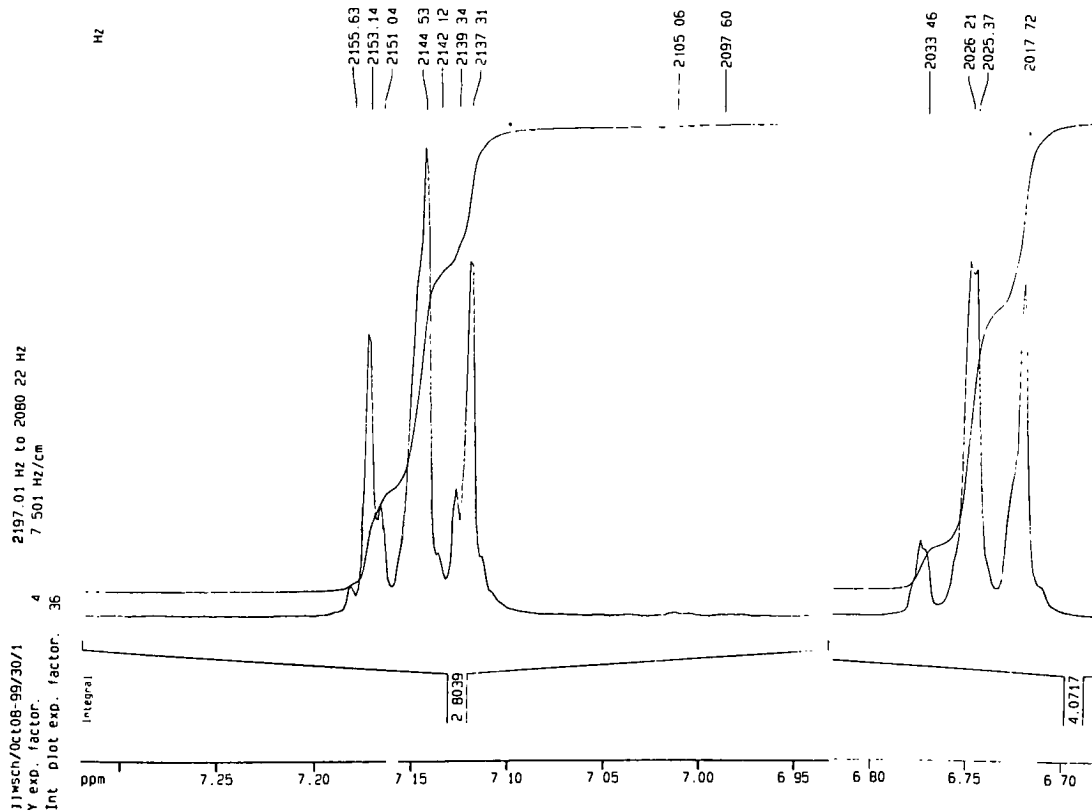
PHENOL + AQUEOUS HCl + DMSO

Spectrum 13.



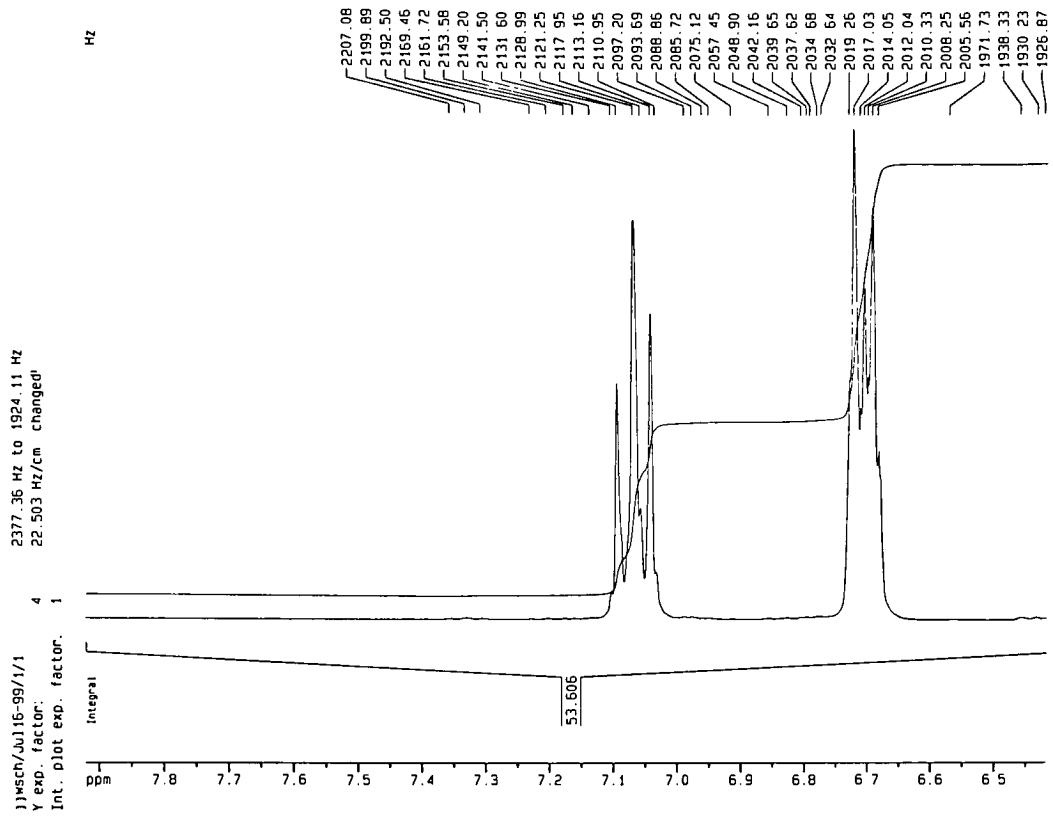
PHENOL + AQUEOUS HBr + DMSO

Spectrum 14.



PHENOL + AQUEOUS HI + DMSO

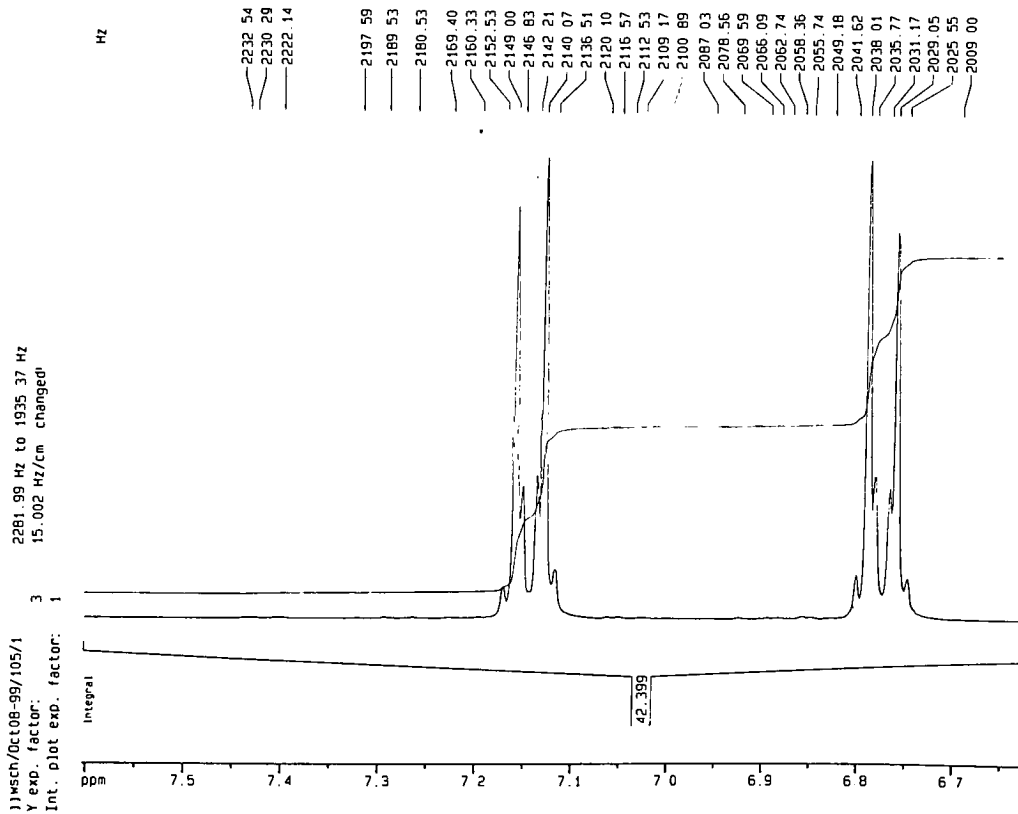
Spectrum 15.



A9

ANISOLE (STANDARD) + DMSO

Spectrum 16.



ANISOLE + GASEOUS HCL + DMSO

Spectrum 17.

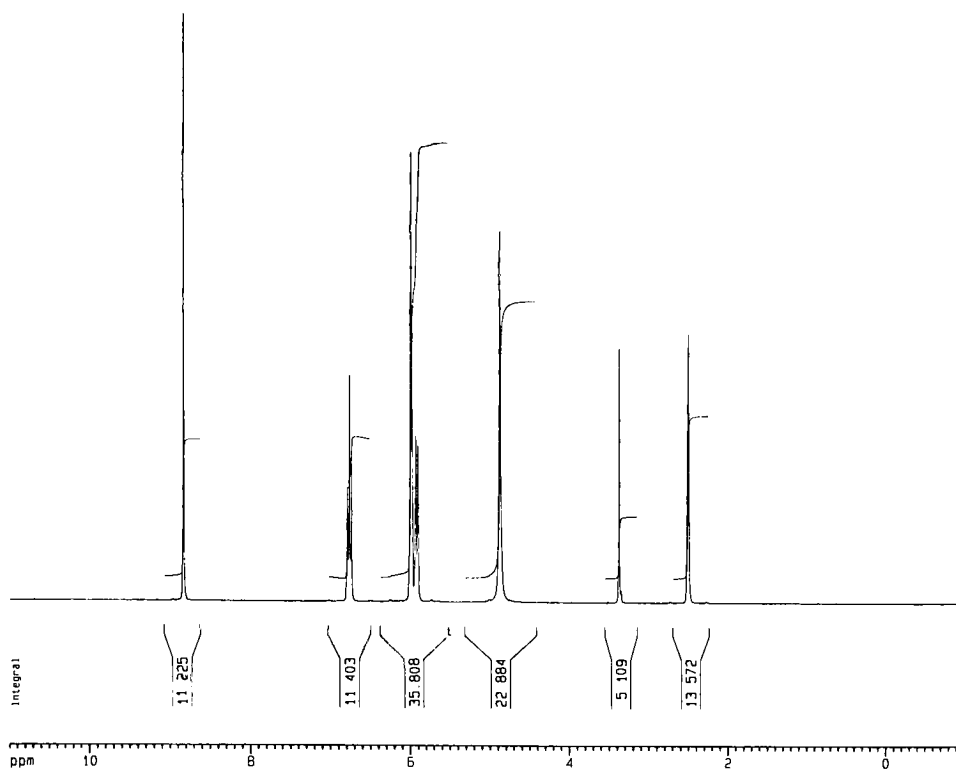
A10

Current Data Parameters
 NAME Jul24-00
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 500000
 Time 16 05
 INSTRUM sdeci
 PROBD0 5 mm ONP 1H
 PULPROG zg30
 TO 32768
 SOLVENT DMSO
 NS 16
 OS 2
 SMH 6172 839 Hz
 FT0RES 0 188380 Hz
 AQ 2 6542580 sec
 RG 256
 DM B1 000 usec
 OE 6 00 usec
 TE 300 0 K
 O1 1 0000000 sec
 P1 9 00 usec
 DE 6 00 usec
 SFO1 300 1318534 MHz
 NUC1 1H
 PL1 -5 00 dB

F2 - Processing parameters
 S1 16384
 SF 300 1300000 MHz
 MWM EM
 SSB 0
 LB 0 30 Hz
 GB 0
 PC 1 00

10 NMR plot parameters
 CX 20 00 cm
 F1P 11 000 ppm
 F1 3301 43 Hz
 F2P -1 000 ppm
 F2 -300 13 Hz
 PPMCM 0 60000 ppm/cm
 HZCM 180 07800 Hz/cm



O-PHENYL PHENOL (STANDARD) + DMSO

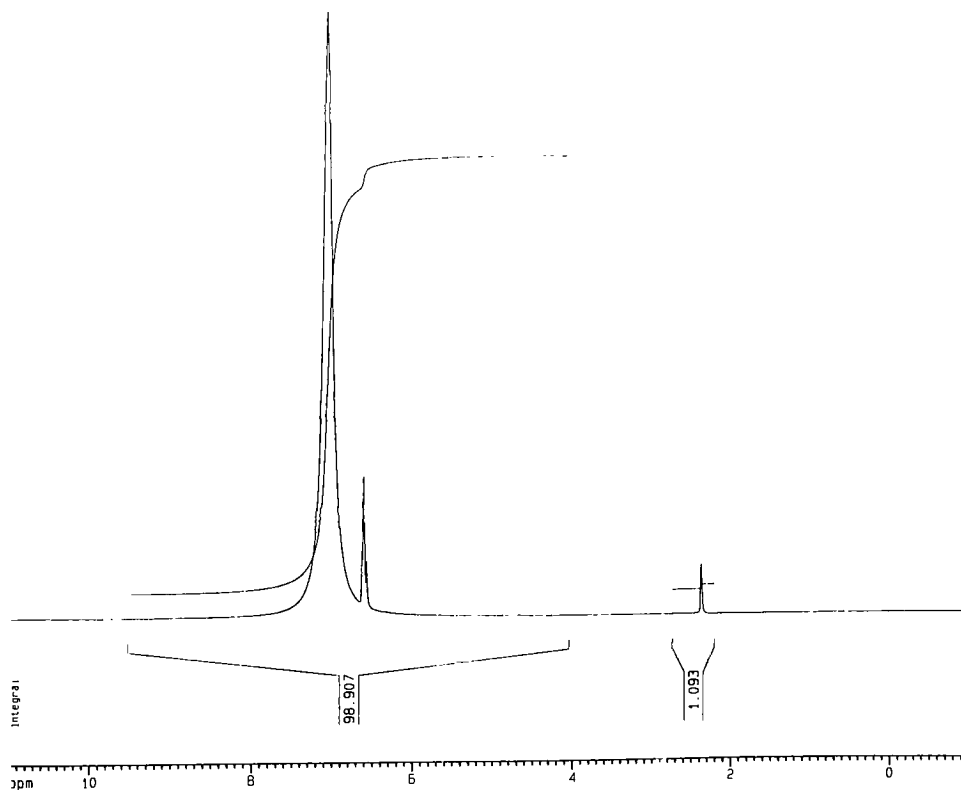
Spectrum 18.

Current Data Parameters
 NAME Jul24-00
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 500000
 Time 16 18
 INSTRUM sdeci
 PROBD0 5 mm ONP 1H
 PULPROG zg30
 TO 32768
 SOLVENT DMSO
 NS 16
 OS 2
 SMH 6172 839 Hz
 FT0RES 0 188380 Hz
 AQ 2 6542580 sec
 RG 28.5
 DM B1 000 usec
 OE 6 00 usec
 TE 300 0 K
 O1 1 0000000 sec
 P1 9 00 usec
 DE 6 00 usec
 SFO1 300 1318534 MHz
 NUC1 1H
 PL1 -5 00 dB

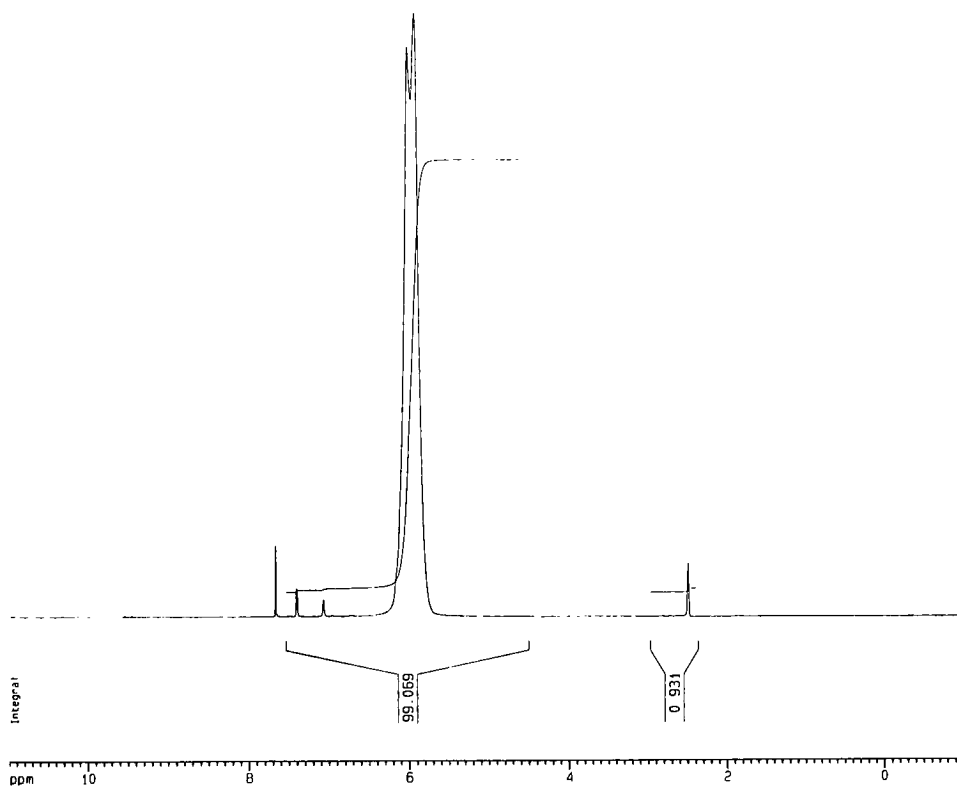
F2 - Processing parameters
 S1 16384
 SF 300 1300393 MHz
 MWM EM
 SSB 0
 LB 0 30 Hz
 GB 0
 PC 1 00

10 NMR plot parameters
 CX 20 00 cm
 F1P 11 000 ppm
 F1 3301 43 Hz
 F2P -1 000 ppm
 F2 -300 13 Hz
 PPMCM 0 60000 ppm/cm
 HZCM 180 07802 Hz/cm



O-PHENYL PHENOL + AQUEOUS HCl + DMSO

Spectrum 19.



Current Data Parameters
 NAME Jun19-00
 EXPNO 7
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 500000
 Time 17 18
 INSTRUM spect
 PROBHD 5 mm QNP 1H
 PULPROG zg30
 TD 32768
 SOLVENT DMSO
 NS 16
 DS 2
 SWH 6172.839 Hz
 FIDRES 0.180380 Hz
 AQ 2.6542580 sec
 RG 25.4
 OW 81.000 usec
 DE 6.00 usec
 TE 300.0 K
 O1 1.00000000 sec
 P1 9.00 usec
 OE 6.00 usec
 SF01 300.1318534 MHz
 NUC1 1H
 PL1 -5.00 dB

F2 - Processing parameters
 S1 16384
 SF 300.1300000 MHz
 MDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 3301.43 Hz
 F2P -1.000 ppm
 F2 -300.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 180.07800 Hz/cm

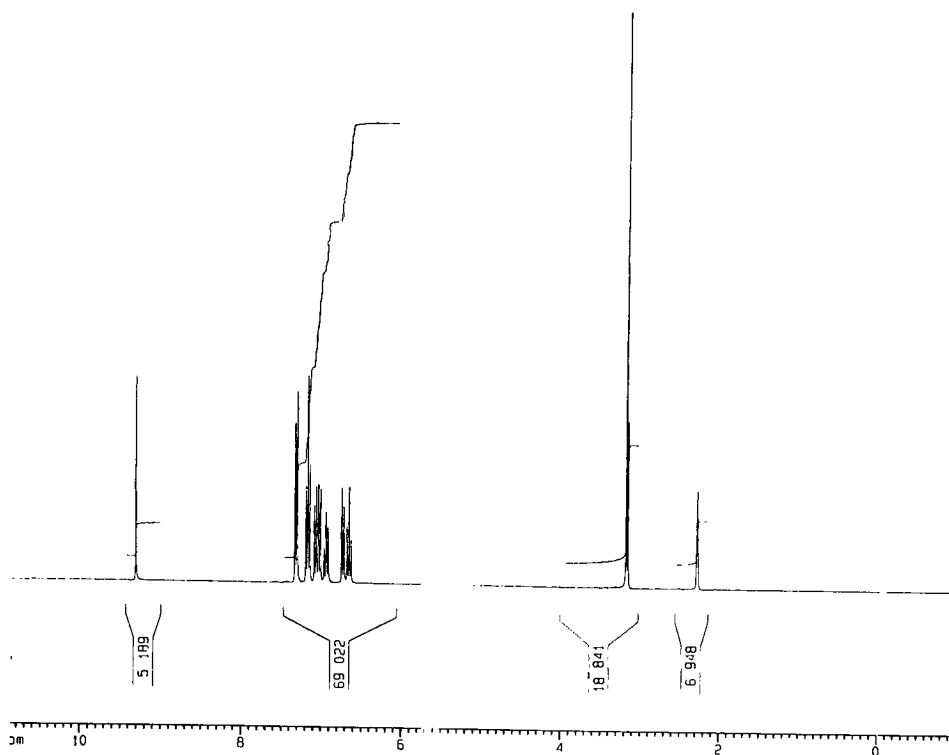
O-PHENYL PHENOL + AQUEOUS HBr + DMSO

Current Data Parameters
 NAME Jun19-00
 EXPNO 5
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 500000
 Time 16 40
 INSTRUM soect
 PROBHD 5 mm ONP 1H
 PULPROG zg30
 TO 32768
 SOLVENT DMSO
 NS 16
 OS 2
 SWH 6172 839 Hz
 FIDRES 0 188380 Hz
 AQ 2.6542580 sec
 RG 181
 OW 81 000 usec
 DE 6 00 usec
 TE 300 0 K
 O1 1 00000000 sec
 P1 9 00 usec
 DE 6 00 usec
 SF01 300 1318534 MHz
 NUC1 1H
 PL1 -5 00 dB

F2 - Processing parameters
 SI 16384
 SF 300 1300638 MHz
 WDW EM
 SSB 0
 LB 0 30 Hz
 GB 0
 PC 1 00

10 NMR plot parameters
 CX 20 00 cm
 F1P 11 000 ppm
 F1 3301 43 Hz
 F2P -1 000 ppm
 F2 -300 13 Hz
 PPMCM 0 60000 ppm/cm
 HZCM 180 07803 Hz/cm



3-AMINO PHENOL (STANDARD) + DMSO

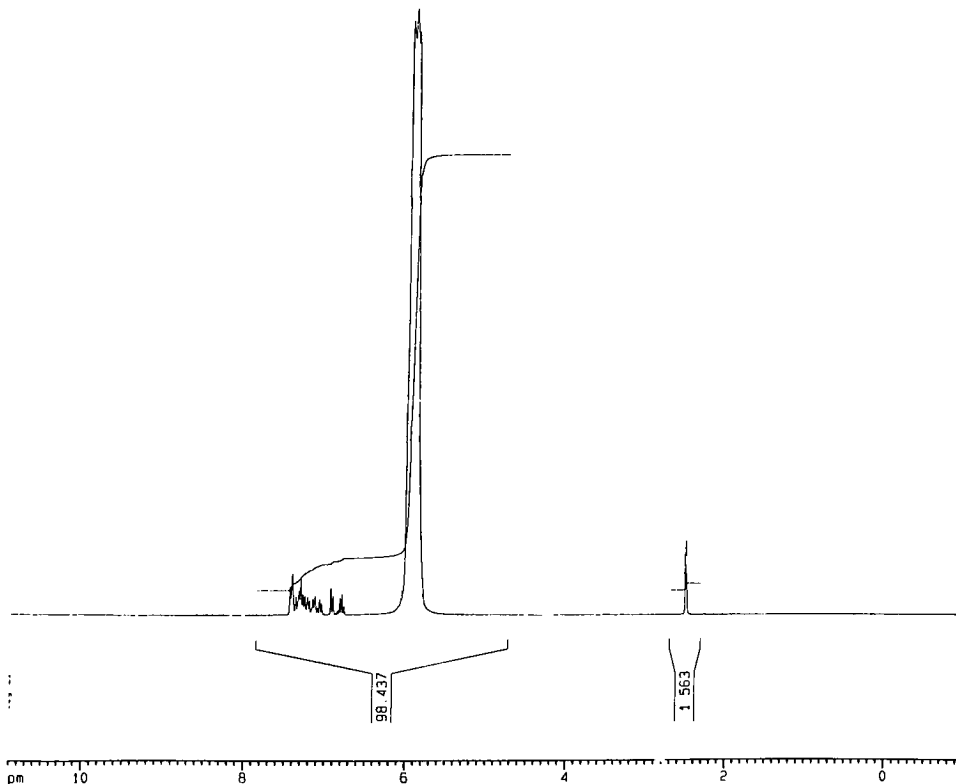
Spectrum 21.

Current Data Parameters
 NAME Jun19-00
 EXPNO 4
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 500000
 Time 16 29
 INSTRUM soect
 PROBHD 5 mm ONP 1H
 PULPROG zg30
 TO 32768
 SOLVENT DMSO
 NS 16
 OS 2
 SWH 6172 839 Hz
 FIDRES 0 188380 Hz
 AQ 2.6542580 sec
 RG 35 9
 OW 81 000 usec
 DE 6 00 usec
 TE 300 0 K
 O1 1 00000000 sec
 P1 9 00 usec
 DE 6 00 usec
 SF01 300 1318534 MHz
 NUC1 1H
 PL1 -5 00 dB

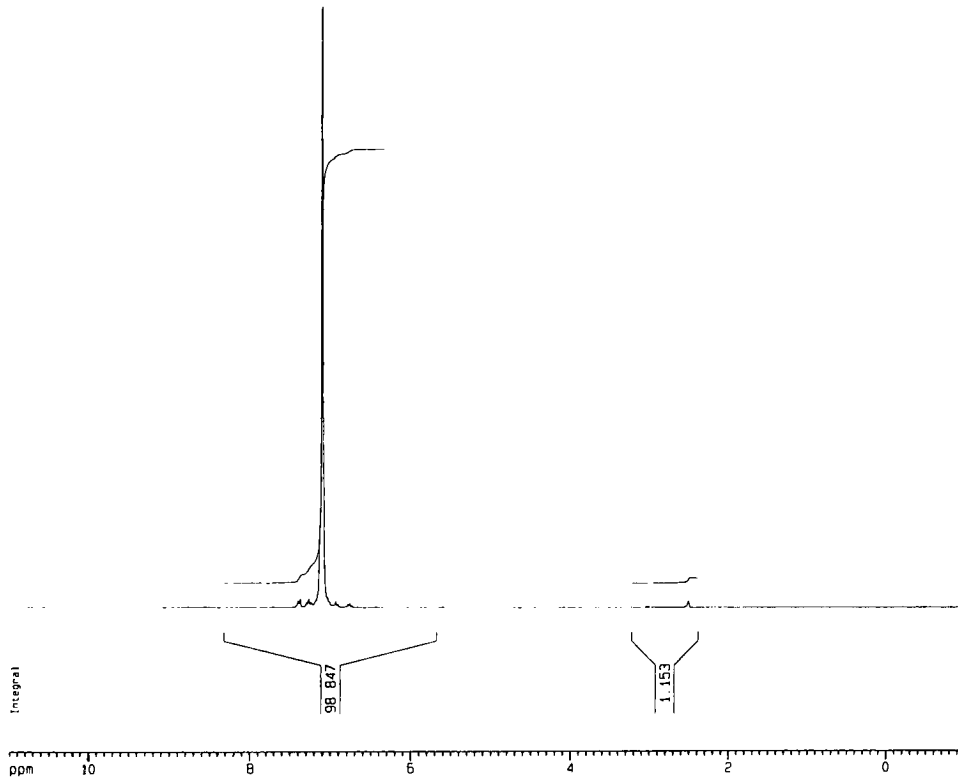
F2 - Processing parameters
 SI 16384
 SF 300 1300000 MHz
 WDW EM
 SSB 0
 LB 0 30 Hz
 GB 0
 PC 1 00

10 NMR plot parameters
 CX 20 00 cm
 F1P 11 000 ppm
 F1 3301 43 Hz
 F2P -1 000 ppm
 F2 -300 13 Hz
 PPMCM 0 60000 ppm/cm
 HZCM 180 07800 Hz/cm



3-AMINO PHENOL + AQUEOUS HCl + DMSO

Spectrum 22.



Current Data Parameters
 NAME Jun19-00
 EXPNO 6
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 500000
 Time 16 51
 INSTRUM spect
 PROBHD 5 mm QNP 1H
 PULPROG zg30
 TO 32768
 SOLVENT DMSO
 NS 16
 OS 2
 SMH 6172.839 Hz
 FIDRES 0.189380 Hz
 AQ 2.6542580 sec
 RG 28.5
 OW 81.000 usec
 DE 6.00 usec
 TE 300.0 K
 O1 1.0000000 sec
 P1 9.00 usec
 DE 6.00 usec
 SFO1 300.1318534 MHz
 NUC1 1H
 PL1 -5.00 dB

F2 - Processing parameters
 SI 16384
 SF 300.1300000 MHz
 WMW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 3301.43 Hz
 F2P -1.000 ppm
 F2 -300.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 180.07800 Hz/cm

3-AMINO PHENOL + AQUEOUS HBr + DMSO

APPENDIX B

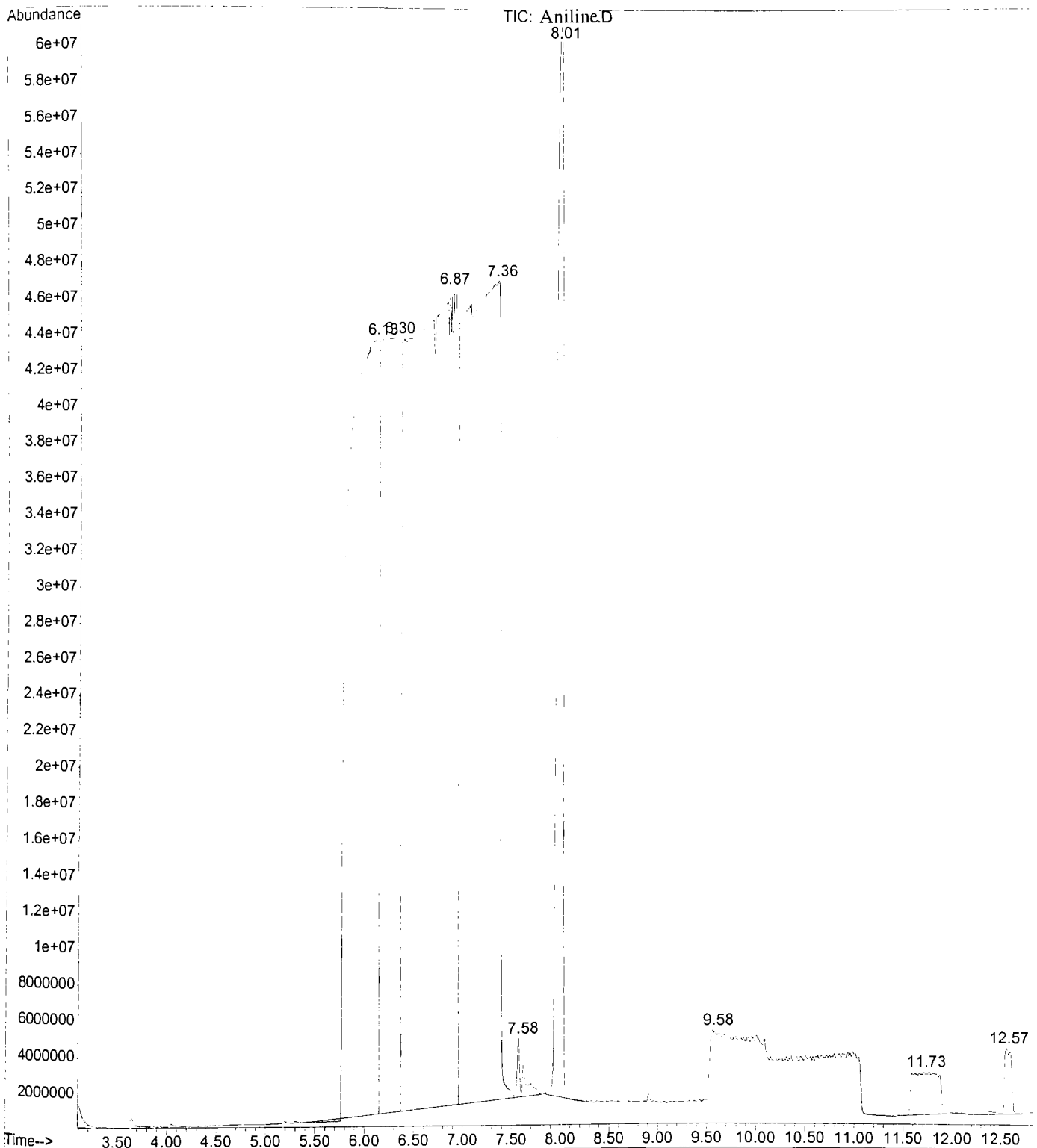
GC Spectrum 1. Area Percent Report

B1

Data File : C:\HPCHEM\1\DATA\ Aniline.D
Acq On : 13 Jul 2000 15:13
Sample :
Misc :

Vial: 1
Operator:
Inst : RIT Chemi
Multiplr: 1.00
Sample Amount: 0.00

MS Integration Params: autoint1.e
Method : C:\HPCHEM\1\5973\PANKAJ.M (Chemstation Integrator)
Title :

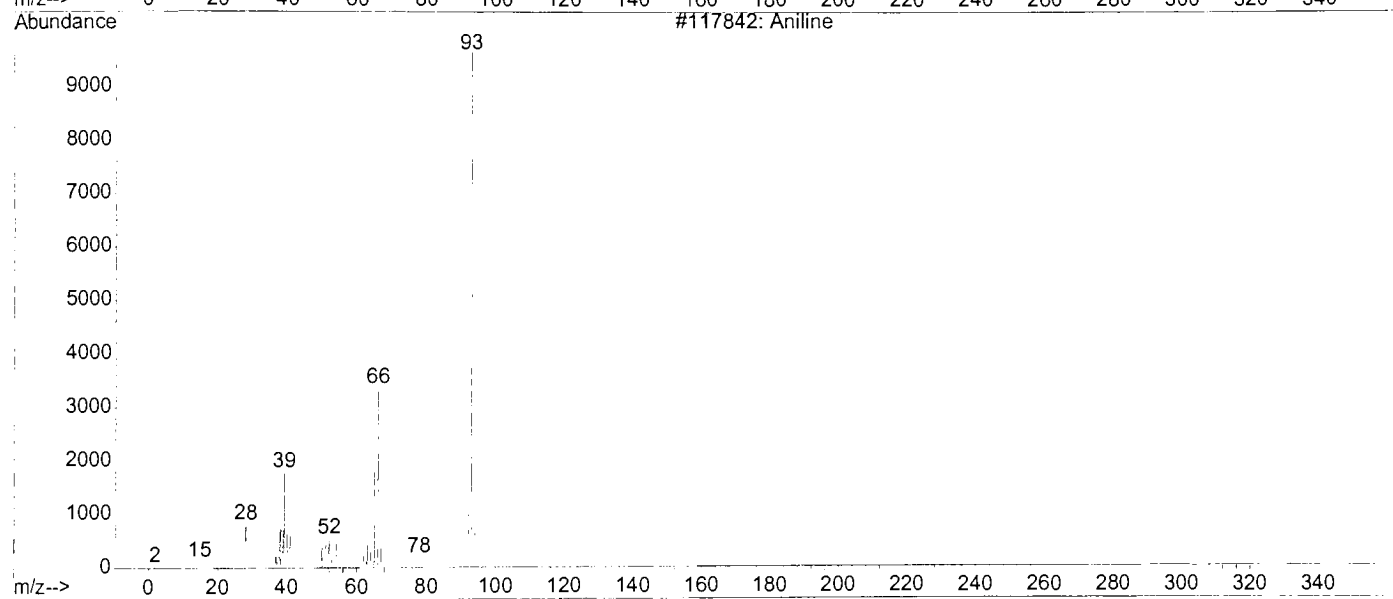
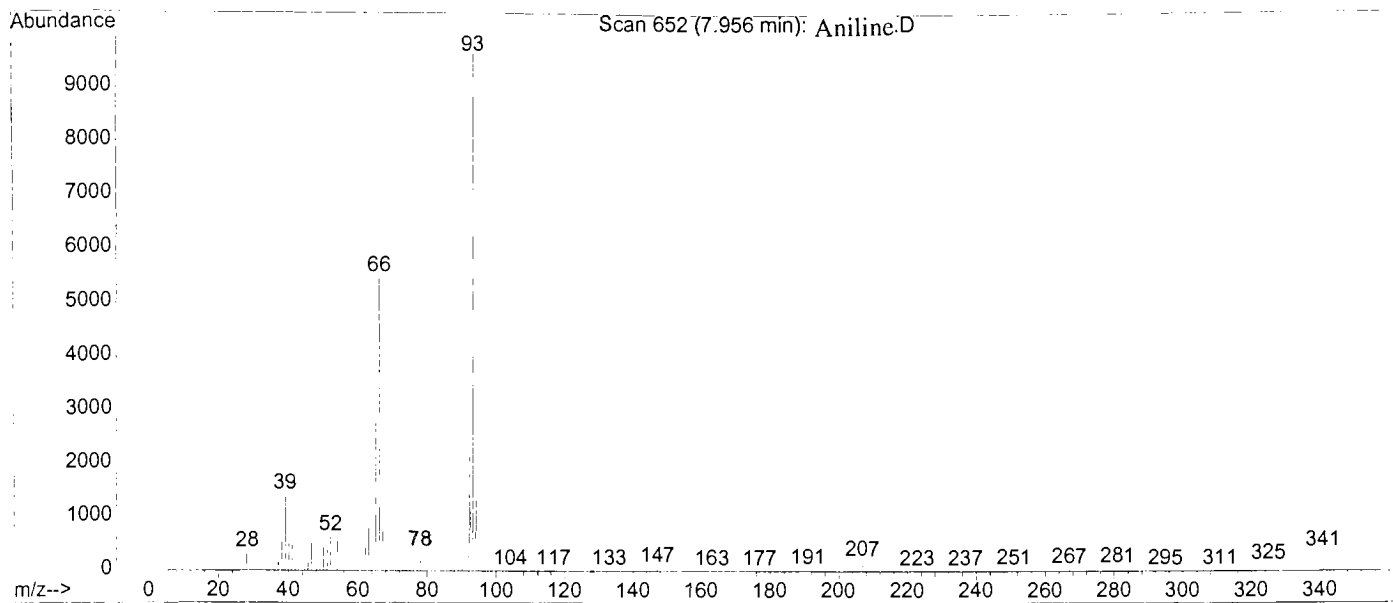


Library Searched : C:\DATABASE\NIST98.L

B1

Quality : 93

ID : Aniline MS Spectrum 1.

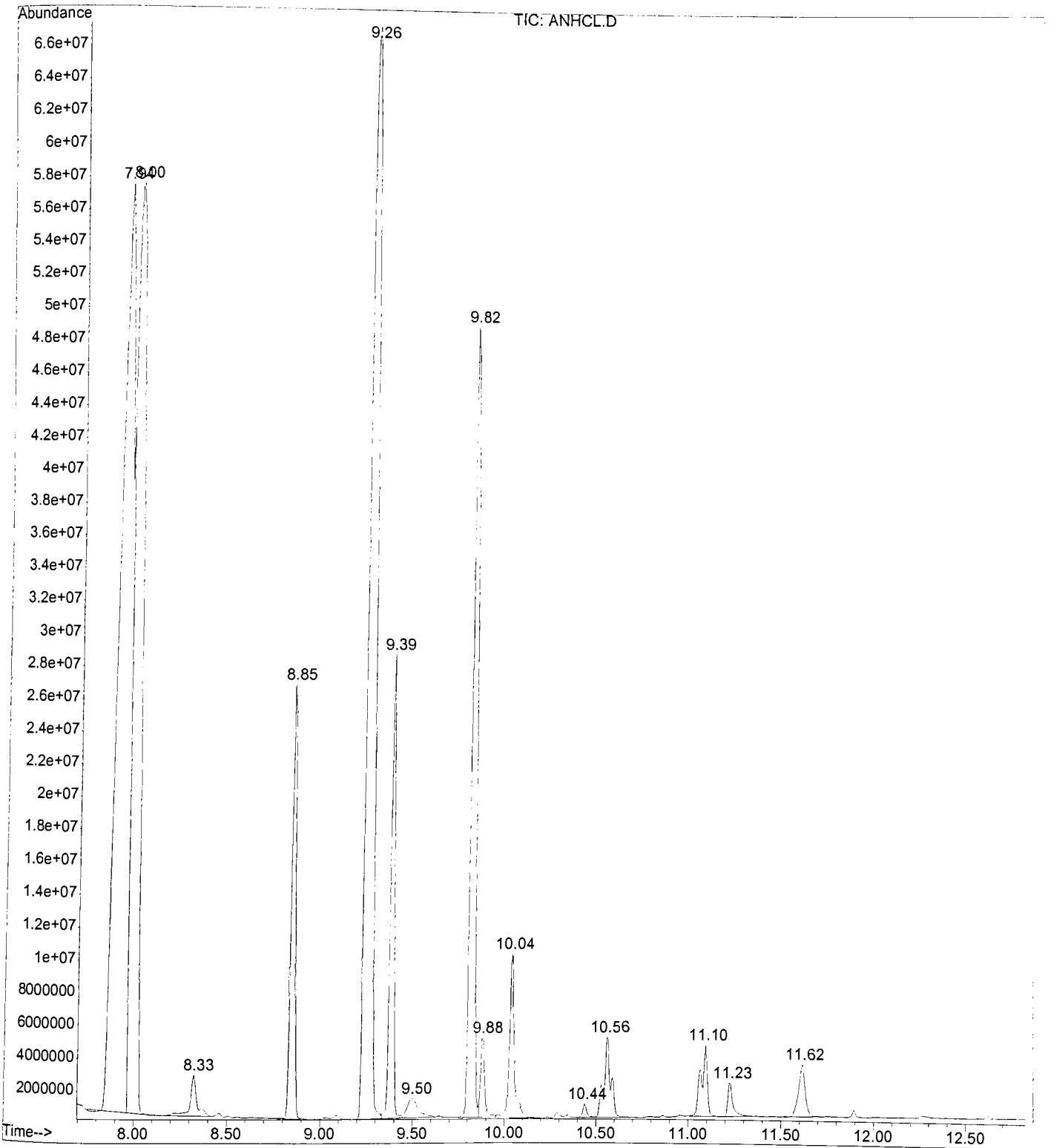


GC Spectrum 2. Area Percent Report

Data File : C:\HPCHEM\1\DATA\ANHCL.D
Acq On : 13 Jul 2000 16:13
Sample :
Misc :

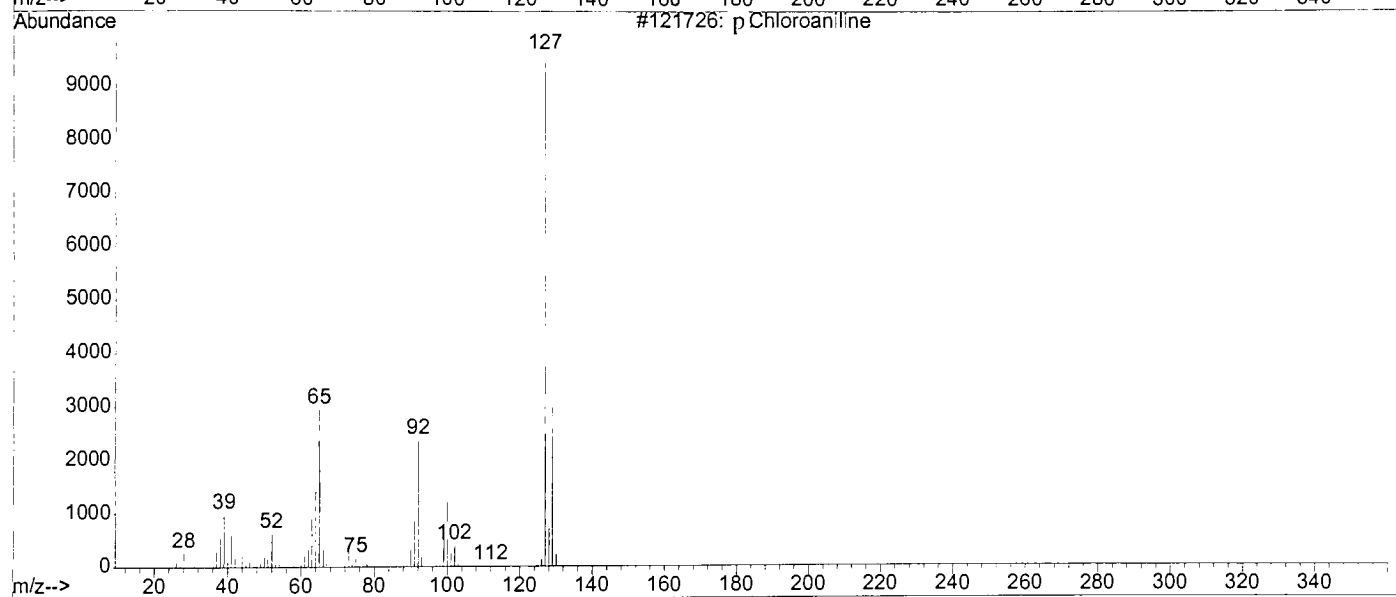
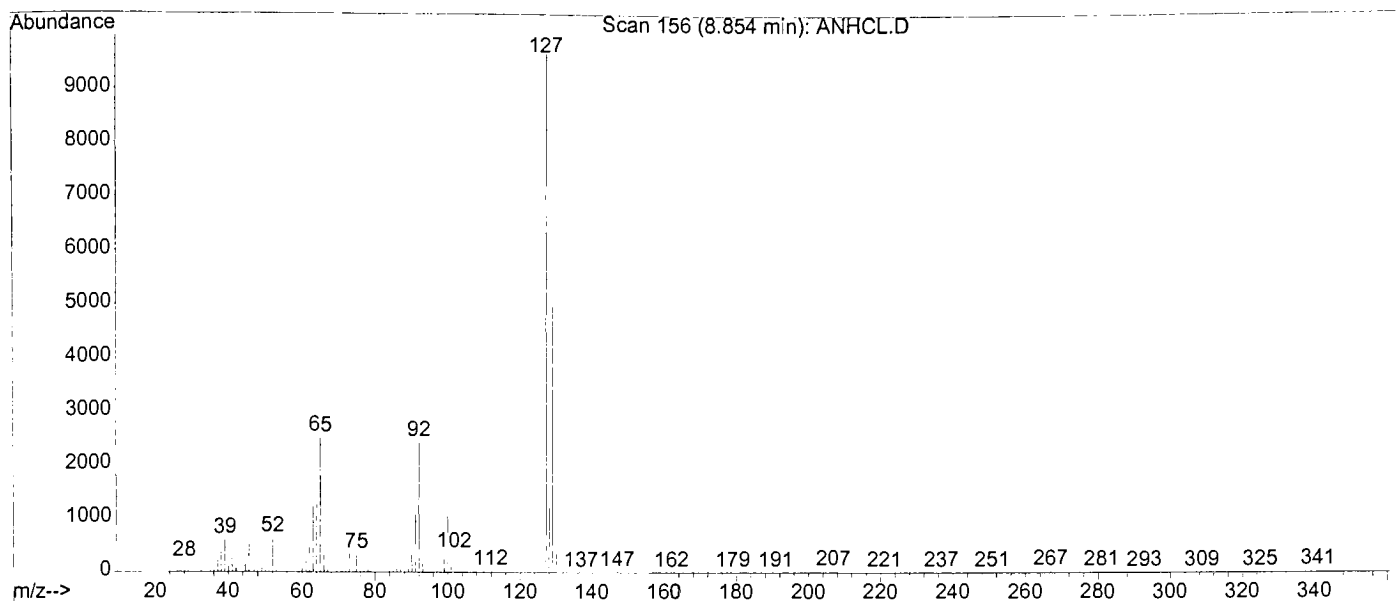
Vial: 1 B2
Operator:
Inst : RIT Chemi
Multiplr: 1.00
Sample Amount: 0.00

MS Integration Params: autoint1.e
Method : C:\HPCHEM\1\5973\PANKAJ7.M (Chemstation Integrator)
Title :



Library Searched : C:\DATABASE\NIST98.L
Quality : 93
ID : p-Chloroaniline MS Spectrum 2.

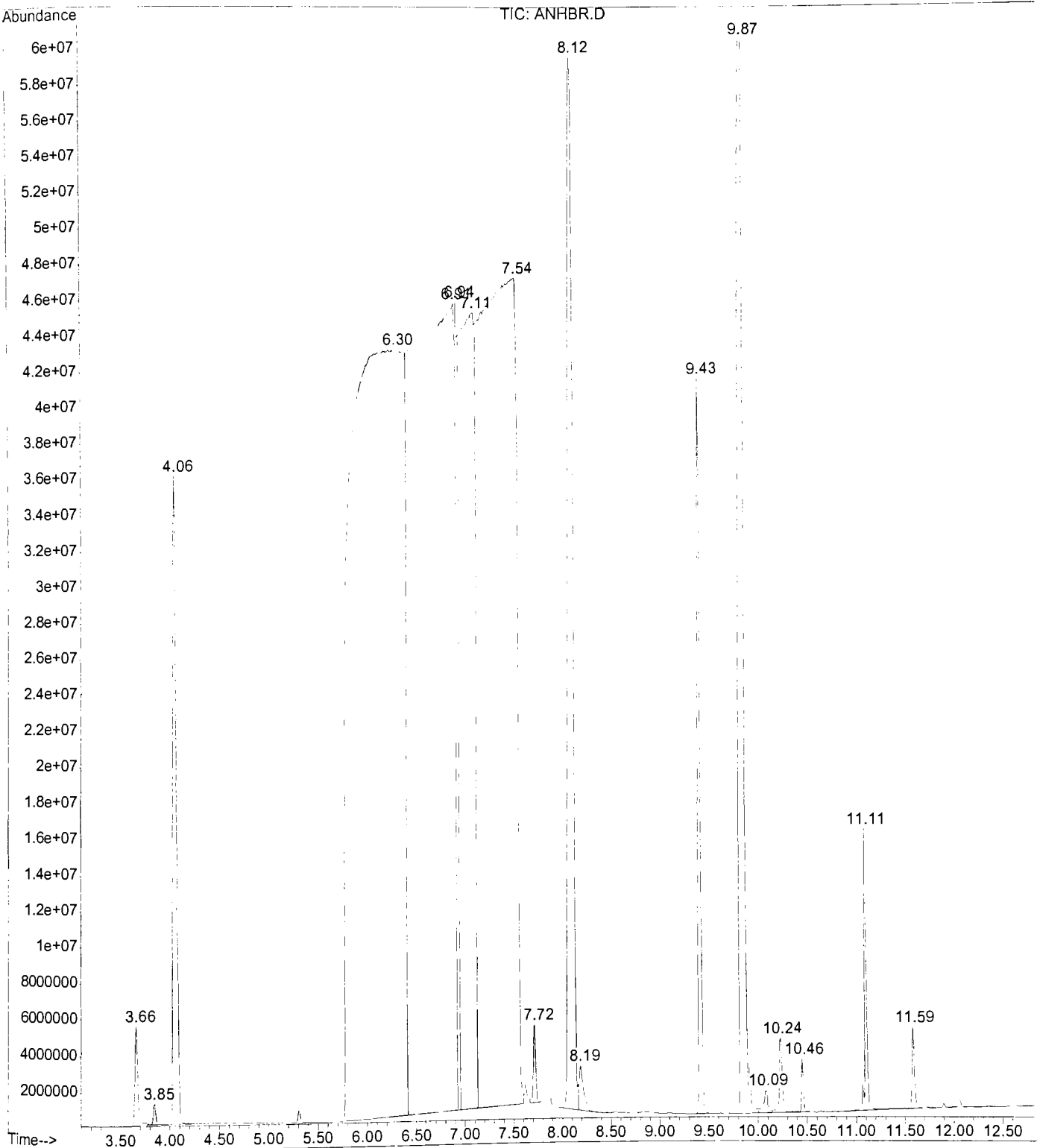
B2



Data File : C:\HPCHEM\1\DATA\ANHBR.D
Acq On : 13 Jul 2000 15:35
Sample :
Misc :

Vial: 1
Operator:
Inst : RIT Chemi
Multiplr: 1.00
Sample Amount: 0.00

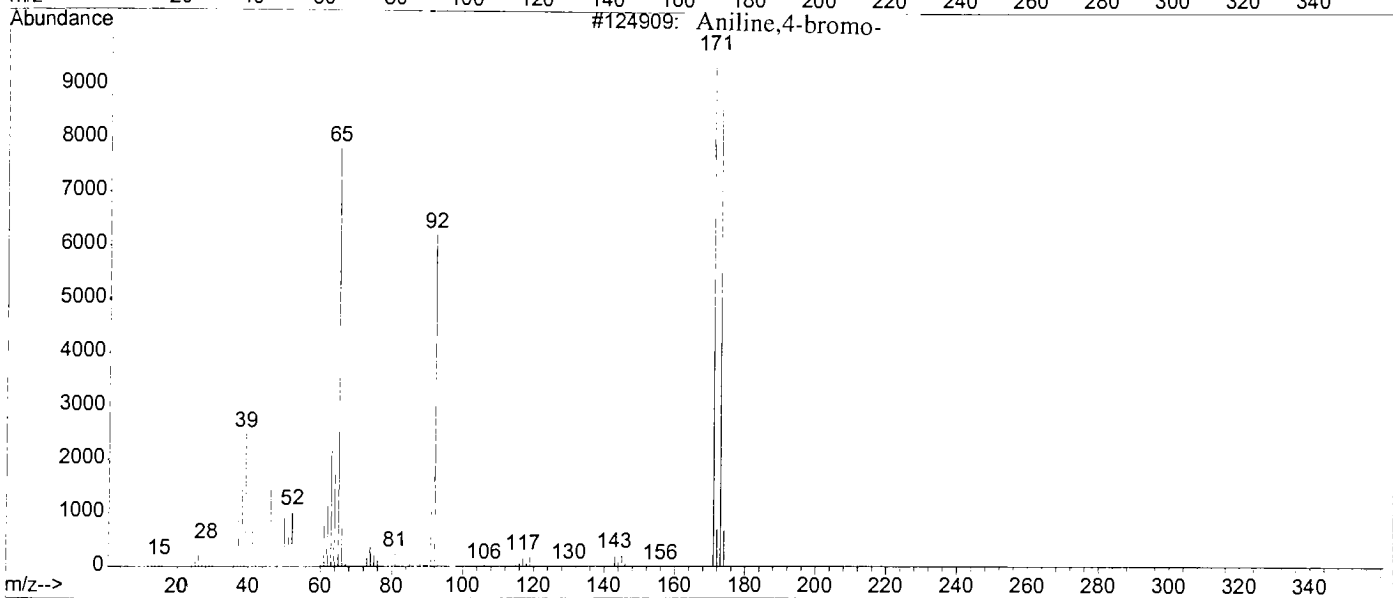
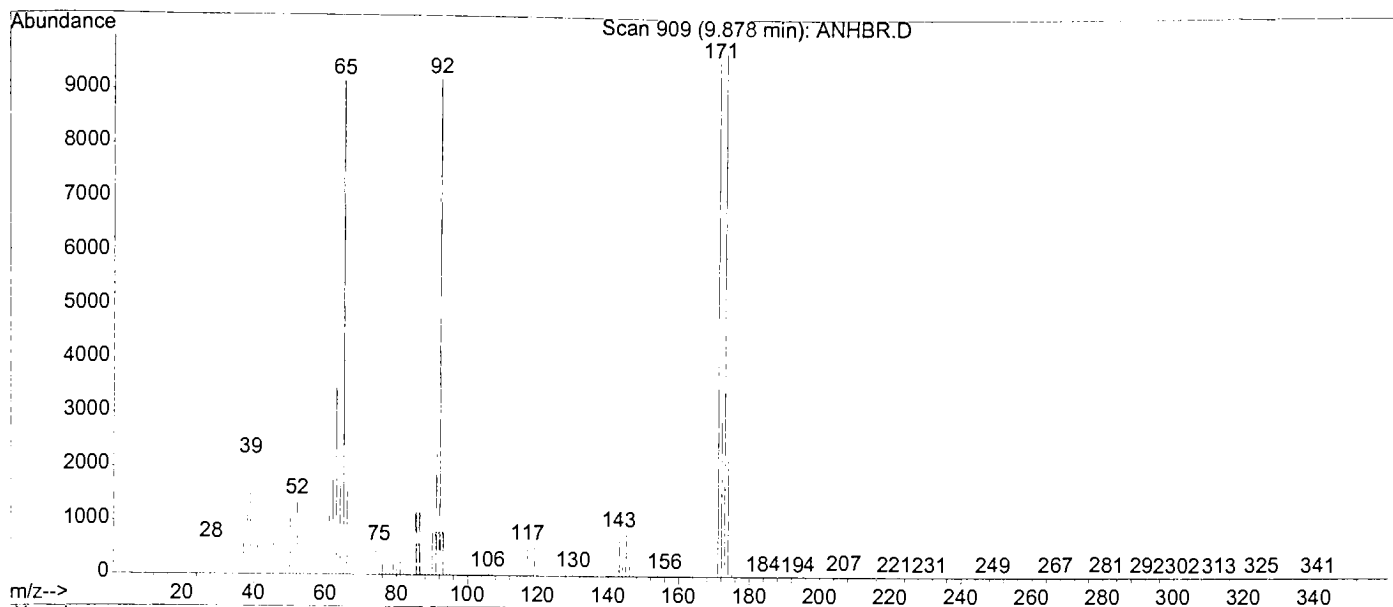
MS Integration Params: autoint1.e
Method : C:\HPCHEM\1\5973\PANKAJ.M (Chemstation Integrator)
Title :



Library Searched : C:\DATABASE\NIST98.L
Quality : 94
ID : Aniline, 4 - bromo-

B3

MS Spectrum 3.



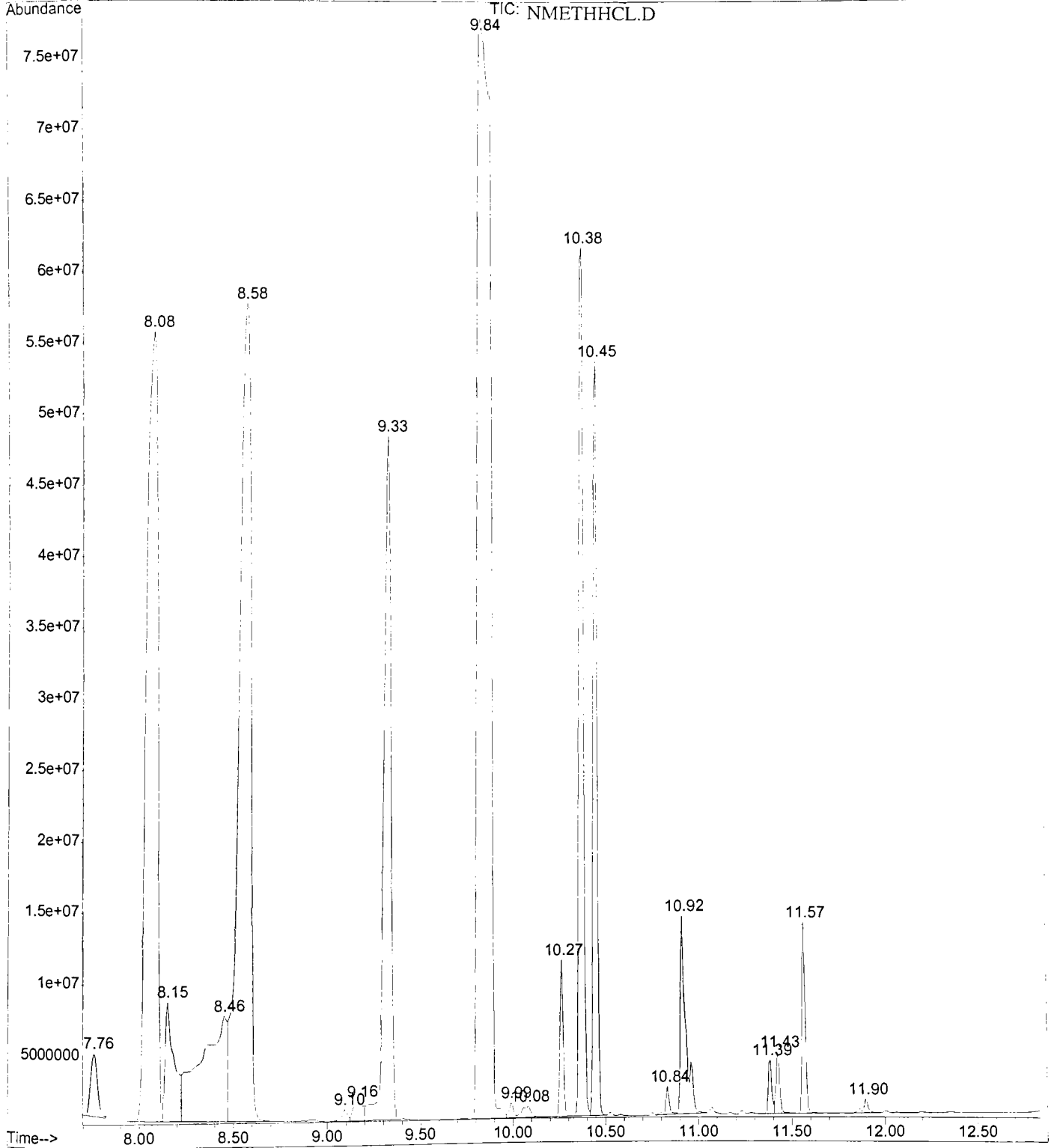
GC Spectrum 4. Area Percent Report

B4

Data File : C:\HPCHEM\1\DATA\NMETHHCL.D
Acq On : 13 Jul 2000 17:54
Sample :
Misc :

Vial: 1
Operator:
Inst : RIT Chemi
Multiplr: 1.00
Sample Amount: 0.00

MS Integration Params: autoint1.e
Method : C:\HPCHEM\1\5973\PANKAJ7.M (Chemstation Integrator)
Title :

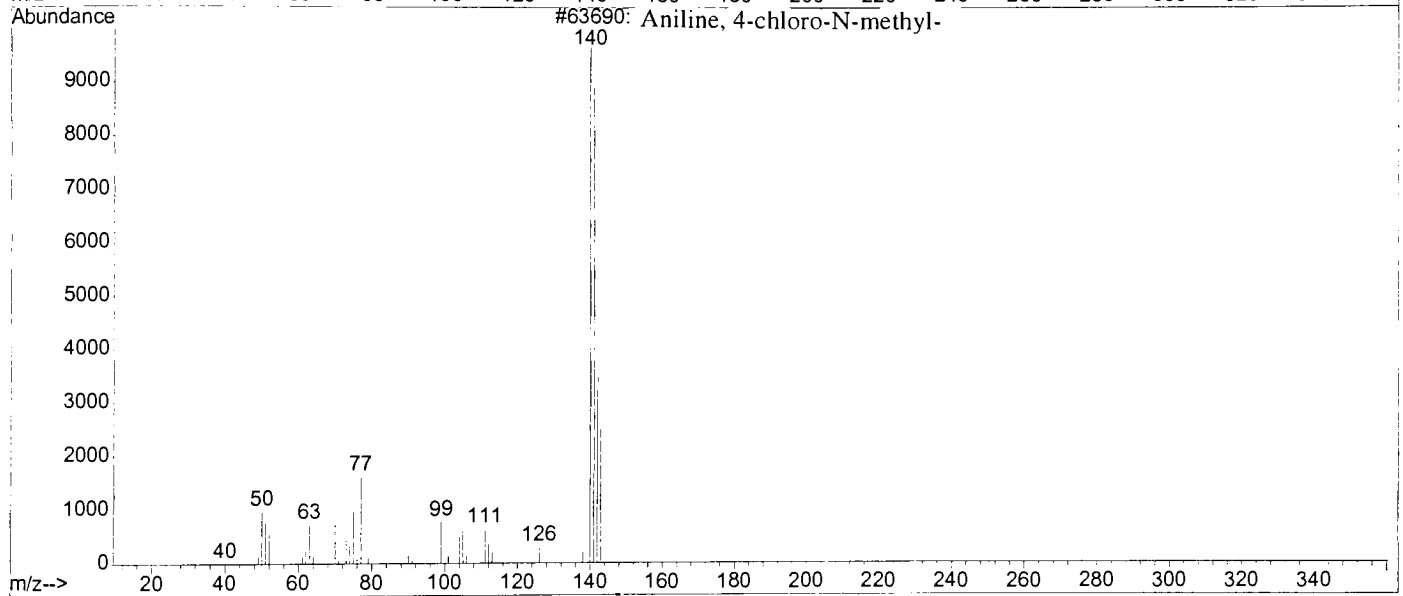
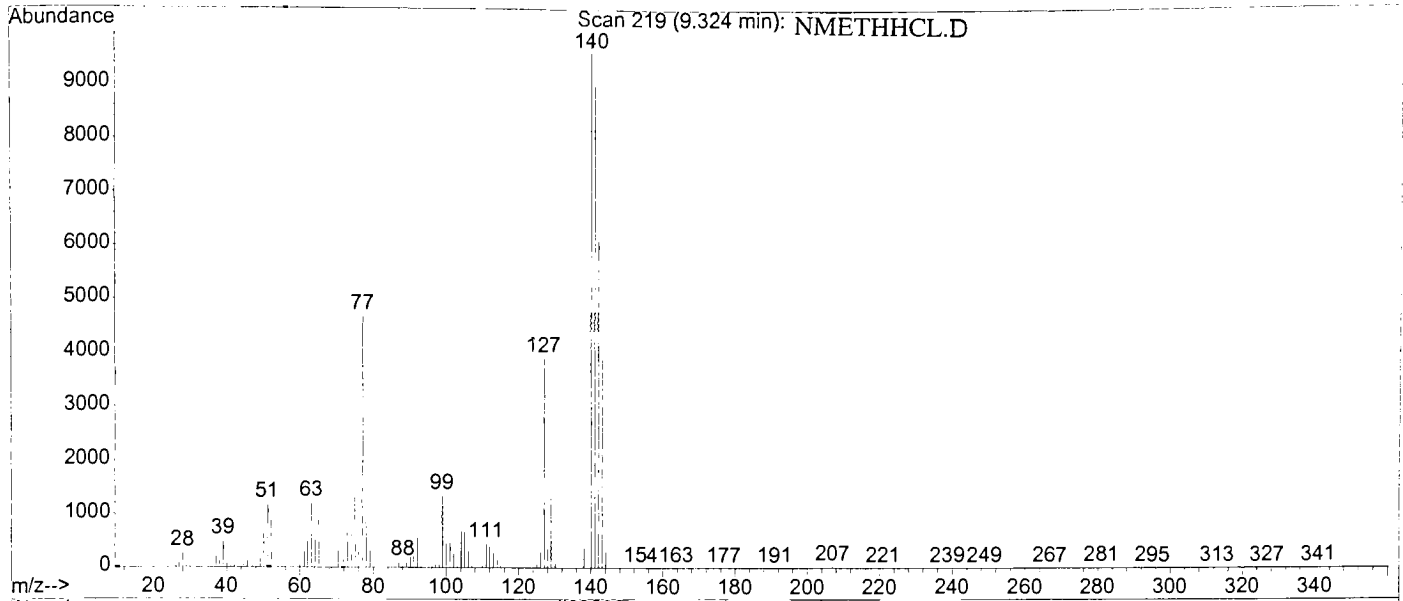


Library Searched : C:\DATABASE\NIST98.L

Quality : 60

B4

ID : Aniline, 4-chloro-N-methyl- MS Spectrum 4.



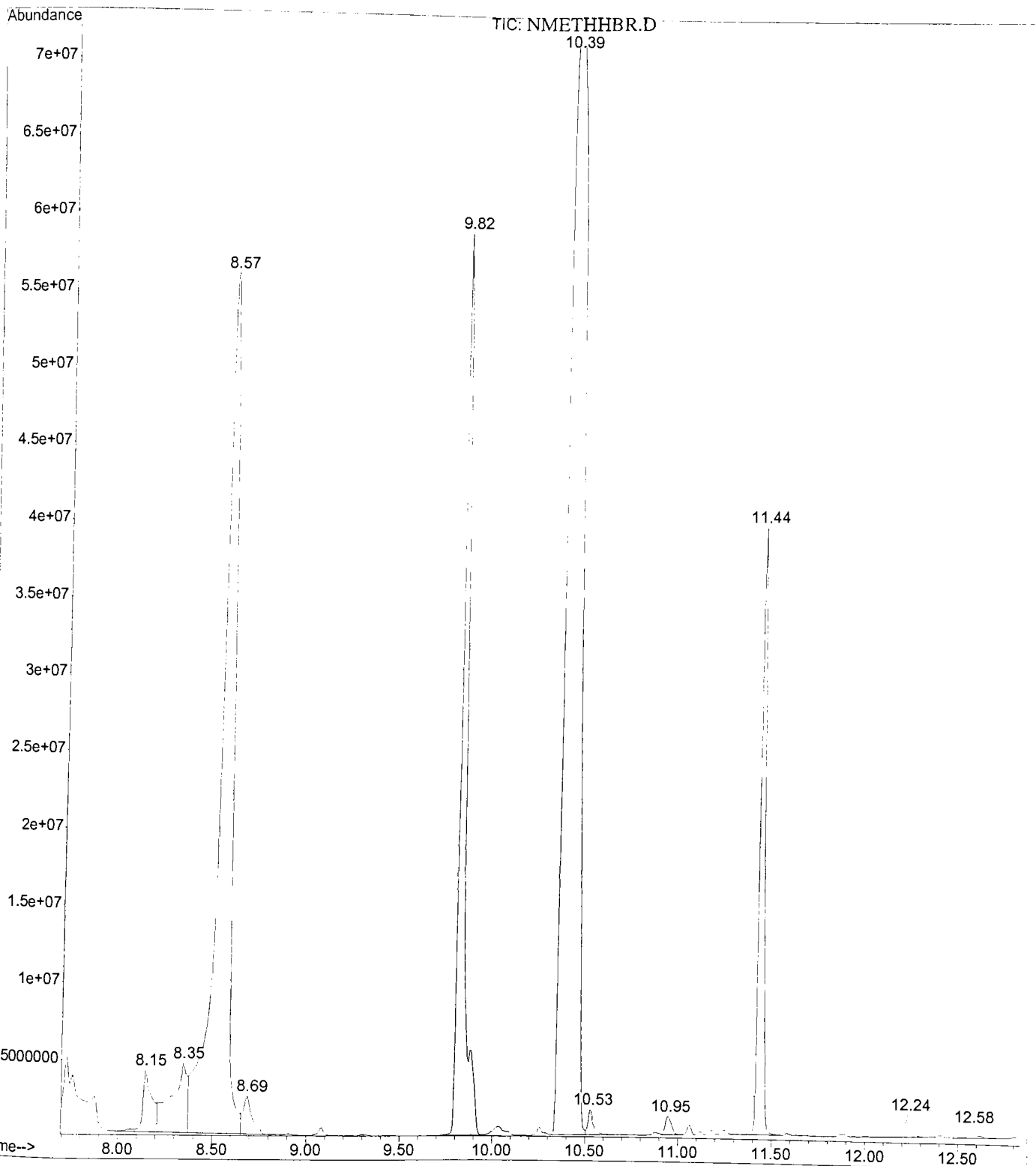
GC Spectrum 5. Area Percent Report

B5

Data File : C:\HPCHEM\1\DATA\NMETHHBR.D
 Acq On : 13 Jul 2000 18:40
 Sample :
 Misc :

Vial: 1
 Operator:
 Inst : RIT Chemi
 Multiplr: 1.00
 Sample Amount: 0.00

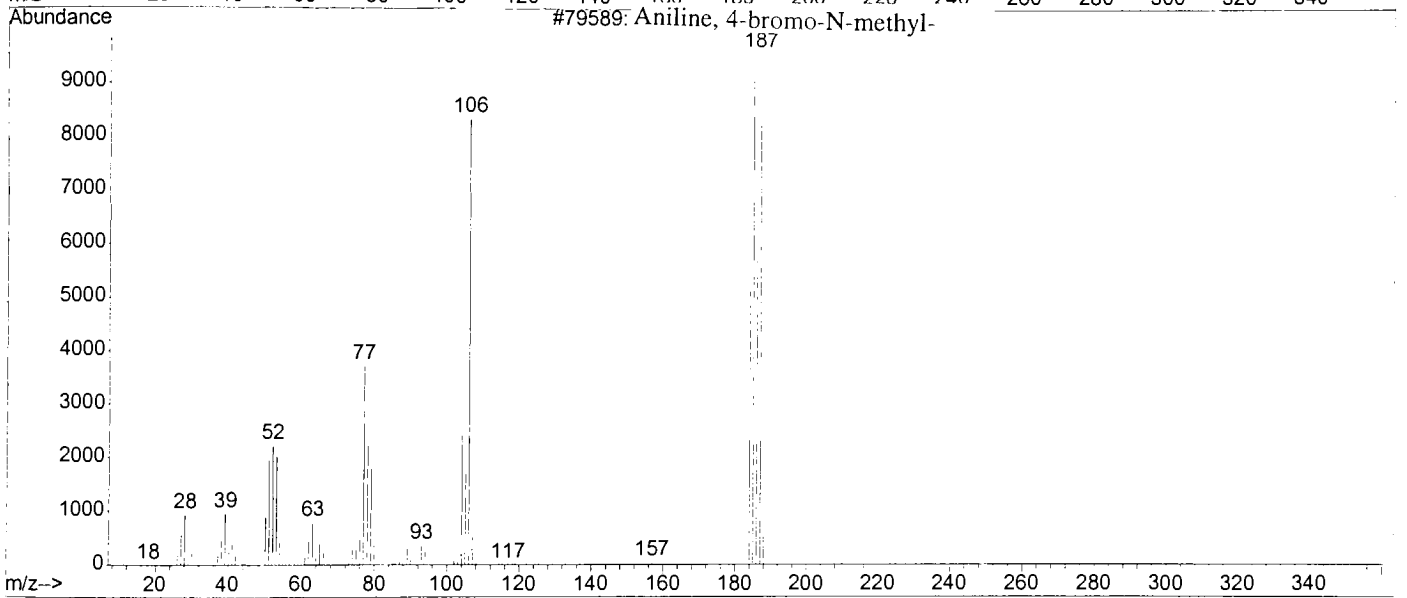
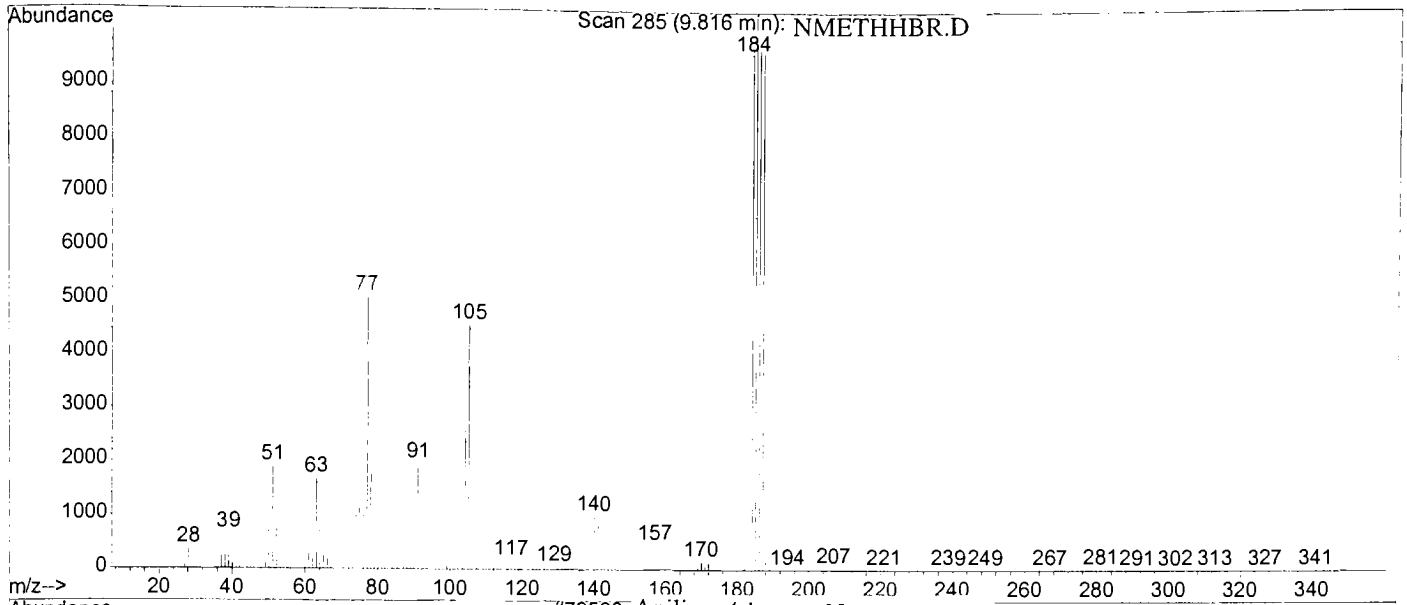
MS Integration Params: autoint1.e
 Method : C:\HPCHEM\1\5973\PANKAJ7.M (Chemstation Integrator)
 Title :



Library Searched : C:\DATABASE\NIST98.L
Quality : 47
ID : Aniline, 4-bromo-N-methyl-

B5

MS Spectrum 5.



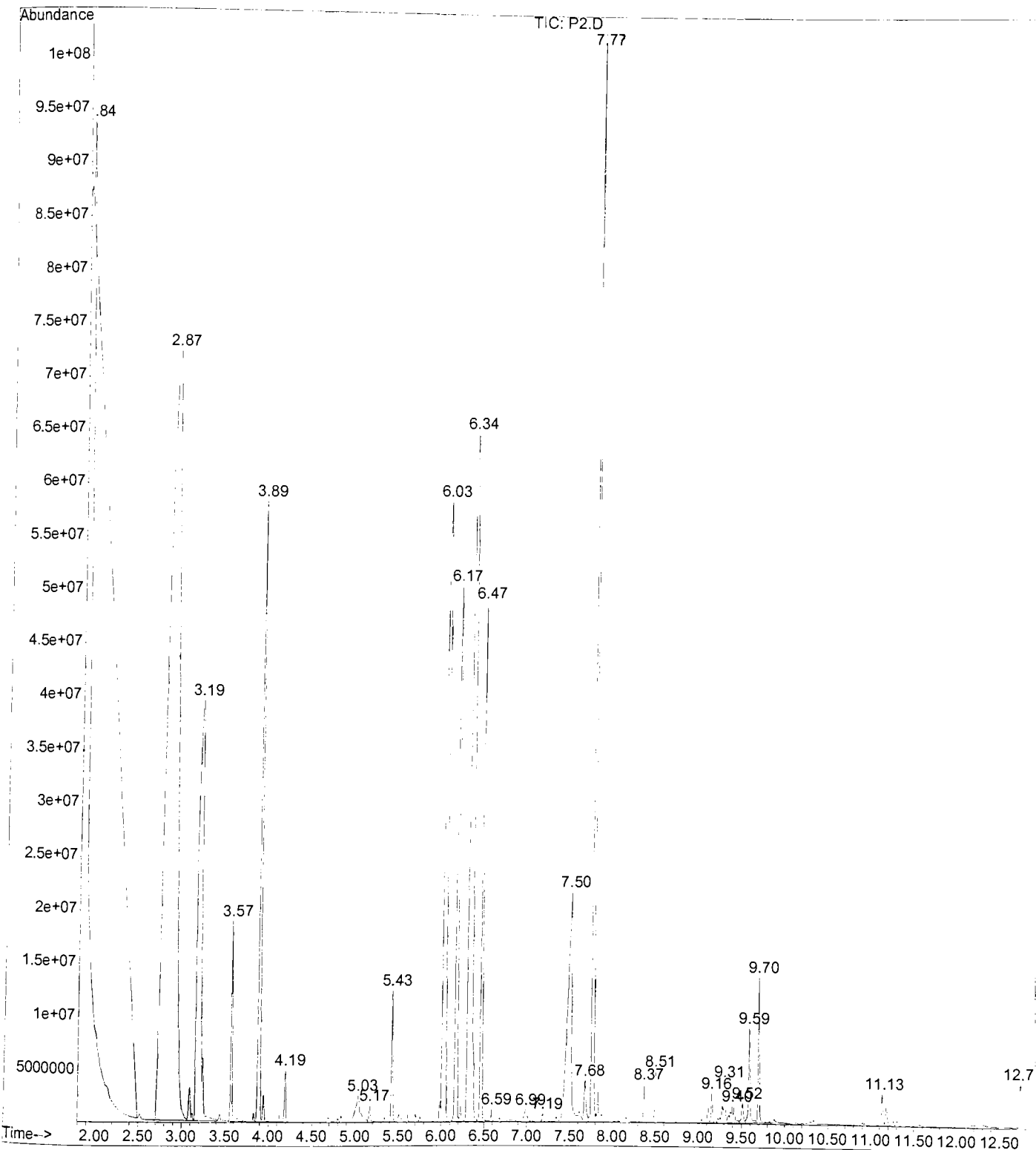
GC Spectrum 6. Area Percent Report

B6

Data File : C:\HPCHEM\1\DATA\P2.D
 Acq On : 4 Jan 2000 14:47
 Sample : Aniline, 4- bromo- N,N - dimethyl-
 Misc :

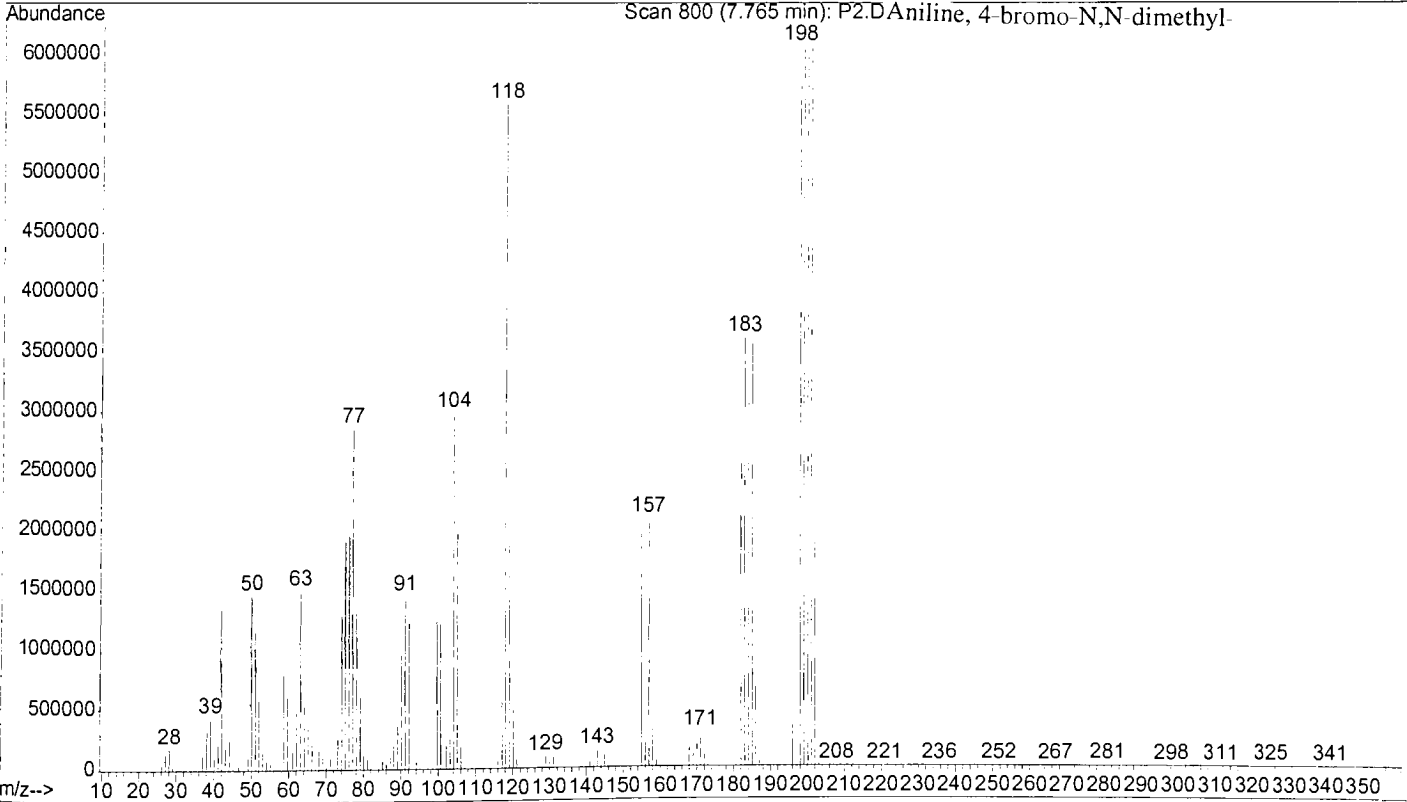
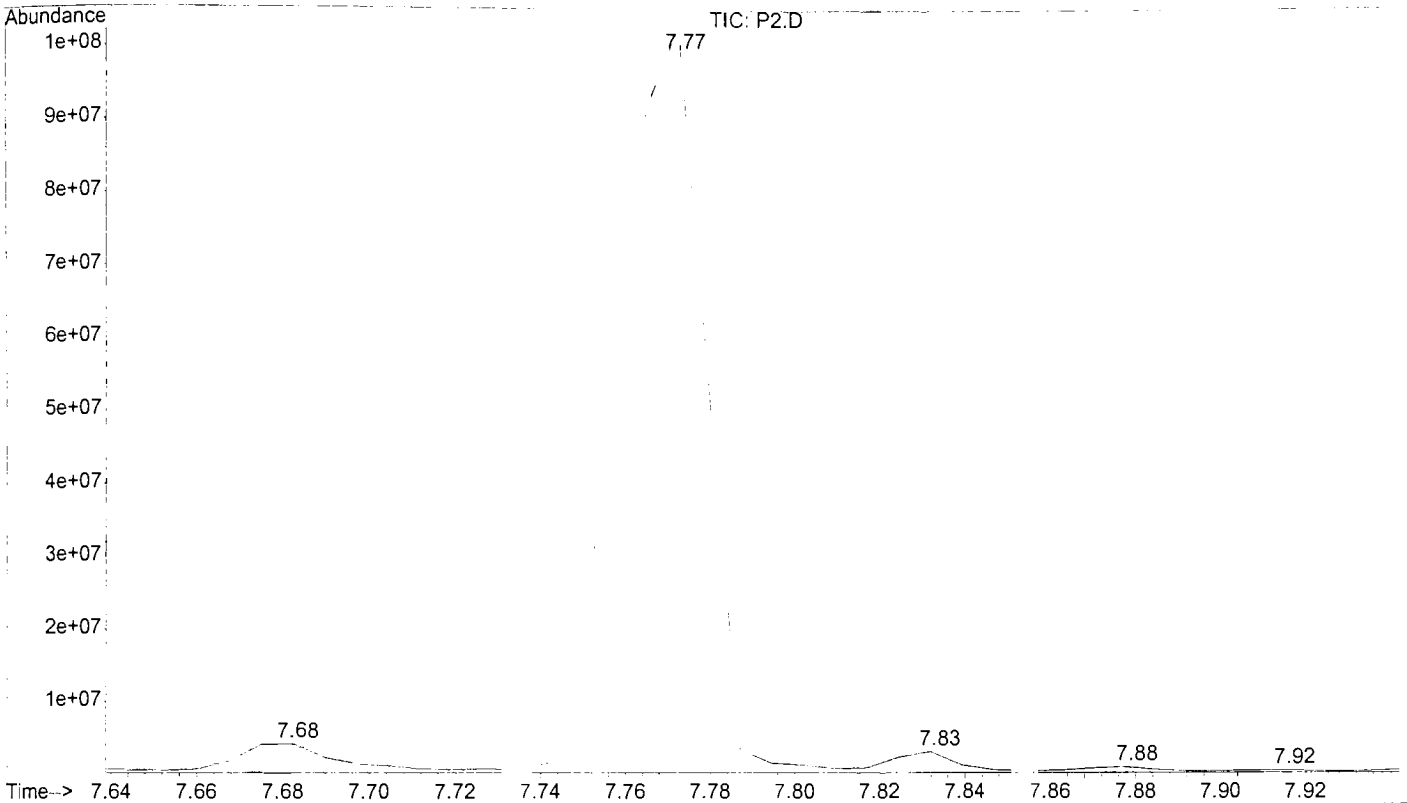
Vial: 1
 Operator: Pankaj
 Inst : RIT Chemi
 Multiplr: 1.00
 Sample Amount: 0.00

MS Integration Params: autoint1.e
 Method : C:\HPCHEM\1\5973\PANKAJ.M (Chemstation Integrator)
 Title :



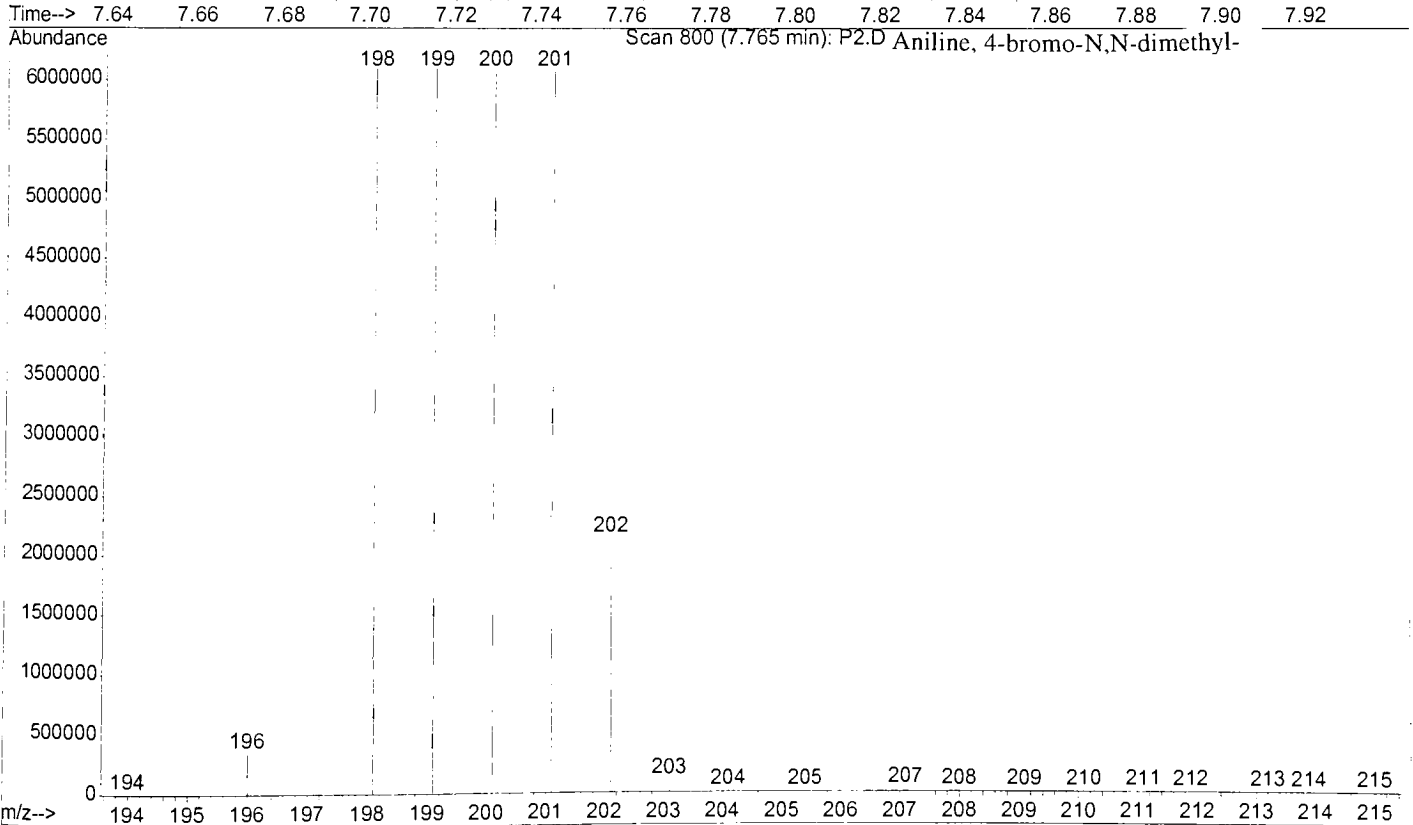
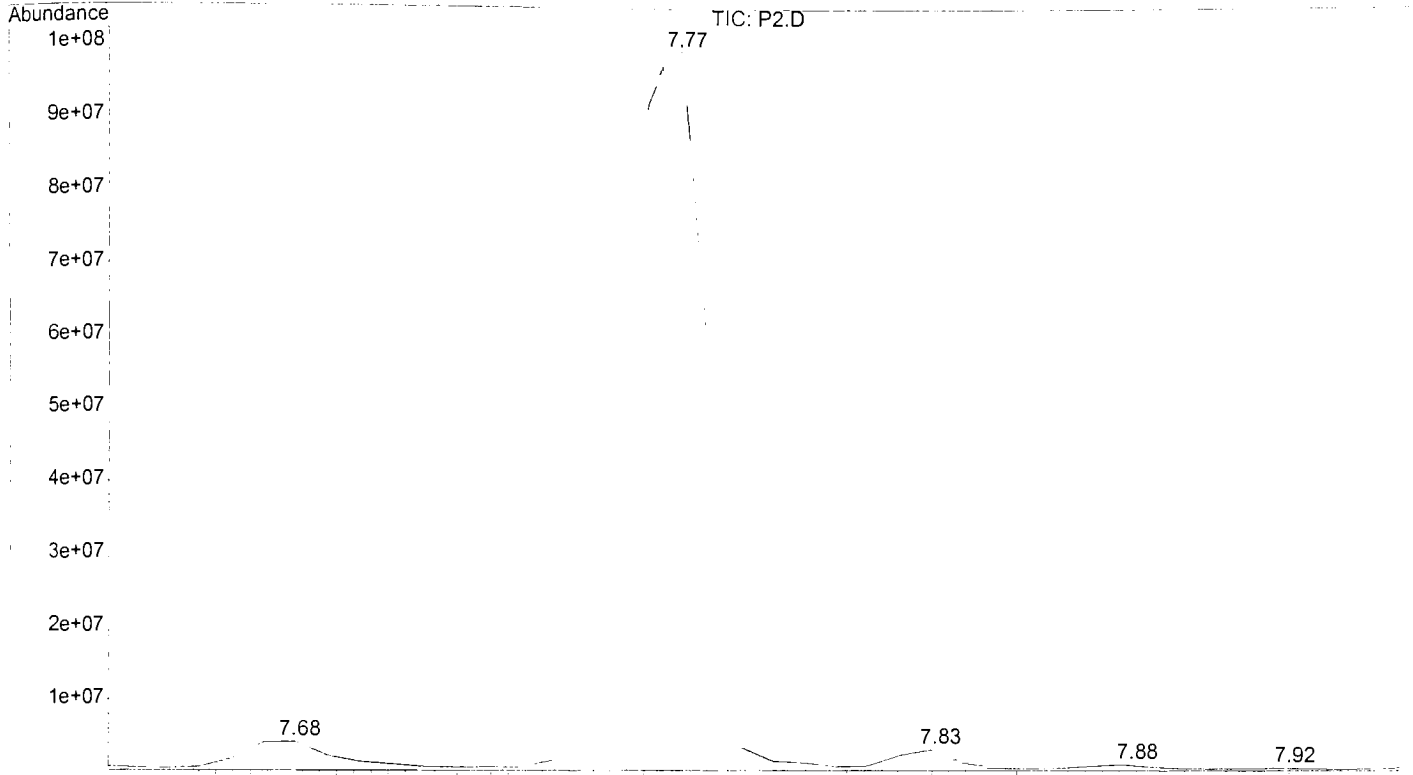
File : C:\HPCHEM\1\DATA\P2.D
Operator : Pankaj
Acquired : 4 Jan 2000 14:47 using AcqMethod PANKAJ
Instrument : RIT Chemi
Sample Name: Aniline, 4-bromo-N,N-dimethyl- MS Spectrum 6.
Scan Info :
Scan Number: 1

B6



File : C:\HPCHEM\1\DATA\P2.D
Operator : Pankaj
Acquired : 4 Jan 2000 14:47 using AcqMethod PANKAJ
Instrument : RIT Chemi
Sample Name: Aniline, 4-bromo- N,N - dimethyl- MS Spectrum 6.
Scan Info :
Scan Number: 1

B6



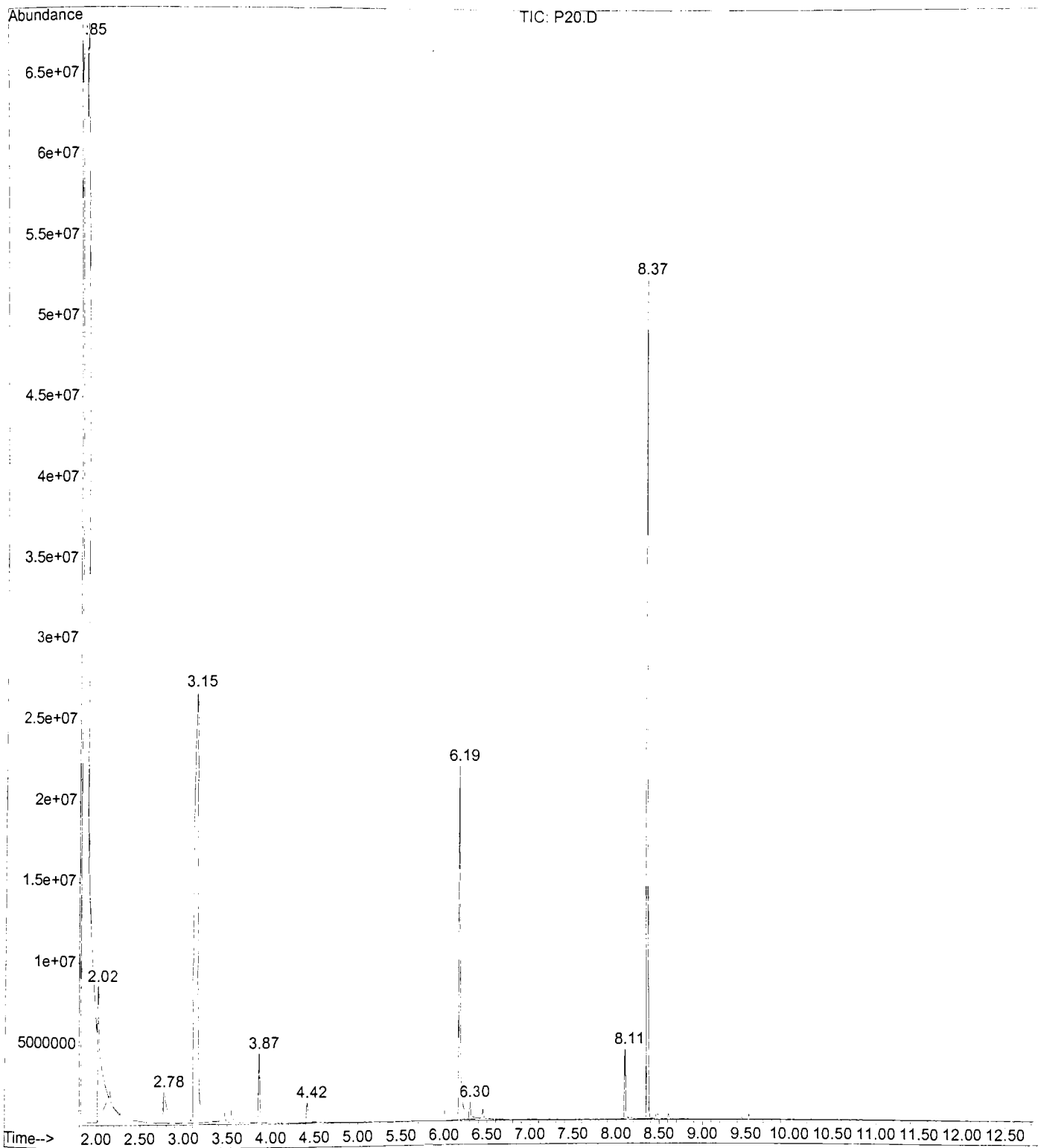
GC Spectrum 7. Area Percent Report

B7

Data File : C:\HPCHEM\1\DATA\P20.D
Acq On : 9 Mar 2000 10:42
Sample : phenol+hcl
Misc :

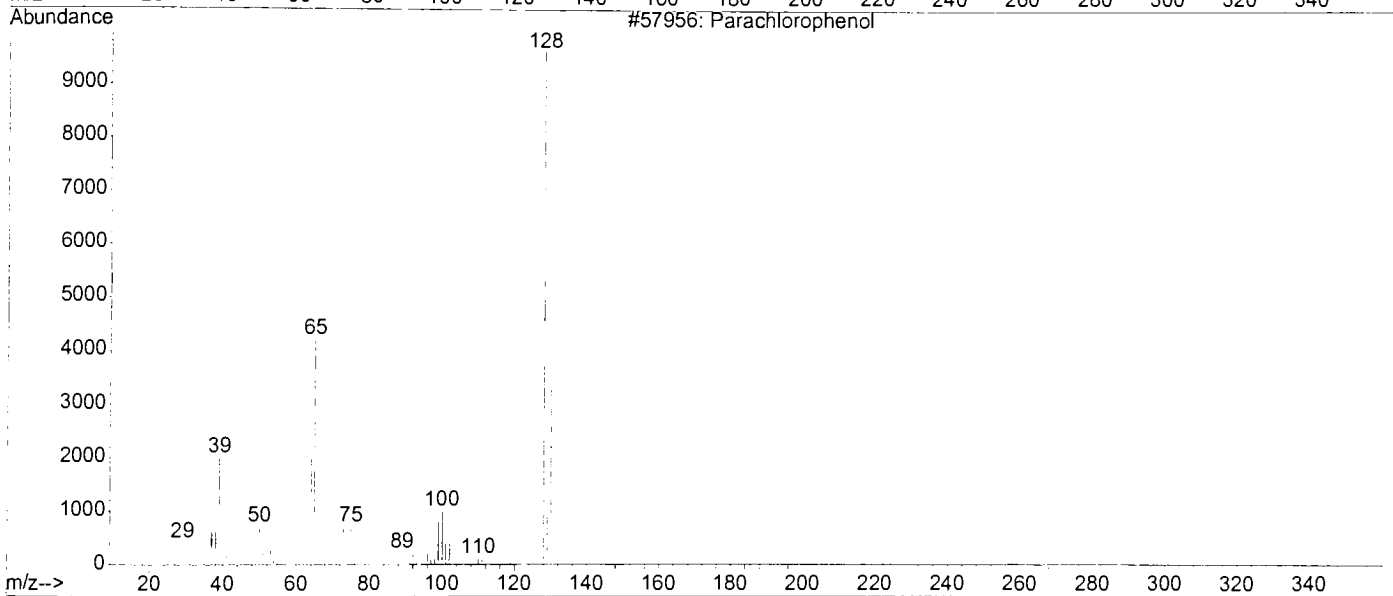
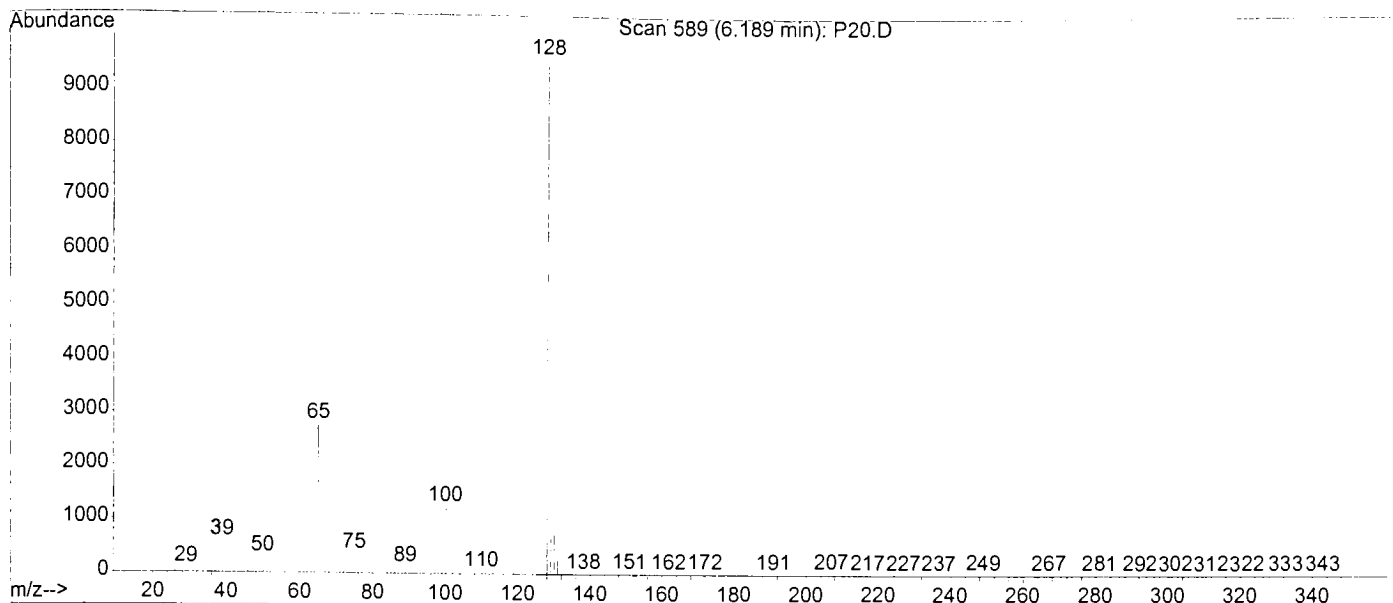
Vial: 1
Operator:
Inst : RIT Chemi
Multiplr: 1.00
Sample Amount: 0.00

MS Integration Params: autoint1.e
Method : C:\HPCHEM\1\5973\PANKAJ.M (Chemstation Integrator)
Title :



Library Searched : C:\DATABASE\NIST98.L
Quality : 94
ID : Parachlorophenol MS Spectrum 7.

B7



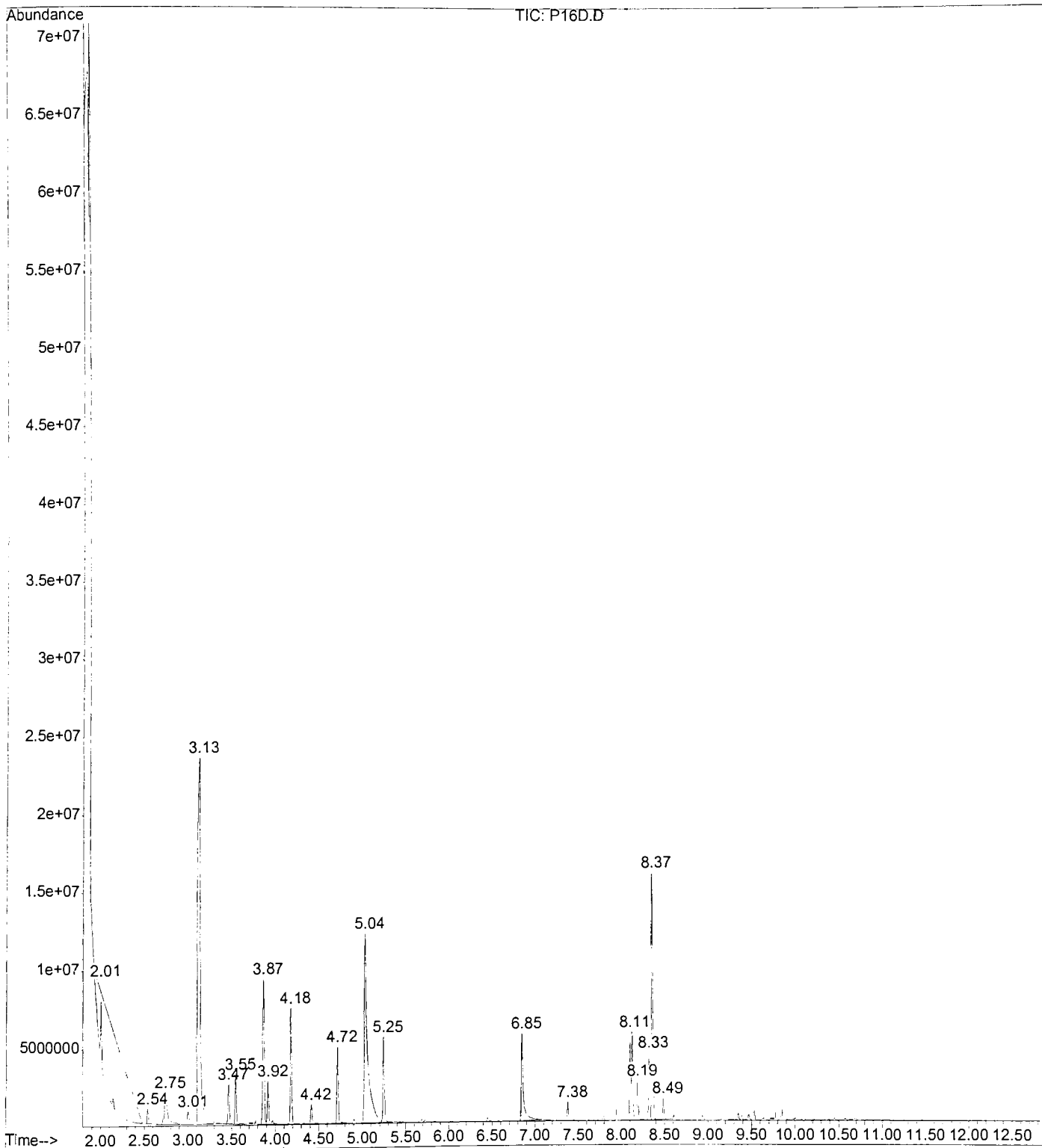
GC Spectrum 8. Area Percent Report

B8

Data File : C:\HPCHEM\1\DATA\P16D.D
 Acq On : 9 Mar 2000 11:12
 Sample : phenol+++hbr
 Misc :

Vial: 1
 Operator:
 Inst : RIT Chemi
 Multiplr: 1.00
 Sample Amount: 0.00

MS Integration Params: autoint1.e
 Method : C:\HPCHEM\1\5973\PANKAJ.M (Chemstation Integrator)
 Title :

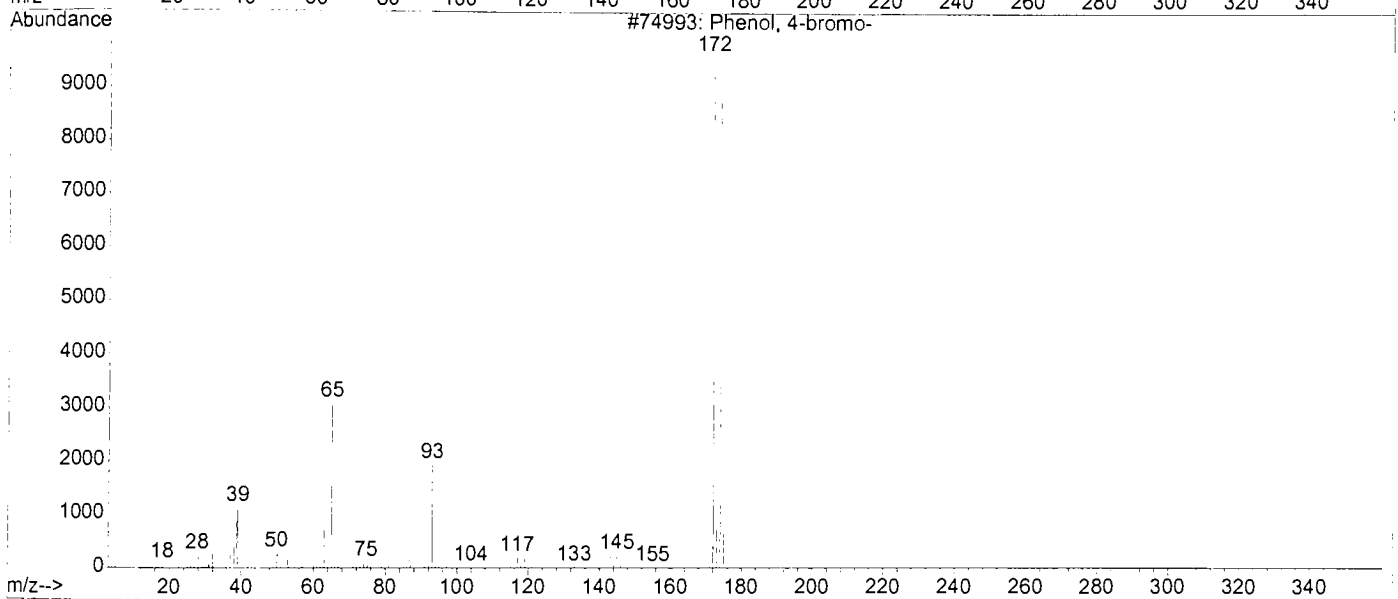
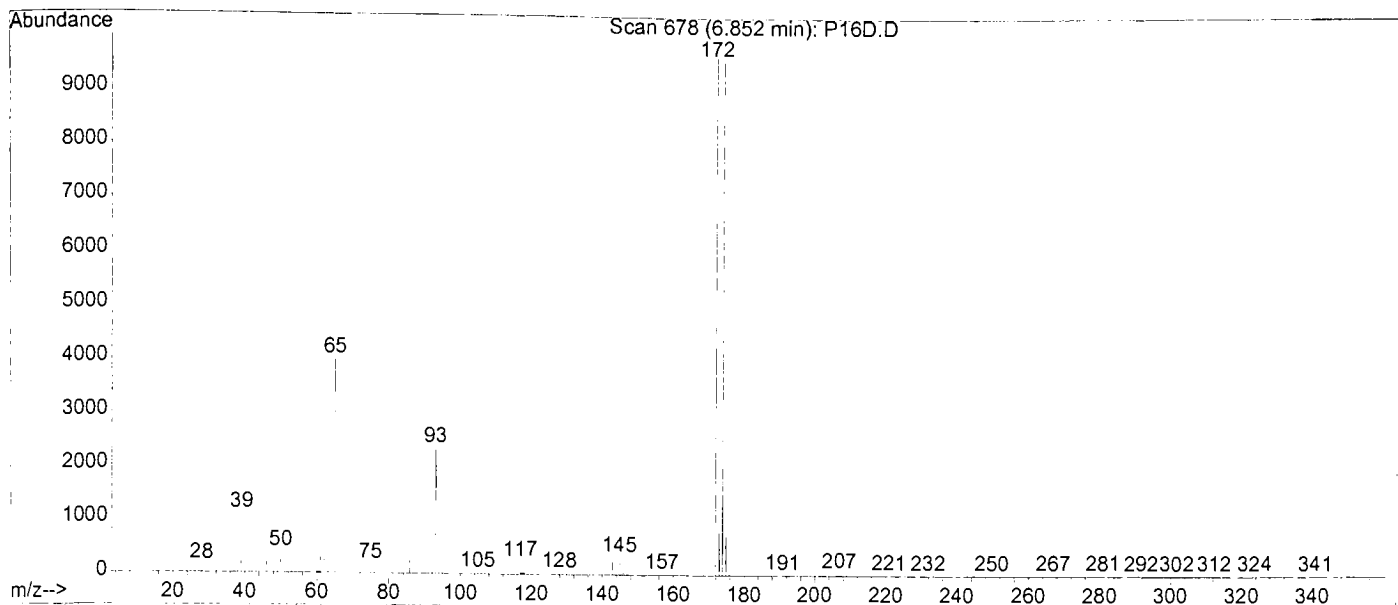


Library Searched : C:\DATABASE\NIST98.L

Quality : 94

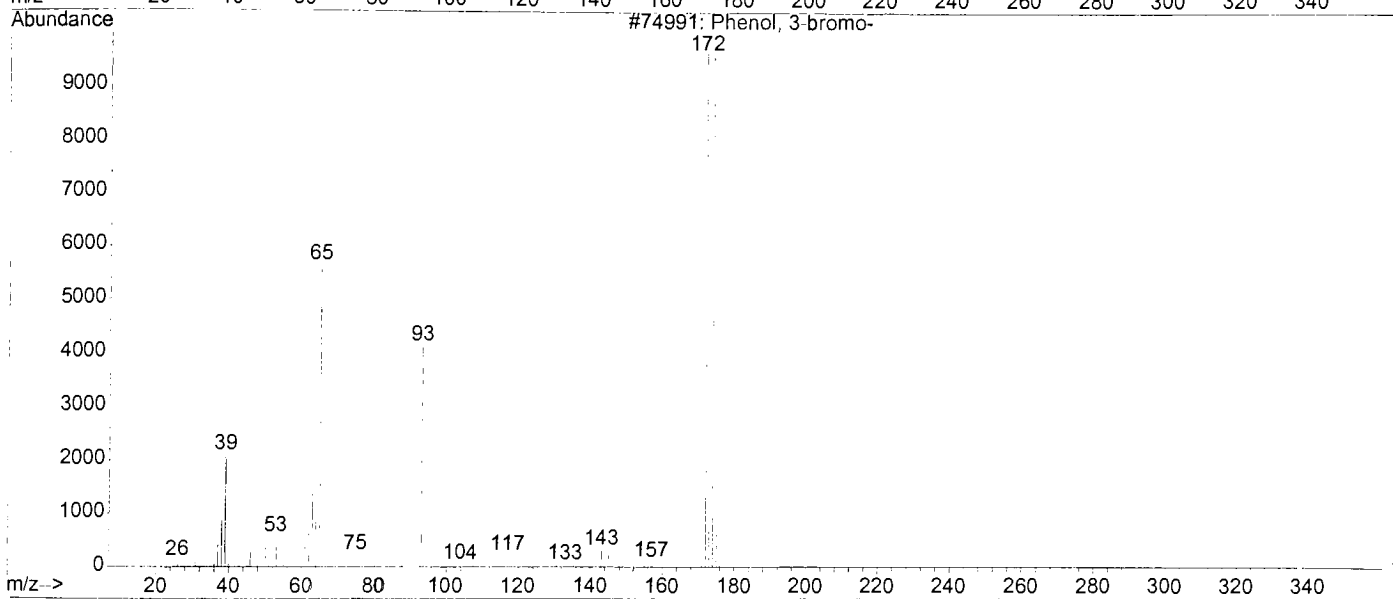
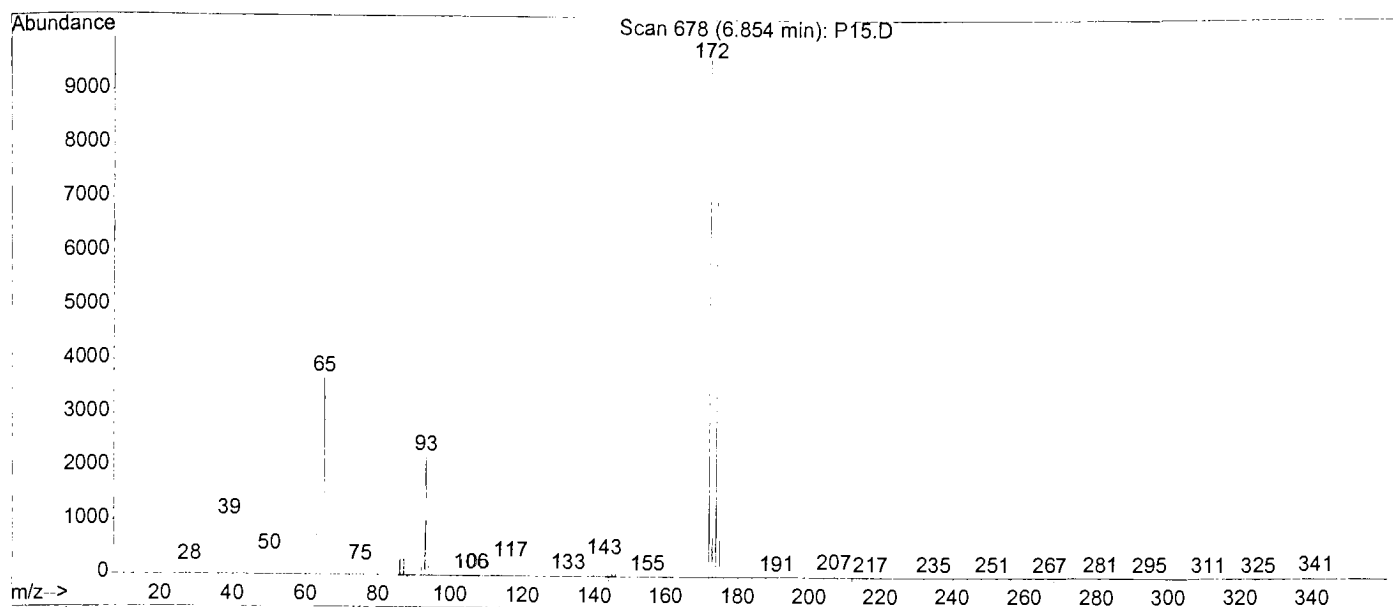
ID : Phenol, 4-bromo- MS Spectrum 8.

B8



Library Searched : C:\DATABASE\NIST98.L
Quality : 94
ID : Phenol, 3-bromo- MS Spectrum 8.

B8

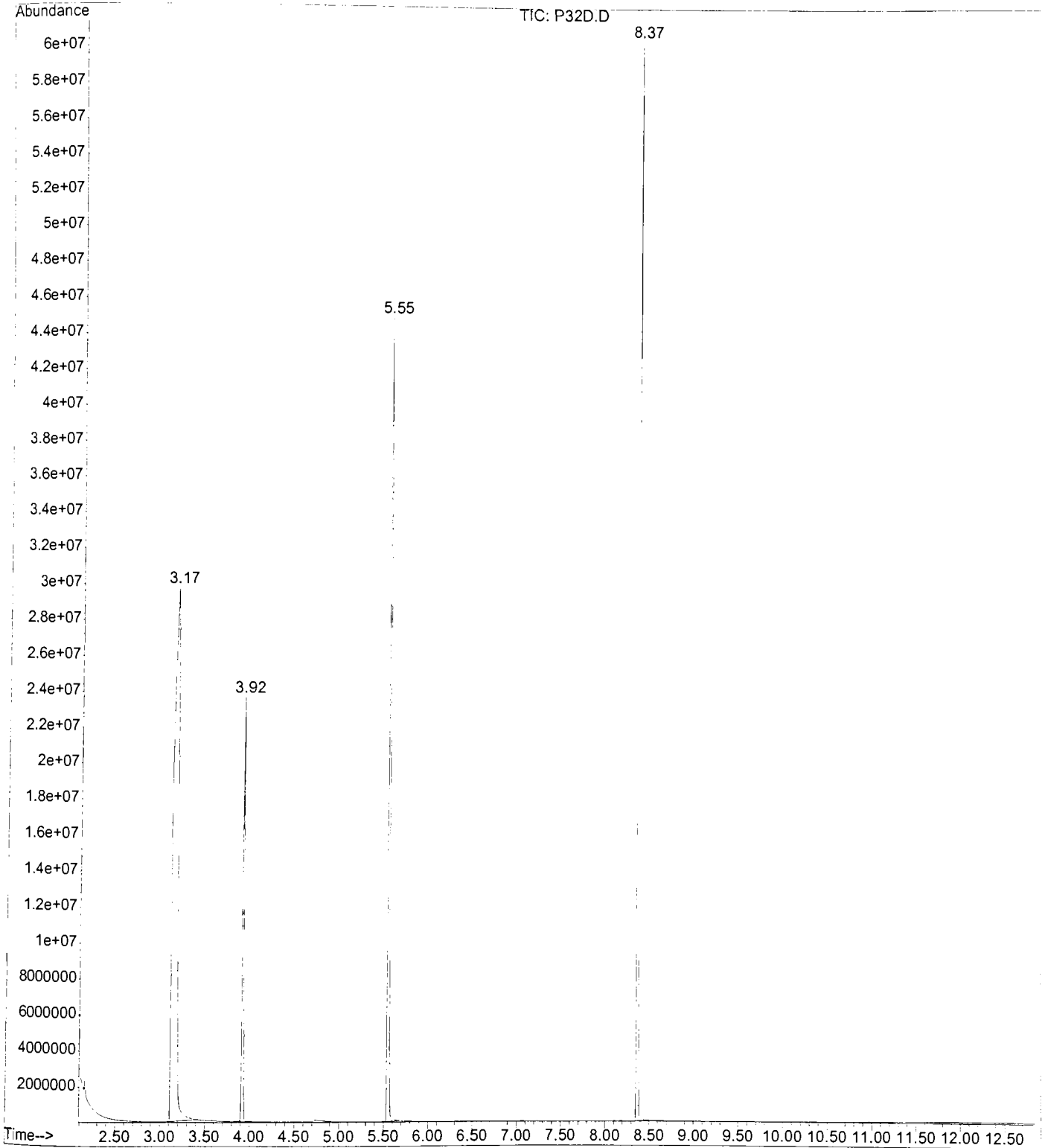


GC Spectrum 9. Area Percent Report

Data File : C:\HPCHEM\1\DATA\P32D.D
Acq On : 9 Mar 2000 15:23
Sample : anisole++hcl no.2
Misc :

Vial: 1 B9
Operator:
Inst : RIT Chemi
Multiplr: 1.00
Sample Amount: 0.00

MS Integration Params: autoint1.e
Method : C:\HPCHEM\1\5973\PANKAJ.M (Chemstation Integrator)
Title :



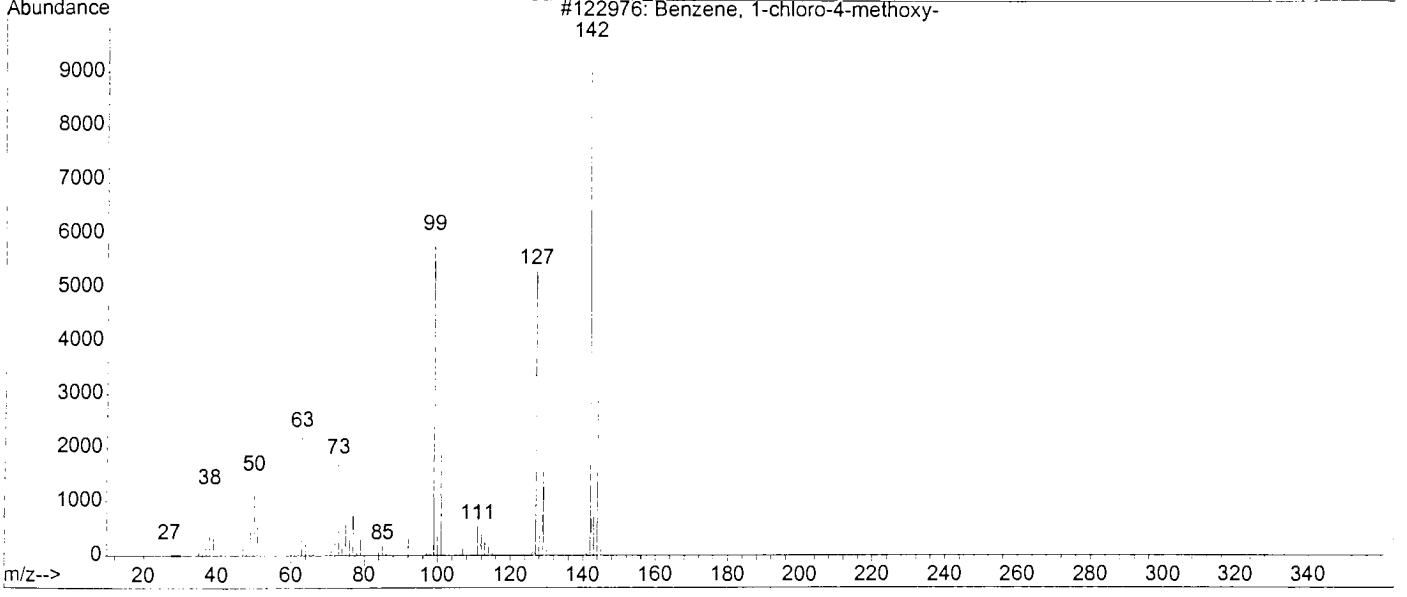
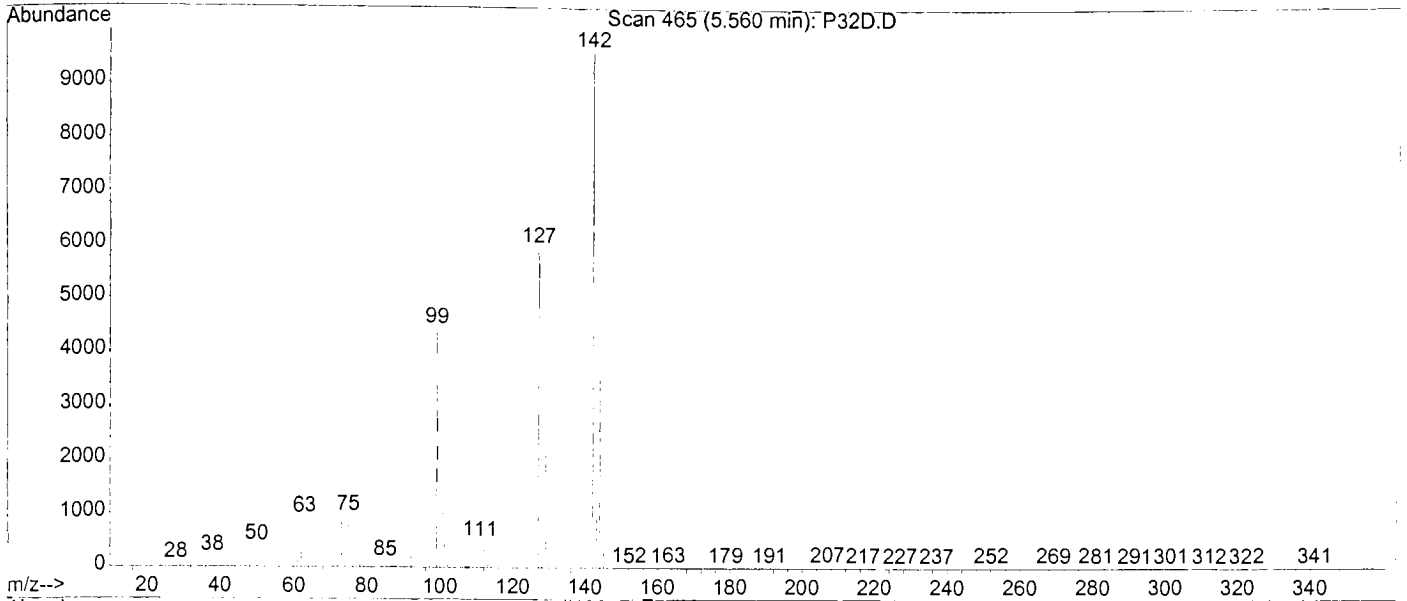
Library Searched : C:\DATABASE\NIST98.L

Quality : 95

ID : Benzene, 1-chloro-4-methoxy-

MS Spectrum 9.

B9

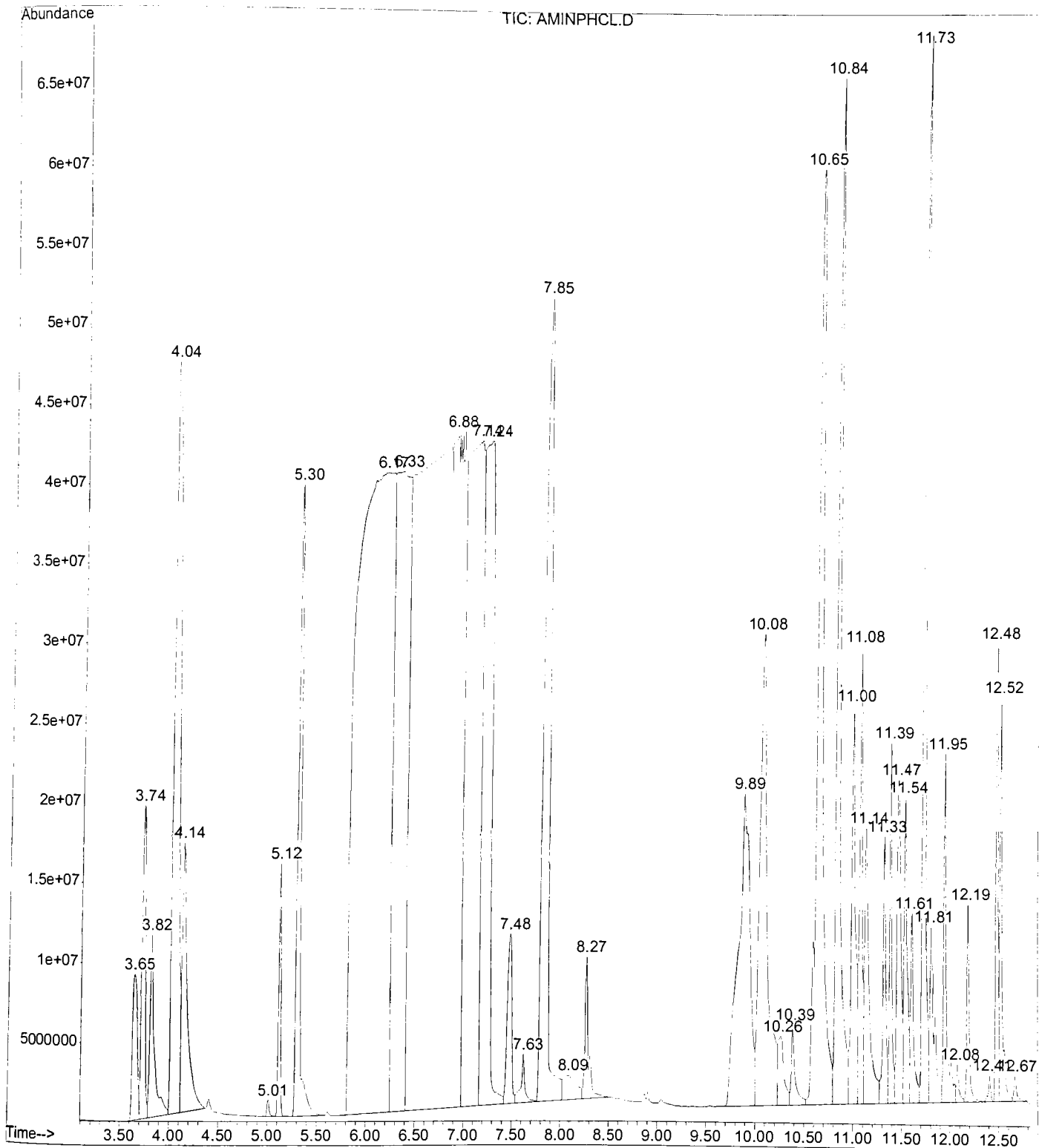


GC Spectrum 10. Area Percent Report

Data File : C:\HPCHEM\1\DATA\AMINPHCL.D
 Acq On : 13 Jul 2000 20:10
 Sample :
 Misc :

Vial: 1 B10
 Operator:
 Inst : RIT Chemi
 Multiplr: 1.00
 Sample Amount: 0.00

MS Integration Params: autoint1.e
 Method : C:\HPCHEM\1\5973\PANKAJ.M (Chemstation Integrator)
 Title :



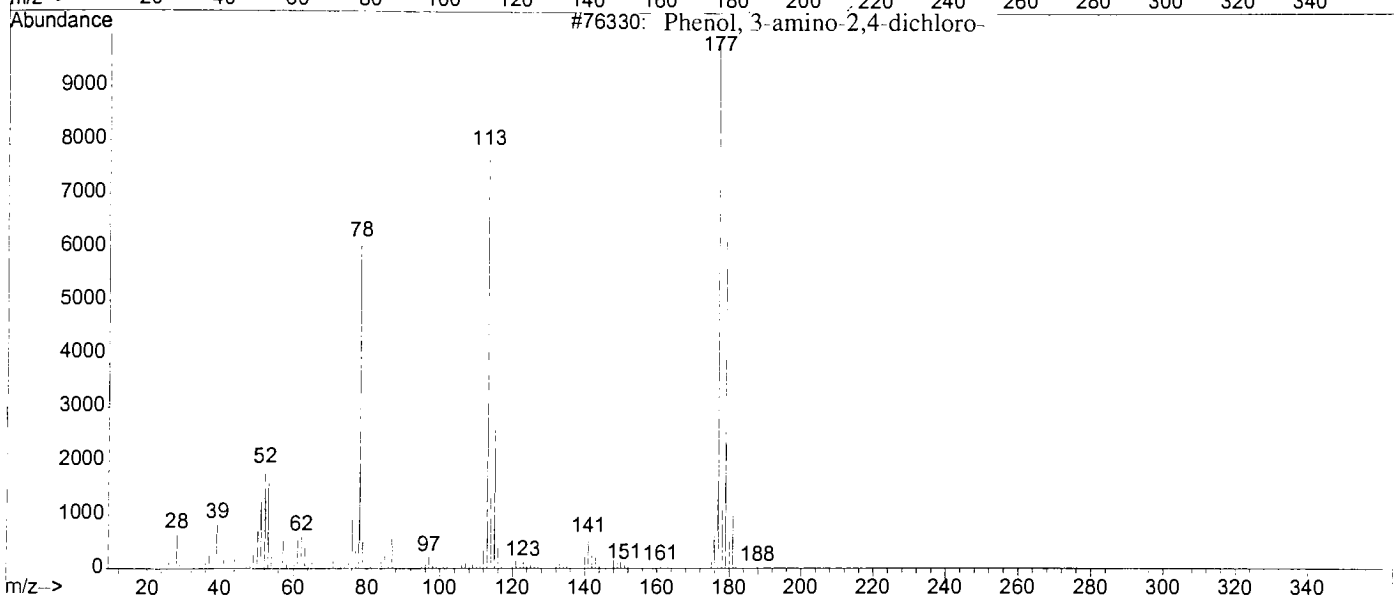
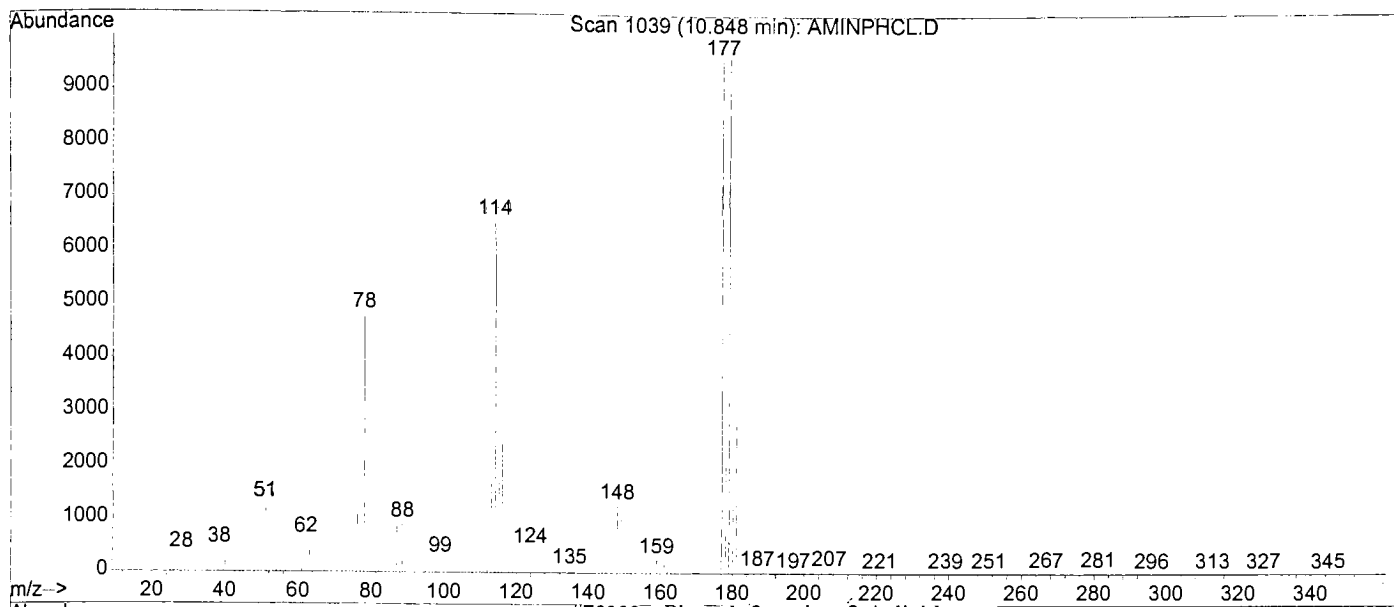
Library Searched : C:\DATABASE\NIST98.L

Quality : 42

ID

Phenol, 3-amino-2,4-dichloro- MS Spectrum 10.

B10

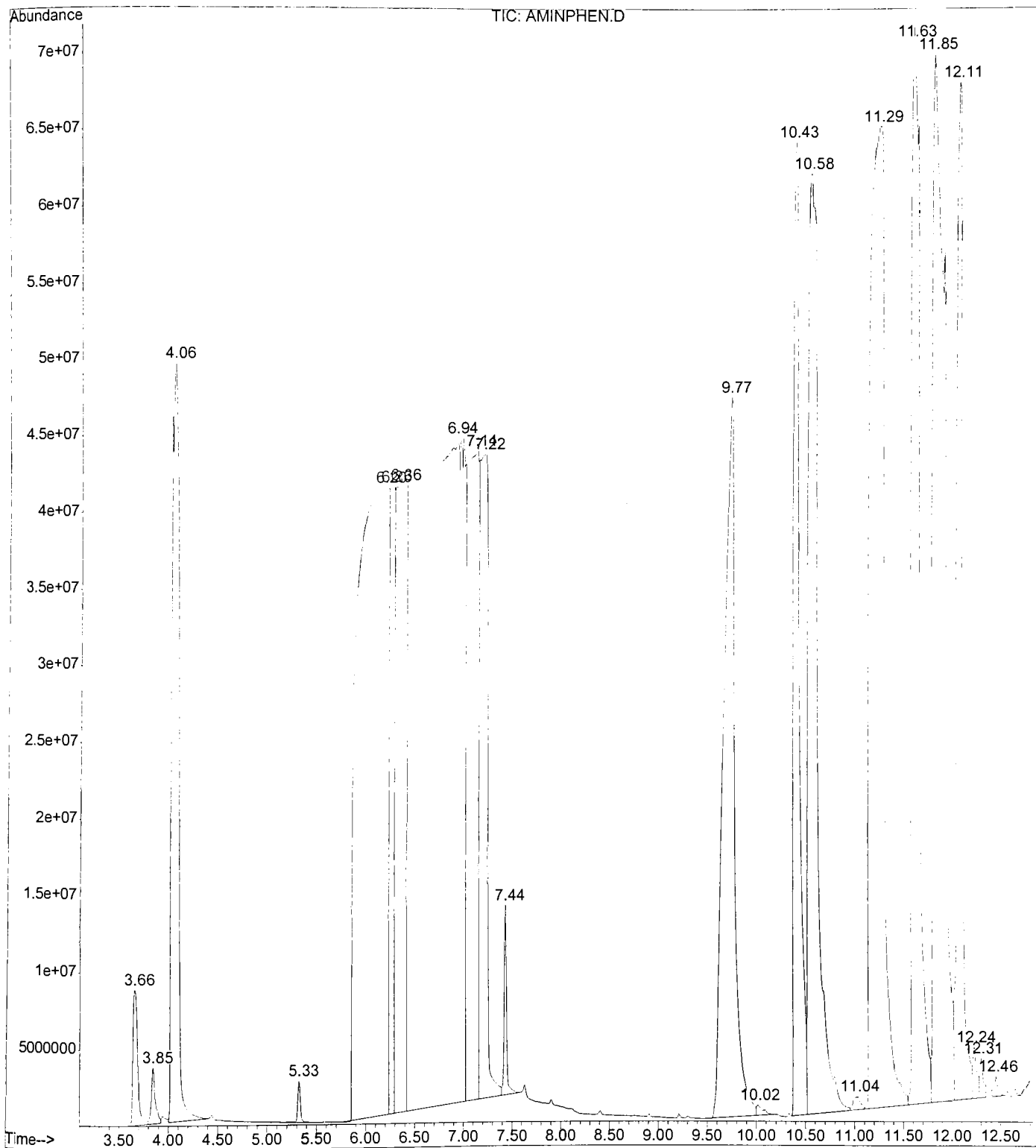


GC Spectrum 11. Area Percent Report

Data File : C:\HPCHEM\1\DATA\AMINPHEN.D
Acq On : 13 Jul 2000 19:43
Sample :
Misc :

Vial: 1 B11
Operator:
Inst : RIT Chemi
Multiplr: 1.00
Sample Amount: 0.00

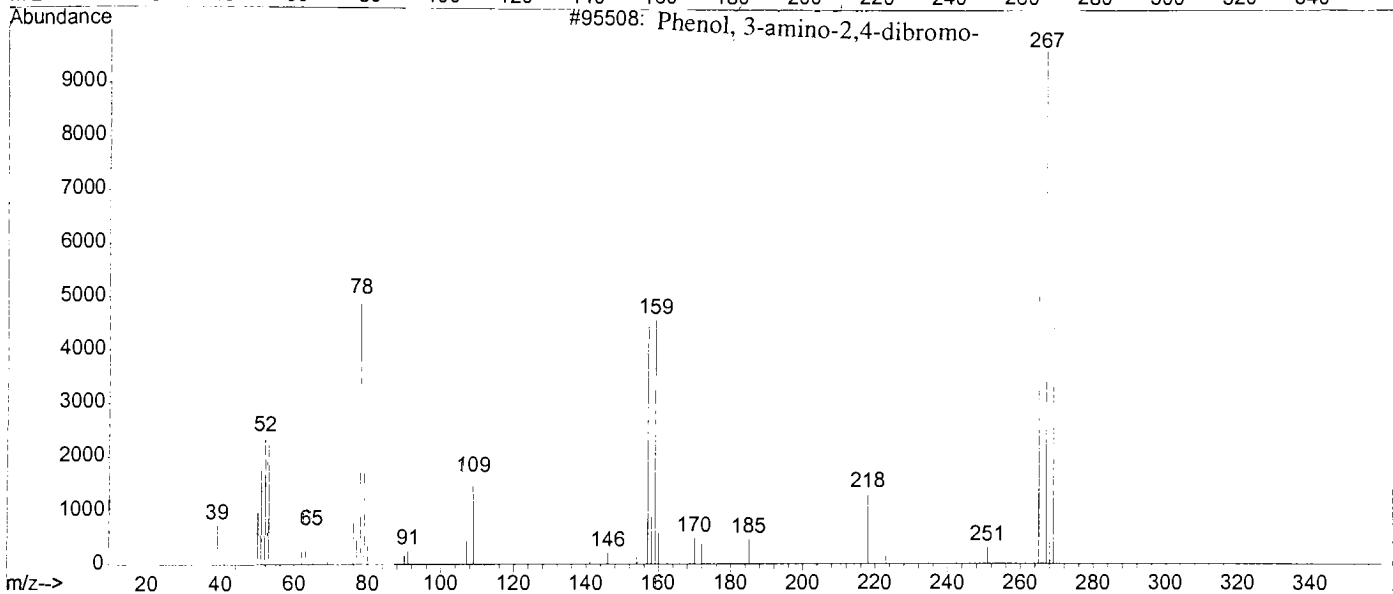
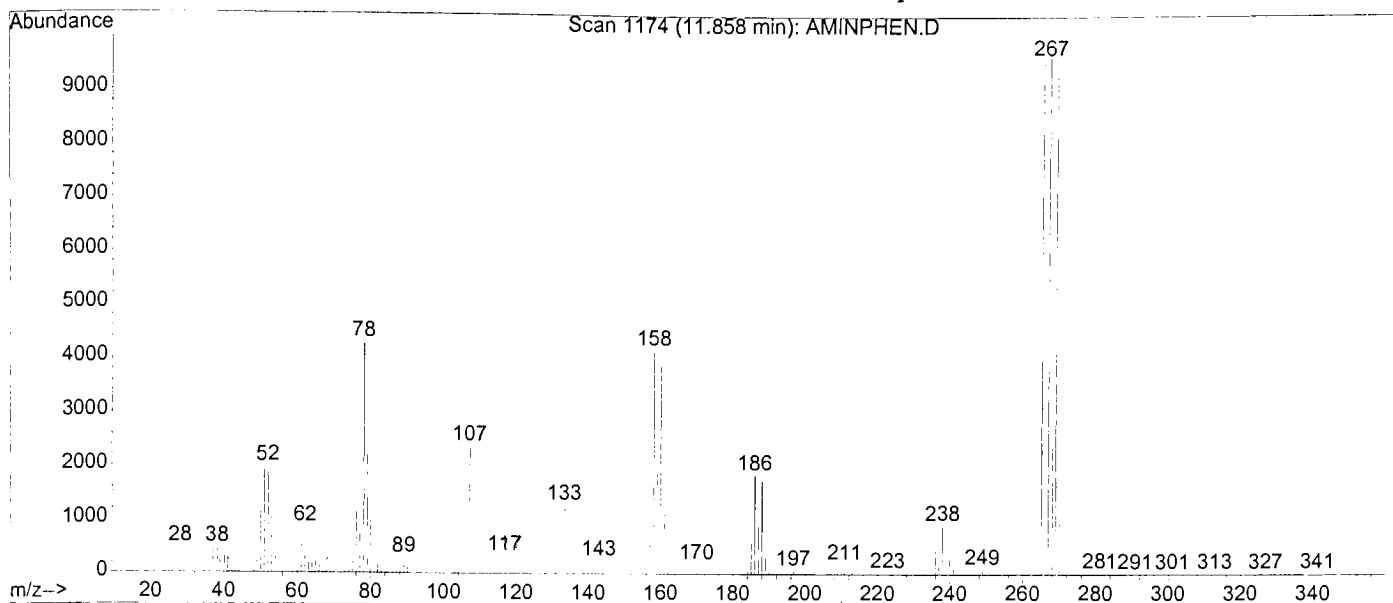
MS Integration Params: autoint1.e
Method : C:\HPCHEM\1\5973\PANKAJ.M (Chemstation Integrator)
Title :



Library Searched : C:\DATABASE\NIST98.L
Quality : 25
ID :

B11

Phenol, 3-amino-2,4-dibromo- MS Spectrum 11.

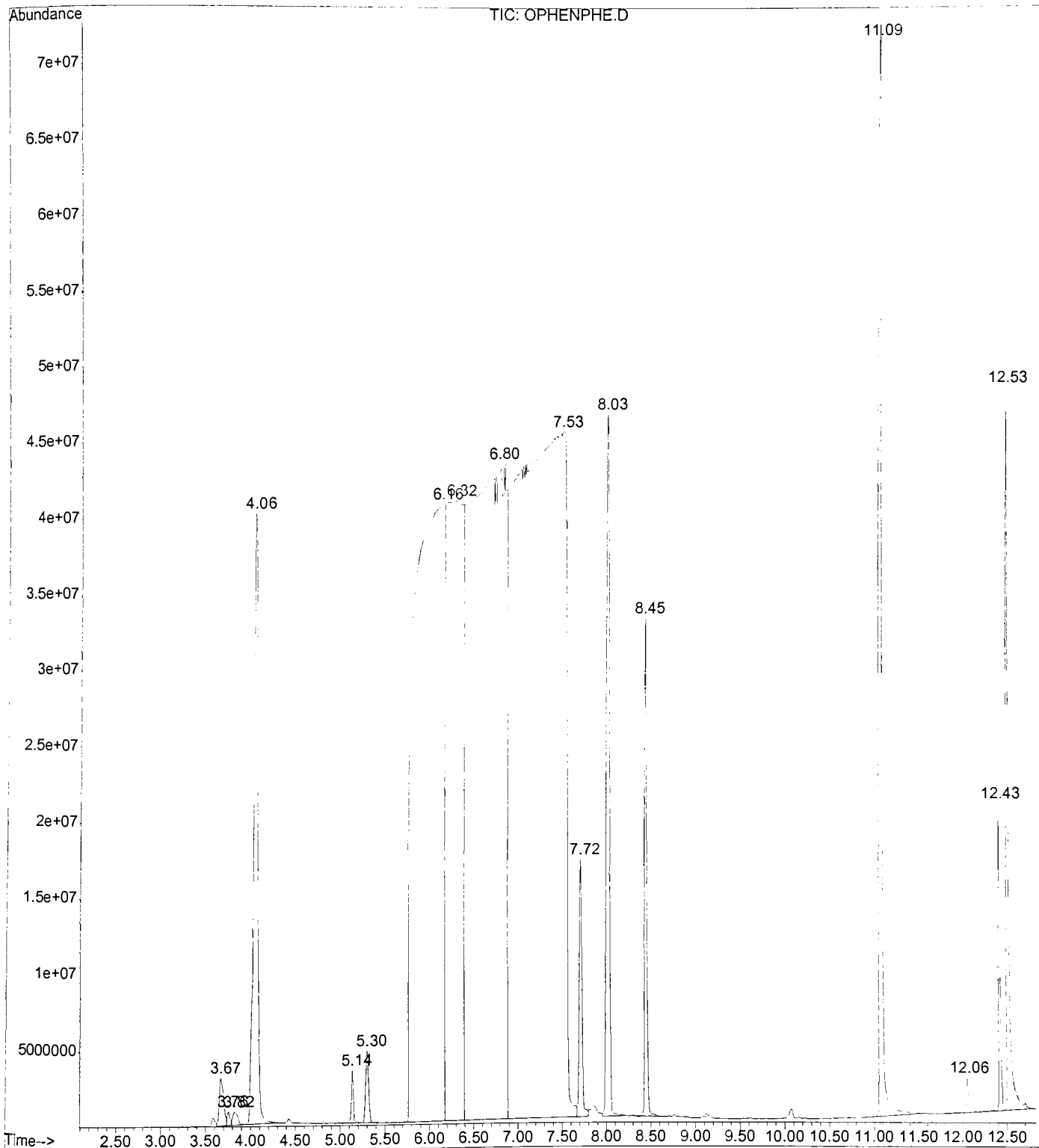


GC Spectrum 12. Area Percent Report

Data File : C:\HPCHEM\1\DATA\OPHENPHE.D
Acq On : 21 Jun 2000 16:16
Sample :
Misc :

Vial: 1 B12
Operator:
Inst : RIT Chemi
Multiplr: 1.00
Sample Amount: 0.00

MS Integration Params: autoint1.e
Method : C:\HPCHEM\1\5973\PANKAJ.M (Chemstation Integrator)
Title :

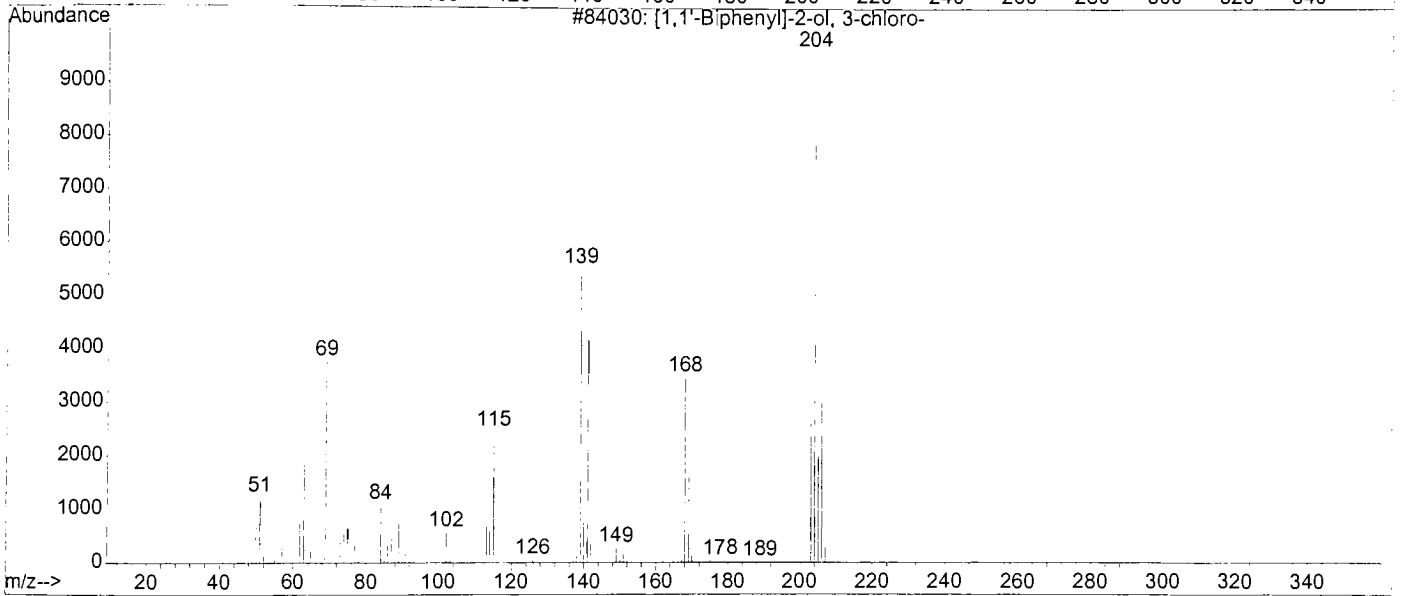
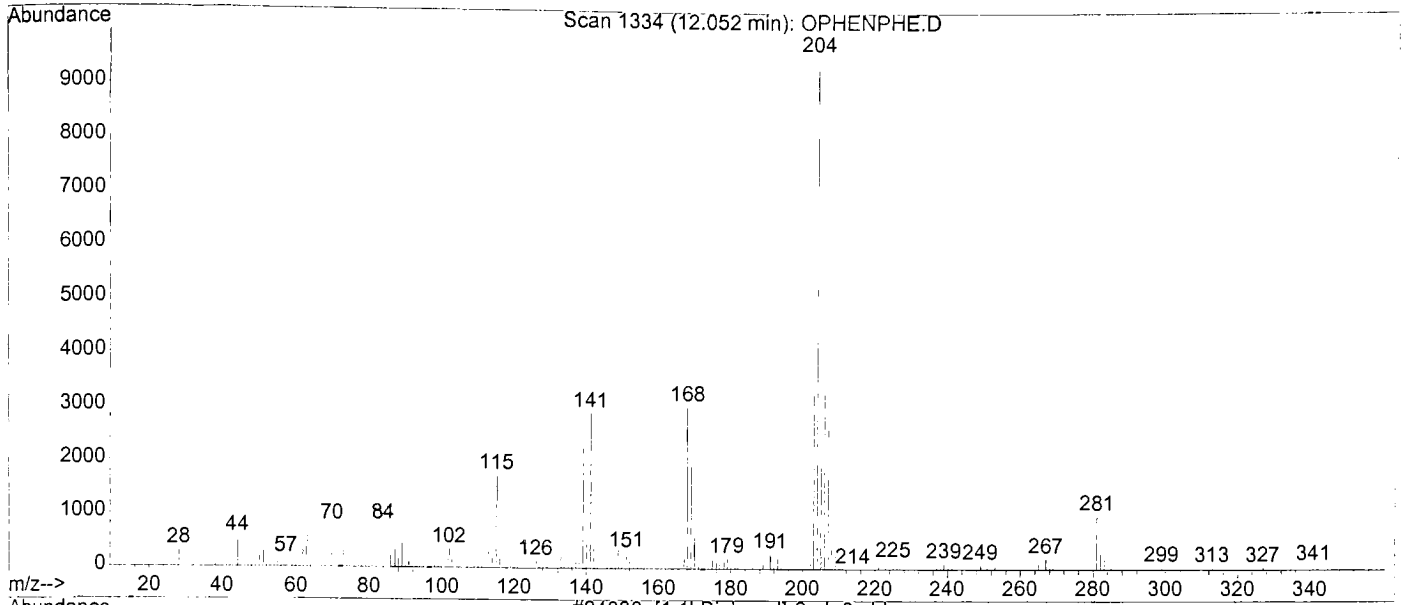


Library Searched : C:\DATABASE\NIST98.L

Quality : 90

ID : [1,1'-Biphenyl]-2-ol, 3-chloro-

MS Spectrum 12. B12

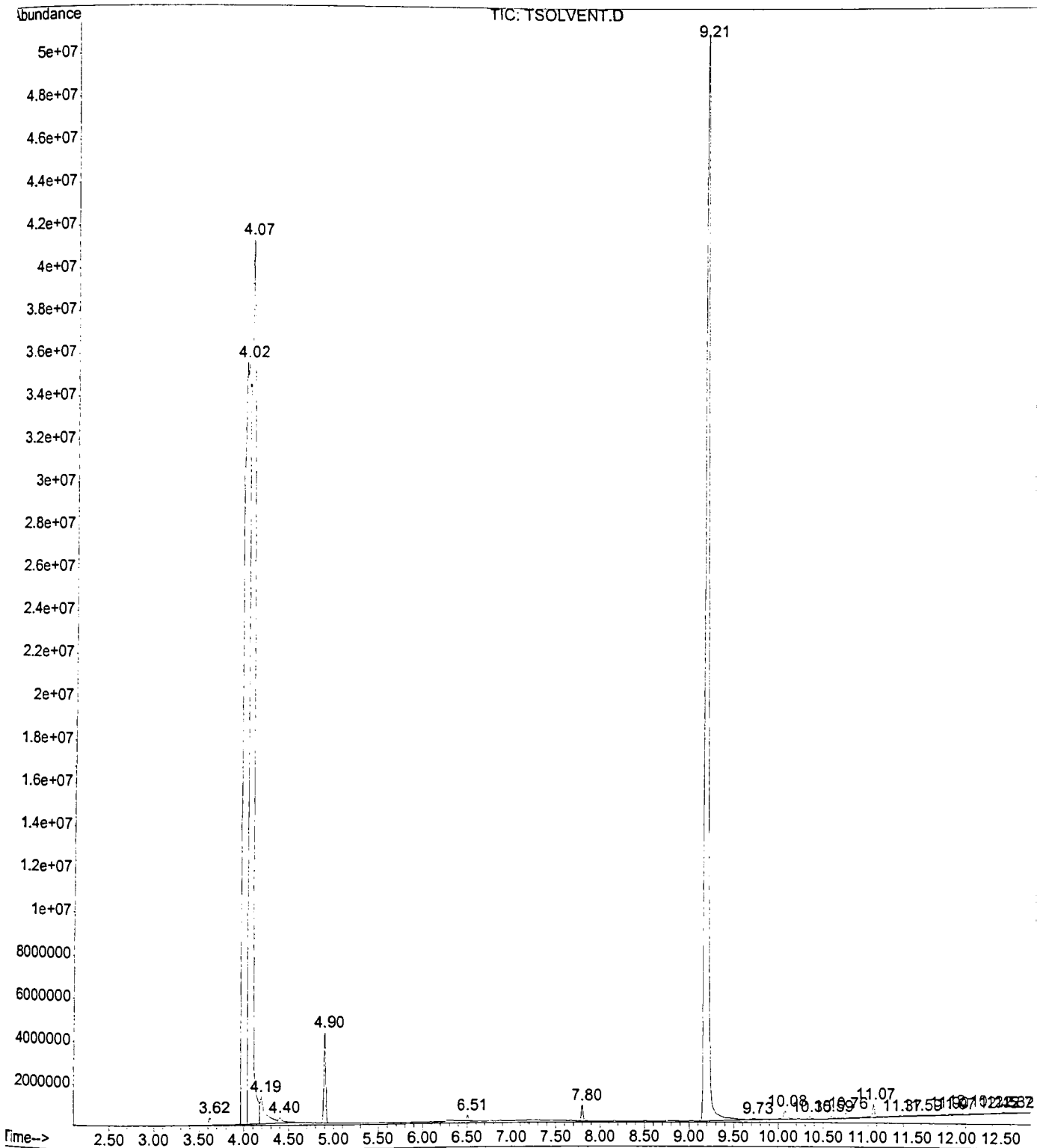


GC Spectrum 13. Area Percent Report

Data File : C:\HPCHEM\1\DATA\OPHENHBR.D
Acq On : 21 Jun 2000 16:57
Sample :
Misc :

Vial: 1 B13
Operator:
Inst : RIT Chemi
Multiplr: 1.00
Sample Amount: 0.00

MS Integration Params: autoint1.e
Method : C:\HPCHEM\1\5973\TRACY.M (Chemstation Integrator)
Title :

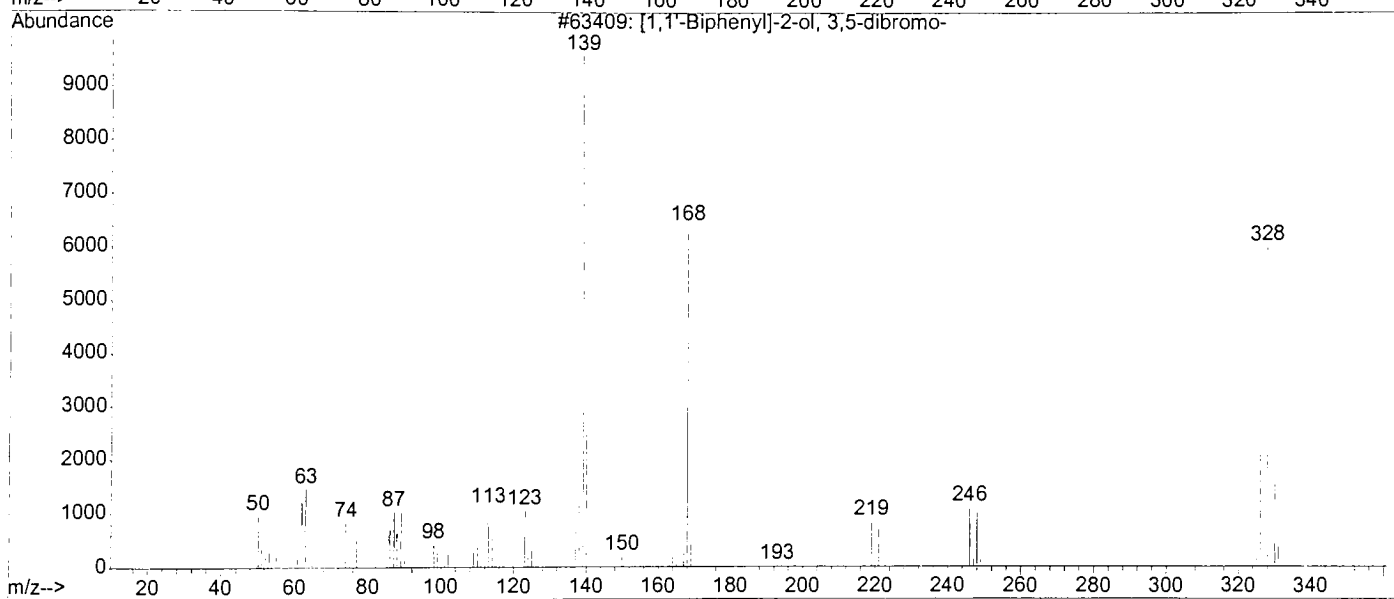
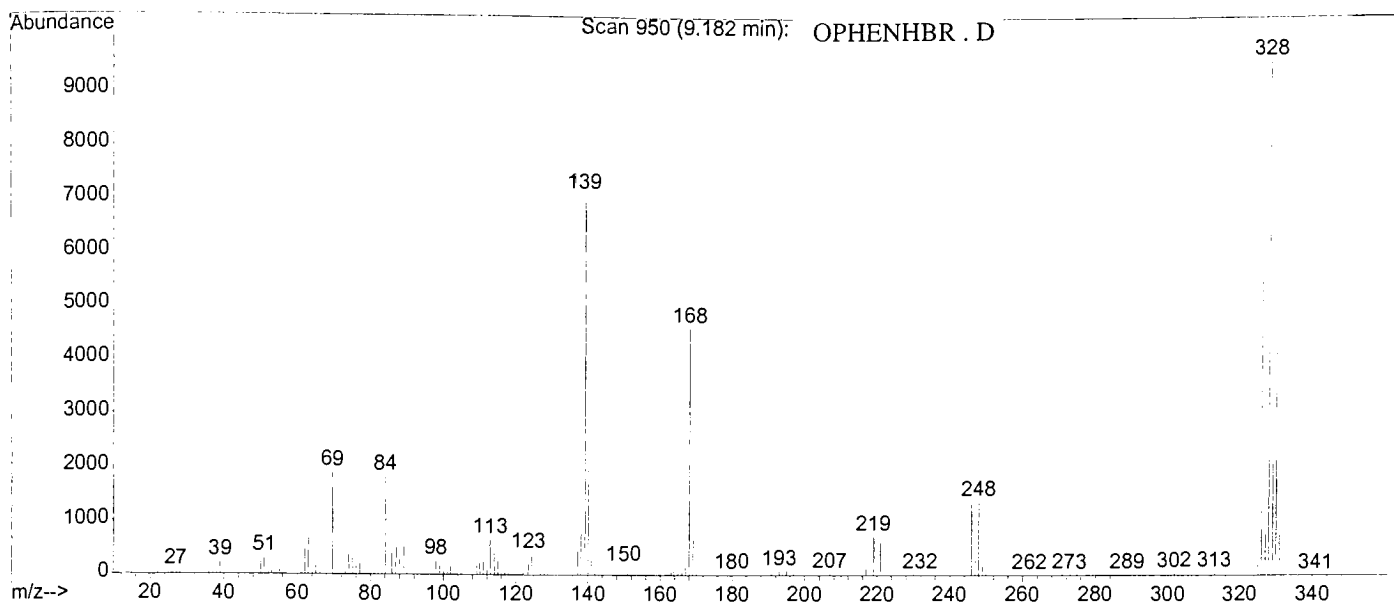


Library Searched : C:\DATABASE\NIST98.L

B13

Quality : 95

ID : [1,1'-Biphenyl]-2-ol, 3,5-dibromo- MS Spectrum 13.



APPENDIX C

Table of NMR Parameters

Current Data Parameters

NAME	Jun19-00
EXPNO	6
PROCNO	1

F2 - Acquisition Parameters

Date_	500000
Time	16.51
INSTRUM	spect
PROBHD	5 mm QNP 1H
PULPROG	zg30
TD	32768
SOLVENT	DMSO
NS	16
DS	2
SWH	6172.839 Hz
FIDRES	0.188380 Hz
AQ	2.6542580 sec
RG	28.5
DW	81.000 usec
DE	6.00 usec
TE	300.0 K
D1	1.00000000 sec
P1	9.00 usec
DE	6.00 usec
SFO1	300.1318534 MHz
NUC1	1H
PL1	-5.00 dB

F2 - Processing parameters

SI	16384
SF	300 1300000 MHz
WDW	EM
SSB	0
LB	0.30 Hz
GB	0
PC	1.00

1D NMR plot parameters

CX	20.00 cm
F1P	11.000 ppm
F1	3301.43 Hz
F2P	-1.000 ppm
F2	-300.13 Hz
PPMCM	0.60000 ppm/cm
HZCM	180.07800 Hz/cm

APPENDIX D

Data Analysis Parameters

Method Name: C:\HPCHEM\1\5973\CHRIS.M

Percent Report Settings

Sort By: Retention Time

Output Destination

Screen: Yes
Printer: Yes
File: No

Integration Events: AutoIntegrate

Generate Report During Run Method: Yes

Signal Correlation Window: 0.020

Qualitative Report Settings

Peak Location of Unknown: Apex

Library to Search	Minimum Quality
C:\DATABASE\NIST98.L	0

Integration Events: AutoIntegrate

Report Type: Summary

Output Destination

Screen: No
Printer: Yes
File: No

Generate Report During Run Method: Yes

Quantitative Report Settings

Report Type: Summary

Output Destination

Screen: Yes
Printer: No

Calibration Last Updated: Mon Nov 20 15:48:49 1995

Reference Window: 10.00 Percent
Non-Reference Window: 5.00 Percent
Correlation Window: 0.02 minutes
Default Multiplier: 1.00
Default Sample Concentration: 0.00

Compound Information

1) mass 284 ()

Ret. Time 7.28 min., Extract & Integrate from 6.78 to 7.78 min.

Signal	Rel Resp.	Pct. Unc.(rel)	Integration
Tgt 283.90			*** AUTO ***

Lvl ID	Conc ()	Response
3	1.000	371748
4	10.000	3849624
5	100.000	18762380
2	0.100	31150

Qualifier Peak Analysis ON
Curve Fit: Avg. RF-----
2) mass 283 ()

Ret. Time 7.28 min., Extract & Integrate from 6.78 to 7.78 min.

Signal	Rel Resp.	Pct. Unc.(rel)	Integration
Tgt 283.90			*** AUTO ***

Lvl ID	Conc ()	Response
3	1.000	371748
4	10.000	3849624
5	100.000	18762380
2	0.100	31150

Qualifier Peak Analysis ON
Curve Fit: Avg. RF-----
3) mass 247 ()

Ret. Time 7.28 min., Extract & Integrate from 6.78 to 7.78 min.

Signal	Rel Resp.	Pct. Unc.(rel)	Integration
Tgt 246.90			*** AUTO ***

1.000	44562
10.000	479738
100.000	2662997
0.100	-1

D1

Qualifier Peak Analysis ON
Curve Fit: Avg. RF

Mass 212

()

Retention Time 7.28 min., Extract & Integrate from 6.78 to 7.78 min.

Signal	Rel Resp.	Pct. Unc.(rel)	Integration
Peak 212.00			*** AUTO ***

Peak ID	Conc ()	Response
	1.000	25707
	10.000	264918
	100.000	1532237
	0.100	-1

Qualifier Peak Analysis ON
Curve Fit: Avg. RF

END OF DATA ANALYSIS PARAMETERS

Thu Jul 13 15:56:22 2000